

## EFFICACY OF FUNGICIDES, SODIUM HYPOCHLORITE AND NEEM SEED POWDER TO CONTROL SEED BORNE PATHOGENS OF MAIZE

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### Abstract

Using standard blotter and deep freezing techniques, 7 genera and 11 species of fungi viz., *Aspergillus niger*, *A. flavus*, *A. wentii*, *Chaetomium* sp., *Drechslera* sp., *Fusarium chlamydosporum*, *F. oxysporum*, *F. moniliforme*, *F. semitectum*, *F. nivale*, *Nigrospora* sp., *Phoma* sp. and *Rhizopus* sp. were isolated from maize seeds. Seed treatment with fungicides viz., Antracol (70% WP), Aliette (80% w/w), Ridomyl Gold (MZ 68% WP), Neem seed powder @ 0.1%, 0.2% & 0.3% and Sodium hypochlorite @ 10% were used. No adverse effects were observed on the germination of seeds in blotter method whereas the germination was reduced due to dead/frozen embryo in deep-freezing method. Ridomyl Gold was found to be effective against seed borne mycoflora of maize followed by Aliette, Neem seed powder, Antracol and Sodium hypochlorite.

### Introduction

In Pakistan maize is third important cereal crop after wheat and rice. It accounts for 4.8% of the total cropped area and 3.5% of the value of agricultural output. It is planted over an estimated area of 981.8 thousand hectares with an annual production of 2797.0 thousand tones, with an average yield of 2849 kg/ha (Anon, 2005). Maize is attacked by more than 60 diseases (Anon., 1980). *Fusarium moniliforme* produces gibberella ear rot, kernel rot, stalk rot, seedling blight, seed rot, wilt and stunt (Kocharek & Kommedhol, 1966; Ullstrup, 1978; Ochar *et al.*, 1987; Leon & Pandey, 1989; Thiel *et al.*, 1991). *Aspergillus flavus* becomes systemic and produces aflatoxin and virescence in seedling of maize and damage stored corn (Blat, 1969). *Sclerotinia sclerotiorum* produces sclerotium blight in maize (Ahmed *et al.*, 1987). Maize is also infected by downy mildew pathogen (Adenle & Cardwell, 2000; Ajala *et al.*, 2003; Ahmed *et al.*, 2006). *Fusarium* spp., invade more than 50% maize grain before harvest and produce mycotoxins (Bakan *et al.*, 2002). The rank of fungi is second after insects as the cause of deterioration and loss of maize (Ominski *et al.*, 1994). In Pakistan the most common seed borne fungi isolated from maize seeds are *Alternaria* spp., *Aspergillus* spp., *Curvularia* spp., *Helminthosporium maydis*, *Monilia* spp., *Penicillium* spp., *Rhizopus* spp. and *Trichoderma* spp. (Ghafoor & Khan, 1976). Seed borne fungi can be controlled by treatment with fungicides (Crosier & Patrick, 1946; Siddiqui & Zaman, 2004). Experiments were conducted to study the efficacy of some fungicides, Sodium hypochlorite and Neem seed powder against seed borne mycoflora of maize.

### Materials and Methods

Seeds of maize were tested by ISTA techniques using blotter and deep freezing methods for the detection of seed borne fungi. Sodium hypochlorite (10%) was used for

surface sterilization of seeds while fungicides viz., Antracol (70% WP), Aliette (80% w/w), Ridomyl Gold (MZ 68% WP) and Neem seed powder were used in addition to find their efficacy as a fungicide @ 0.1%, 0.2% & 0.3% in reducing seed borne mycoflora.

The fungicides and Neem seed powder were applied on seeds in conical flasks separately. The seeds treated with fungicides, Neem seed powder and Sodium hypochlorite (10%) were plated @ 10 seeds / plate on 3 layers of moistened blotter in 9 cm glass Petri plates, incubated at  $25\pm 1^\circ\text{C}$  in alternate cycle of 12 hours light and 12 hours darkness for 7 days. In deep-freezing method (Limonard, 1966) the treated and untreated seeds in Petri plates were incubated for one day at  $25\pm 1^\circ\text{C}$  and then in deep freezer at  $-4^\circ\text{C}$  for 24 hours. After deep-freezing the Petri plates were taken out and incubated for 7 days at  $25\pm 1^\circ\text{C}$ . In both methods the growth of fungi were observed after 7 days and isolated on Potato Dextrose Agar (PDA) slant. The fungi were identified up to species level after reference to Barnett & Hunter (1972), Booth (1972), Ellis (1970) & Nelson *et al.*, (1983).

### Results and Discussion

In blotter method the fungi isolated and identified were *Aspergillus niger*, *A. flavus*, *A. fumigatus*, *A. candidus*, *Rhizopus* sp., *Phoma* sp., *Cheatomium* sp., and *Nigrospora* sp., as compared to control. These results fully supported the results obtained by Orisi *et al.*, (2000). Sodium hypochlorite 10% completely controlled the growth of *Phoma* sp., and *Cheatomium* sp., (Table 1). The growth of *A. niger* was not reduced with Antracol and Neem seed powder whereas Aliette was effective @ 0.2% and 0.3%. Ridomyl Gold @ 0.3% controlled all fungi except *A. niger* which showed only 2% growth. The growth of *A. flavus* was not reduced by Sodium hypochlorite, Neem seed powder and fungicides (Antracol and Aliette), while 0.3% Ridomyl Gold completely controlled this fungus (Fig.1). For the control of seed borne mycoflora Ridomyl Gold was found to be the most effective followed by Neem seed powder, Aliette, Antracol and Sodium hypochlorite. The rest of the fungi viz., *A. fumigatus*, *A. candidus*, *Rhizopus* sp., *Phoma* sp., *Cheatomium* sp., and *Nigrospora* sp., showed a positive response of fungicides tested and Neem seed powder. Aziz (1988) reported that powder of neem provided protection to maize grain against fungi.

In deep freezing method, the fungi isolated were *A. flavus*, *A. niger*, *A. fumigatus*, *A. wentii*, *Drechslera* sp., *Fusarium nivale*, *F. oxysporum*, *F. chlamydosporum*, *F. moniliforme* and *F. semitectum*. The most dominant fungi were *Aspergillus* and *Fusarium* species as reported by Askun (2006) and Fandohan *et al.*, (2003) on the same seed. The result revealed that Aliette, Ridomyl Gold and Neem seed powder completely controlled the growth of *A. fumigatus*, *A. wentii*, *Drechslera* sp., *Fusarium nivale*, *F. oxysporum* and *F. moniliforme* (Table 2). Agbenin *et al.*, (2004), reported that *Fusarium* spp., was also controlled by using Neem seed powder, however in the present study, *A. flavus*, *A. niger*, *F. semitectum* were not controlled by Antracol, Aliette and Neem seed powder whereas Ridomyl Gold (0.3%) gave complete reduction in infection of all fungi except *F. semitectum* (1%) (Fig. 2). Only *Drechslera* sp., was fully controlled by Sodium hypochlorite (Table 2). Ridomyl Gold was found to be effective in all doses in both blotter and deep freezing method for the control of seed borne fungi, followed by Aliette, Neem seed powder, Antracol and Sodium hypochlorite. Deep-freezing method was also found best for isolation of *Fusarium* spp. These results are in close conformity with the findings of Khanzada *et al.*, (1988); Bilgrami & Ghaffar, (1993); Hussein *et al.*, (2002).



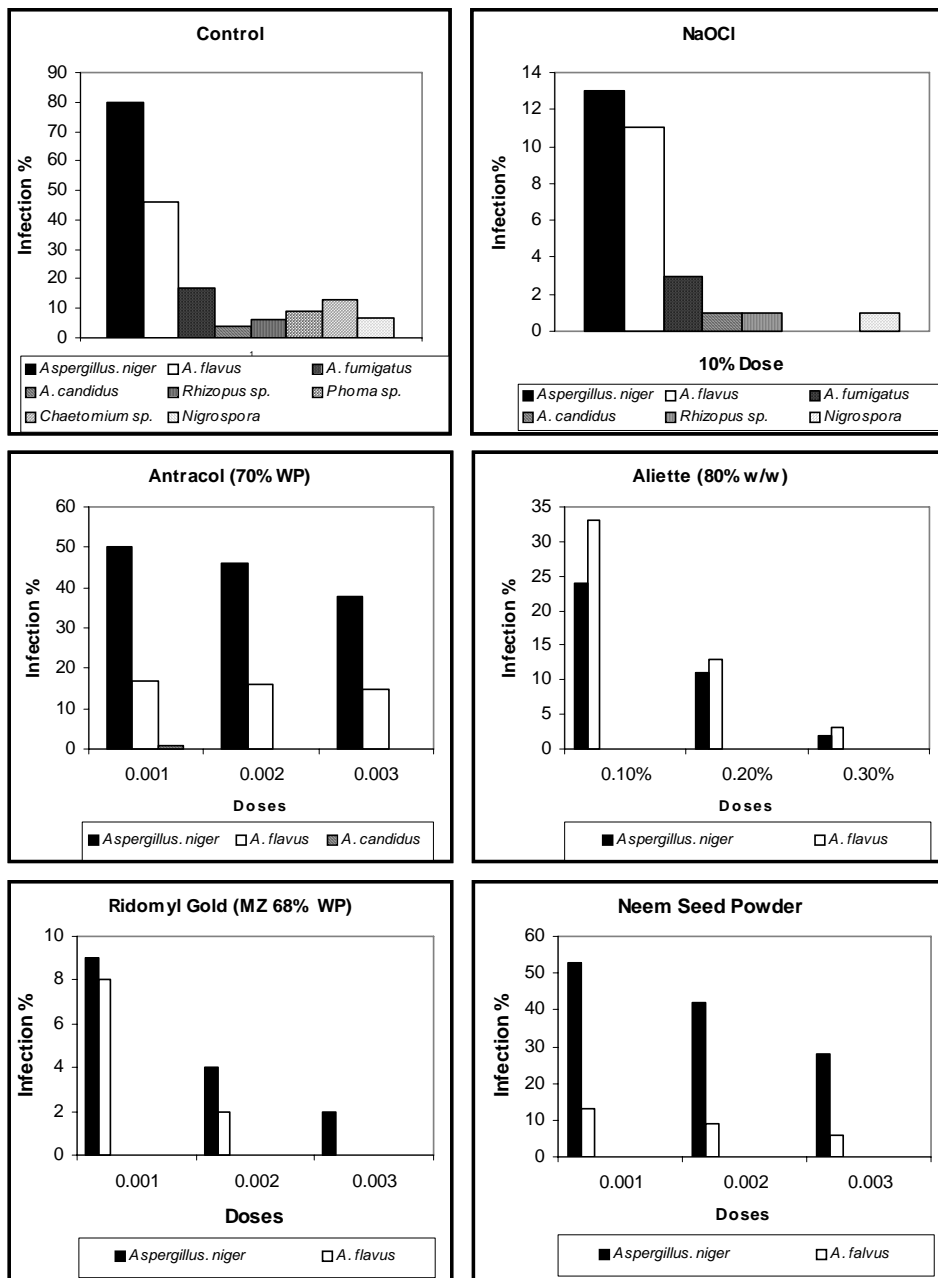


Fig. 1. Infection of seed borne fungi of Maize (Blotter method).

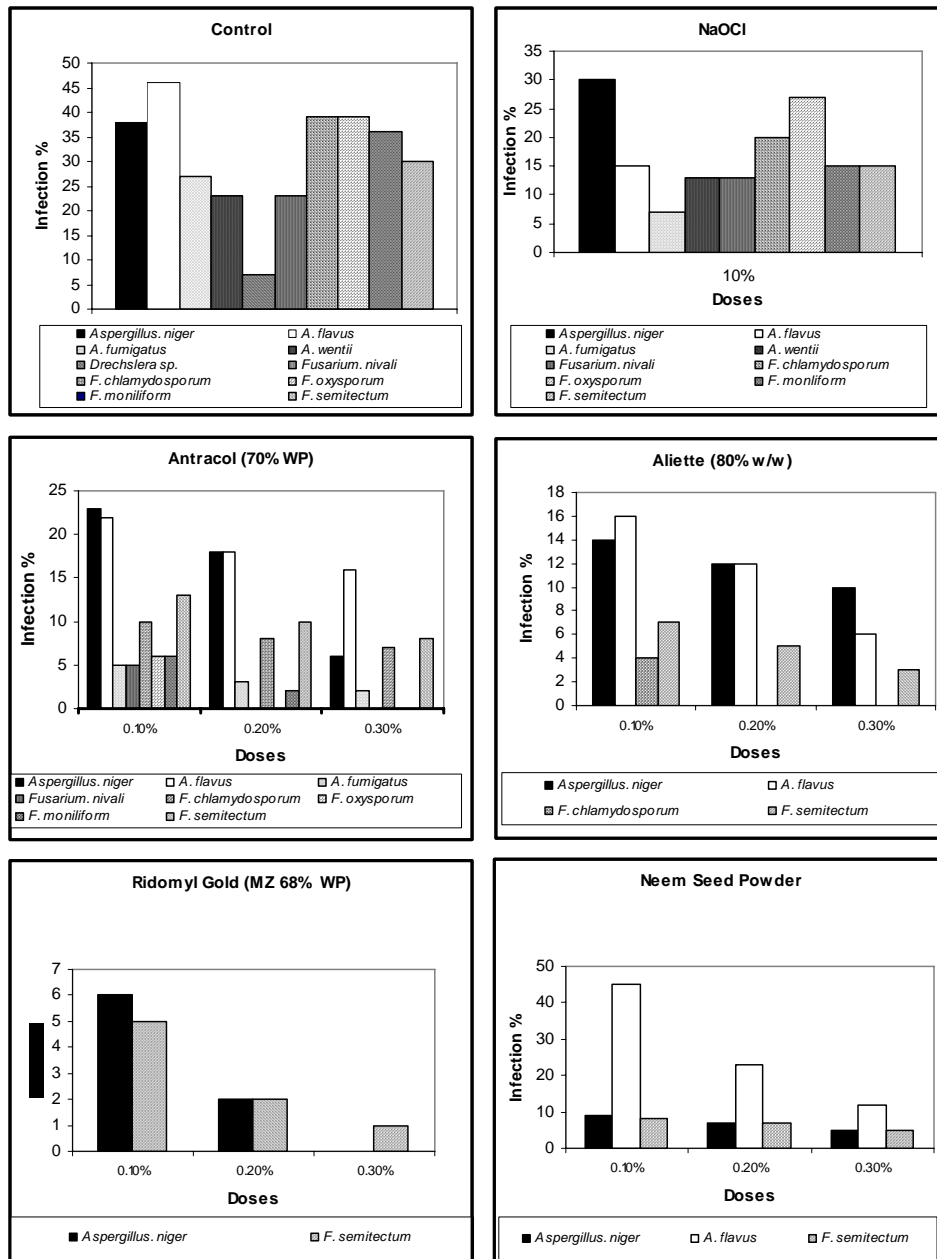


Fig. 2. Infection of seed borne fungi of Maize (Deep freezing method).

Table 3. Analysis of variance comparisons of fungicides with NaOCl\* (Blotter Method).

S. #	Doses	Treatments										F Value	P Value
		Antracol (70% WP)		Aliette (80% w/w)		Ridomyl gold (MZ 68% WP)		Neem seed powder		F Value	P Value		
		Mean	St. Dev	Mean	St. Dev	Mean	St. Dev	Mean	St. Dev				
1.	0.1%	8.50	17.78	7.13	13.41	2.12	3.94	8.25	18.65	1.51	0.221		
2.	0.2%	7.75	16.44	3.00	5.58	0.75	1.49	6.37	14.74	**2.41	0.068		
3.	0.3%	6.63	13.72	0.62	1.19	0.25	0.71	4.25	9.82	***3.41	0.019		

\*10% NaOCl was taken as standard in comparison with three concentration of the fungicides

Mean = 22.75

St. Dev = 26.77

\*\*Significant at 0.05

\*\*\*Highly significant at 0.05

Table 4. Analysis of variance comparisons of fungicides with NaOCl\* (Deep Freezing Method).

S. #	Doses	Treatments										F Value	P Value
		Antracol (70% WP)		Aliette (80% w/w)		Ridomyl gold (MZ 68% WP)		Neem seed powder		F Value	P Value		
		Mean	St. Dev	Mean	St. Dev	Mean	St. Dev	Mean	St. Dev				
1.	0.1%	9.000	8.124	4.100	6.226	1.100	2.331	7.100	13.932	**3.90	0.008		
2.	0.2%	5.900	7.279	2.900	5.043	0.400	0.843	4.400	7.321	***7.98	0.000		
3.	0.3%	3.900	5.343	1.900	3.479	0.100	0.316	2.200	4.022	***14.29	0.000		

\*10% NaOCl was taken as standard in comparison with three concentration of the fungicides

Mean = 15.500

St. Dev = 8.746

\*\*Significant at 0.05

\*\*\*Highly significant at 0.05

By using ANOVA, efficacies of fungicides, Neem seed powder showed highly significant result ( $\alpha=0.05$ ) as compared to Sodium hypochlorite (10%). In blotter method 0.2% and 0.3% doses are significant (Table 3) while in deep freezing method all doses (0.1%, 0.2% and 0.3%) are highly significant at 0.05% (Table 4). It was observed that in blotter method fungicides, Neem seed powder and Sodium hypochlorite has no adverse effects on germination of seeds specially Ridomyl Gold controlled the fungi and gave 100% seed germination while in deep freezing method it provided very low germination due to frozen or dead embryo.

The deep freezing method was best for the isolation of deep seated and pathogenic fungi viz., *Fusarium* spp. while blotter method was found suitable for germination test and for isolation of *Aspergillus* spp. Ridomyl Gold, Aliette and Neem seed powder were found to be most effective for the control of fungi associated with maize seeds.

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### References

- Adenle, V.O. and F. Cardwell. 2000. Seed transmission of maize downy mildew (*Peronospora sorghi*). *Nigeria-Plant Pathology*, 49(5): 628.
- Agbenin, N.O., A.M. Emechebe and P.S. Marley. 2004. Evaluation of Neem seed powder for *Fusarium* wilt and *Meloidogne* control on tomato. *Archives of Phytophology and Plant Protection*, 37(4): 319-326.
- Ahmed, S., D. Jeffers, S.K. Vasal, R. Frederiksen and C. Magill. 2006. A region of maize chromosome 2 effects response to downy mildew pathogens. *Theoretical and Applied Genetics*, 113(2): 321.
- Ahmed, Y., M.S. Mirza and M. Aslam. 1987. Sclerotinia blight of maize caused by *Sclerotinia sclerotiorum*. *Pak. J. Agric. Res.*, 8(4): 474-476.
- Ajala, S.O., J.G. Kiling, S.K. Kim and A.O. Obajimi. 2003. Improvement of maize populations for resistance to downy mildew. *Plant Breeding*, 122(4): 328.
- Anonymous. 1976. International rules of seed testing. *Proe. Int. Seed Test. Assoc.*, 4:3-49.
- Anonymous. 1980. *Foods and Agriculture Organization (FAO). Improvement and Production of Maize, Sorghum and Millet*. Vol. 2-Breeding, Agronomy and seed production. *FAO Pl. Prod and Prot paper* No. 24/2. R 72-77.
- Anonymous. 2005. *Agriculture Statistics of Pakistan, 2004-2005*. Ministry of Food and Agricultural and Livestock, Govt of Pakistan, Islamabad.
- Askun, T. 2006. Investigation of fungal species diversity of maize kernels. *Journal of Biological Sciences*, 6(2): 275-281.
- Aziz, P. 1988. Effect of neem leaves and barks powder on the storage of maize and gram. *Pak. J. Agric. Res.*, 9(4): 483-487.
- Bakan, B., D. Meleion, D.R. Molard and B. Cahagnier. 2002. Fungal growth and *Fusarium* mycotoxin contention. *Isogenic Traditional Maize and Genetically Modified Maize Grown in France and Spain*, 50(4): 728-731.
- Barnet, H.L. and B.B. Hunter. 1998. *Illustrated genera of imperfect fungi*. The American Phytopathological Society, St. Paul, Minnesota, 218 pp.
- Bilgrami, Z. and A. Ghaffar. 1993. Detection of seed borne mycoflora in *Pinus gerardiana*. *Pak. J. Bot.*, 25(2): 225-231.
- Blat, G. 1969. *Aflatoxin*. Academic Press, Inc (London), p. 17.

- Booth, C. 1971. *The Genus Fusarium*. Commonwealth Myco. Inst. Kew, Surrey, England, 237 pp.
- Crosier, W. and S. Patrick. 1946. Arasan for control of fungi in germination corn seed. *Phytopathol.*, 36, 162-164.
- Ellis, M. S. 1971. *Dematiaceous Hyphomycetes* (C.M.I., Kew, Surrey, England), 608 pp.
- Fandohan, P., K. Hell, W.F.O. Marasus and M.J. Wingfield. 2003. Infection of maize by *Fusarium* species and contamination with fumonisin in Africa. *African Journal of Biotechnology*, 2(12): 570-579.
- Ghafoor, A. and S.A.J. Khan. 1976. *List of diseases of economic plants in Pakistan*. Ministry of Food and Agriculture, Islamabad, Pakistan. 26 pp.
- Hussein, H.M., M.J. Christensen and M. Baxter. 2002. Occurrence and distribution of *Fusarium* species in maize fields in New Zealand. *Mycopathologia*, 156 (1): 25-30.
- Khanzada, A.K., N. Sultana, S.A.J. Khan and M. Aslam. 1988. Seed mycoflora of vegetable and its control. *Pak. J. Sci. Ind. Res.*, 31(8): 574-576.
- Kocharek, T. and T. Kommedhal. 1966. Kernel infection and corn stalk rot caused by *Fusarium moniliforme*. *Phytopathol.*, 56: 983-984.
- Leon, C. and S.S. Pandey. 1989. Importance of resistance to ear and stalk rots and agronomic traits in tropical maize gene pools. *Crop. Sci.*, 29: 12.
- Limonard, T. 1966. A modified blotter test for seed health. *Neth Pl. Path.* 72: 319-321.
- Nelson, P.E., T.A. Toussoun and W.F.O. Marasas. 1983. *Fusarium* species. *An Illustrated Manual for Identification*. The Pennsylvania State University Press, 193 pp.
- Ochar, T.E., L.E. Trevathan and S.B. King. 1987. Relationships of harvest date and host genotype to infection of maize kernels by *Fusarium moniliforme*. *Plant Dis.*, 71:311.
- Ominski, K.H., R.R. Marquardt, R.N. Sinha and D. Abramson. 1994. Ecological aspects of growth and mycotoxins production by storage fungi. In: *Mycotoxins in Grains. Compounds other than Aflatoxin*. (Eds.): J.D. Miller, H.L. Threnholm. Eagen Press, USA. p 287-305.
- Orisi, R.B., B. Correa, C.R. Possi, E.A. Schammass, J.R. Nogueira, S.M.C. Dias and M.A.B. Malozzi. 2000. Mycoflora and occurrence of fumonisins in freshly harvested and stored hybrid maize. *J. Stor. Prod. Res.*, 36: 75.
- Siddiqui, Z.S. and Arif-uz-Zaman. 2004. Effects of benlate systemic fungicide on seed germination, seedling growth, biomass and phenolic contents in two cultivars of *Zea mays* L. *Pak. J. Bot.*, 36(3): 577-582.
- Thiel, P.G., W.F.O. Marasas, E.W. Sydenham, G.S. Shgephard, W.C.A. Gelderblom and J.J. Hievwenhvis. 1991. Survey of fumonisin production by *Fusarium* spp. *Allp. Environ. Microbiol.*, 57:1089.
- Ullstrup, A.J. 1978. Stalk rot and root rots. In *Corn Diseases in the United State and their Control* ARS/USDA Agriculture Hand book No. 199, Washington DC. p. 8.

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