

STUDIES ON LIMNOLOGICAL CHARACTERISTICS AND PLANKTONIC DIVERSITY IN D.G. KHAN CANAL WATER AT D.G. KHAN (PAKISTAN)

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Abstract

A study was carried out to investigate the seasonal variations in limnological characteristics and planktonic diversity of D.G. Khan Canal water as affected with sewage at D.G. Khan, Pakistan. Water samples were collected on monthly basis and analyzed for estimation of water temperature, light penetration, turbidity, boiling point, surface tension, viscosity, density, specific gravity, pH, EC, dissolved O₂, Free CO₂, alkalinity, carbonates, bicarbonates, sodium, chlorides, acidity, hardness, total solid, total volatile solids, total dissolved solids and total volatile dissolved solids. An attempt was also made to assess the biological parameters including frequency of occurrence, relative abundance and diversity index of plankton life. Density and diversity of plankton was used as a measure of water quality. Phytoplankton were abundant as compared to Zooplankton. 39 Phytoplankton genera were recorded. Among these 08 of Cyanophyta, 12 of Chlorophyta, 11 of Chrysophyta, 4 of Euglenophyta, 2 of each Pyrrophyta and Cryptophyta. 14 genera of Zooplankton were observed including 9 of Protozoan, 4 of Rotifers and one genus of Cladoceran. Total number of organisms was 616, out of which 523 were Phytoplankton and 93 were Zooplankton. Diversity index of Phytoplankton ranged from 2.53 to 2.99 and diversity index of Zooplankton ranged from 1.08 to 1.68. It may be concluded that the quality of canal water is marginally fit as the diversity index of Phytoplankton and Zooplankton was less than three throughout the study period.

Introduction

Our world especially developing countries are facing the problem of water stress due to rapid growth in population. Food and Agriculture Organization has estimated the world's hunger population 923 million (Anon., 2008). The problem is expected to be further aggravated as there will be an additional 2 billion people by the year 2030 (Gany, 2006). The increased population will increase the demand of food accordingly; enhanced water diversions for irrigation need by 14 -17% (Bos *et al.*, 2005). Major causes of lack of food are water shortages due to limited water availability and inefficient use of available water (Laghari *et al.*, 2008).

Most of the cities and great civilizations have developed along the banks of fresh water sources especially canals and rivers for their multi-use. Their untreated sewage discharge not only damage the aquatic life but also hazardous to human health used for drinking and irrigation purpose in the downstream areas (Rather *et al.*, 2010). In Pakistan, only a small fraction of urban sewage is treated before discharge in fresh water bodies (Anon., 2007).

Plankton are very sensitive to the environment where they live; so, any change in the environment may lead to the change in the planktonic communities in terms of tolerance, abundance, diversity and dominance in the habitat. Therefore, planktonic population observation can be used as a reliable tool to assess the pollution status of water bodies (Basu *et al.*, 2010; Prabhakar *et al.*, 2011). Planktonic life is an essential part of aquatic ecosystem to maintain a healthy and productive environment (Khangarot & Das, 2009). The physico-chemical properties and nutrient status of aqueous medium have significant role in production of plankton which is critical to maintain aquatic food web foundations (Rahman & Hussain, 2008). Freshwater ecosystems have lost a greater

proportion of their species and habitat due to threats from dams, over extraction, pollution, and over fishing (Revenge & Mock, 2000). Biological characteristics are related to density and diversity of organisms (Barnabe, 1990). The high relative abundance of Chlorophyta indicates productive water (Boyd & Tucker, 1998).

D.G. Khan City is located on the western side of Indus River and its area is categorized as Barani in general, because the western side of the city receives hill torrents of Sulaiman Range. The groundwater is saline. The only potable water is from the seepage of Manka and D.G. Khan Canals. The annual design discharge of D.G. Khan Canal is 2.205 MAF and its useable seepage is 0.723 MAF. About 90% of the effluents and municipalities are untreated, which are directly polluting the both water resources and estimated pollution load is 60 Cusecs (Anon., 2006).

Keeping in view the importance of freshwater resources, an attempt has been made to study the limnological characteristics and planktonic diversity in D.G. Khan Canal in comparison to water quality standards.

Materials and Methods

The present study was carried out on mixed water of D.G. Khan Canal at D.G. Khan (*longitude* 70° 29' 7" E and *latitude* 29° 57' 38" N), Pakistan. The study site was suitable for limnological studies because the city sewage is added and properly mixed here, the depth and flow of water was also maximum. Water samples from the surface water column (≤ 1 m depth) were collected in plastic bottles of 1.5 L capacity on monthly basis for a period of ten months. At the time of sampling, the water temperatures were recorded by using alcoholic thermometer. Light penetration was recorded with the help of Secchi's disc. Boiling point was measured by

using mercury thermometer. pH and conductivity were measured by using pH meter (HI-8417) and conductivity meter (AGB-1001, Japan), respectively. Density, specific gravity, viscosity and surface tension were determined by the methods given by Nabi *et al.* (1998) while all other parameters including turbidity, dissolved O₂, Free CO₂, carbonates, bicarbonates, sodium, chlorides, alkalinity, acidity, total hardness, total solids (TS), total volatile solids (TVS), total dissolved solids (TDS) and total volatile dissolved solids (TVDS) were determined by the methods as described by Boyd & Tucker (1998).

The water samples for plankton study were preserved by using 4% formalin solution (Battish, 1992) and examined under a microscope using 10X ocular and 10X and 40X objectives. The identification of phytoplankton and zooplankton were done up to generic level with the help of following literature (Ward & Whipple, 1959; Tonapi, 1980; Battish, 1992). Frequency of occurrence and relative abundance of each genus of Phytoplankton and Zooplankton was calculated for each month. Diversity index of plankton was calculated by using formula as described by Boyd (1981). The data were

subjected to analysis of variance to find out statistically significant relationship among different limnological parameters by using MSTATC program (version 2.10).

Results and Discussion

In the present investigation, all the mean data of physico-chemical parameters obtained from the monthly analysis of water samples are summarized in Table 1. During the study period, water temperature ranged from a minimum of 18.3°C (November) to a maximum of 35°C (July). Water temperature was found to increase from March to July and decrease from August to December. Temperature has profound influence, and direct or indirect effect on biodiversity of an ecosystem. Temperature showed an inverse relationship with dissolved oxygen. Basu *et al.*, (2010) made similar observation. Light penetration was maximum (38.1 cm) in June and minimum (21.6 cm) in November. There was highly significant ($p < 0.001$) positive correlation between temperature and light penetration (Table 4).

Table 1. Limnological characteristics of D.G. Khan Canal water as affected with domestic sewage.

| Parameters | Months | | | | | | | | | |
|---|--------|-------|-------|-------|-------|-------|-------|-------|-------|-------|
| | Mar | Apr | May | Jun | Jul | Aug | Sep | Oct | Nov | Dec |
| Water temperature (°C) | 22.4 | 31.5 | 33.3 | 30.1 | 35.0 | 30.5 | 28.7 | 26.5 | 18.3 | 18.5 |
| Light penetration (cm) | 25.9 | 27.7 | 32.5 | 38.1 | 36.0 | 31.2 | 30.2 | 23.4 | 21.6 | 24.1 |
| Boiling point (°C) | 97.3 | 96.0 | 98.2 | 99.1 | 98.2 | 97.5 | 97.4 | 96.7 | 98.3 | 96.2 |
| Density (g L ⁻¹) | 0.994 | 0.990 | 0.989 | 0.988 | 0.995 | 0.990 | 0.995 | 0.997 | 0.995 | 0.993 |
| Specific gravity | 0.997 | 0.993 | 0.993 | 0.991 | 0.998 | 0.993 | 0.998 | 1.001 | 0.998 | 0.995 |
| Turbidity (mg L ⁻¹) | 0.64 | 0.76 | 0.66 | 0.80 | 0.40 | 0.61 | 0.46 | 0.33 | 0.26 | 0.27 |
| Viscosity (mN S m ⁻²) | 0.944 | 0.923 | 0.899 | 0.909 | 0.961 | 0.995 | 0.807 | 0.887 | 0.918 | 0.975 |
| Surface tension (dynes cm ⁻¹) | 77.05 | 78.12 | 76.45 | 76.61 | 73.55 | 80.16 | 76.00 | 74.37 | 75.69 | 74.47 |
| pH | 7.3 | 7.6 | 8.1 | 8.0 | 7.7 | 7.9 | 7.3 | 7.1 | 8.3 | 7.5 |
| Conductivity (dS m ⁻¹) | 0.37 | 0.29 | 0.25 | 0.23 | 0.24 | 0.27 | 0.32 | 0.29 | 0.31 | 0.42 |
| Dissolved O ₂ (mg L ⁻¹) | 6.7 | 6.5 | 5.7 | 4.5 | 5.2 | 6.1 | 6.9 | 7.7 | 6.9 | 6.8 |
| Free CO ₂ (mg L ⁻¹) | 9.97 | 9.22 | 12.5 | 12.3 | 10.5 | 9.73 | 7.80 | 8.57 | 10.9 | 9.56 |
| Acidity (mg L ⁻¹) | 32 | 51 | 92 | 83 | 97 | 53 | 42 | 61 | 85 | 101 |
| Hardness (mg L ⁻¹ as CaCO ₃) | 320 | 211 | 200 | 190 | 200 | 203 | 250 | 220 | 260 | 215 |
| Alkalinity (mg L ⁻¹) | 105 | 132 | 177 | 165 | 146 | 108 | 70 | 121 | 174 | 161 |
| TS (mg L ⁻¹) | 0.61 | 0.35 | 0.21 | 0.57 | 0.58 | 0.44 | 0.48 | 0.35 | 0.32 | 0.20 |
| TVS (mg L ⁻¹) | 0.08 | 0.05 | 0.02 | 0.13 | 0.13 | 0.07 | 0.12 | 0.07 | 0.05 | 0.02 |
| TDS (mg L ⁻¹) | 0.57 | 0.24 | 0.25 | 0.23 | 0.26 | 0.22 | 0.32 | 0.32 | 0.25 | 0.24 |
| TVDS (mg L ⁻¹) | 0.06 | 0.01 | 0.06 | 0.05 | 0.05 | 0.01 | 0.03 | 0.06 | 0.05 | 0.04 |
| SAR | 5.85 | 6.71 | 7.10 | 7.42 | 3.85 | 4.21 | 7.36 | 2.96 | 3.44 | 1.36 |
| RSC (meq L ⁻¹) | Nil | Nil | 0.2 | Nil | Nil | 2.3 | Nil | Nil | Nil | Nil |

TS: Total Solids, TVS: Total Volatile Solids, TDS: Total Dissolved Solids, TVDS: Total Volatile Dissolved Solids, SAR: Sodium Adsorption Ratio, RSC: Residual Sodium Carbonate

High transparency of freshwater ecosystem coincided with the period of dry season when there is little or no rainfall (Achionye-Nzeh & Isimaikaiye, 2010). The boiling point was maximum (99.1°C) in June and minimum (96°C) in April. The maximum water density (0.997 g L⁻¹) and specific gravity (1.001) was observed in October and minimum (0.988 g L⁻¹) and 0.991 in June, respectively. The turbidity was maximum (0.80 mg L⁻¹) in June and minimum (0.26 mg L⁻¹) in November. Turbidity reduces the light penetration and affects the photosynthesis of phytoplankton which ultimately produces less oxygen (Iqbal *et al.*, 2010). The viscosity

(0.995 mN S m⁻²) was observed maximum in August and minimum (0.809 mN S m⁻²) in September. Viscosity showed significant inverse correlation with temperature and photoperiod while positive correlation with density and turbidity.

The Surface tension ranged from 77.39 to 101.1 dynes cm⁻¹ (Table 1). Surface tension ranged from a minimum of 73.55 dynes cm⁻¹ (July) to a maximum of 80.16 dynes cm⁻¹ (August). The D.G. Khan Canal water showed alkaline condition throughout the study period. The pH value was highest in May (8.1) and lowest in October (7.1). pH showed significant ($p < 0.005$) positive

correlation with alkalinity (Table 4). pH value of the water changes from basic nature to the acidic due to regular discharge of domestic sewage (Rather *et al.*, 2010). The maximum EC (0.42 dS m^{-1}) was observed in December and minimum (0.23 dS m^{-1}) in June. EC is imperative to find good quality of irrigation water as its high values cause salinization (Ghafoor *et al.*, 1993). EC showed highly significant ($p < 0.001$) inverse correlation with water temperature and significant ($p < 0.005$) positive correlation with dissolved O_2 (Table 4). The changes in electrical conductivity are due to fluctuations in dissolved solids (Boyd, 1981). The dissolved O_2 of water varied between 4.5 mg L^{-1} (June) to 7.7 mg L^{-1} (October). High dissolved O_2 is an indication of healthy aquatic ecosystem (Chattopadhyay & Banerjee, 2007). The dissolved O_2 evaluated the degree of freshness of an aquatic ecosystem (Agbaire & Obi, 2009).

Free CO_2 was observed highest (12.5 mg L^{-1}) in May, decreased steadily up to September (7.80 mg L^{-1}) and then increased during rest of period. Free CO_2 showed highly significant ($p < 0.001$) positive correlation with pH and alkalinity, while significant ($p < 0.005$) inverse correlation with dissolved O_2 and hardness (Table 4). The acidity of water ranged from 32 mg L^{-1} (March) to 101 mg L^{-1} (December). The hardness of water fluctuated from 190 mg L^{-1} (June) to 320 mg L^{-1} (March). Total hardness more than 75 mg L^{-1} is undesirable for fish production (Abbasi, 1998). Alkalinity was observed maximum (177 mg L^{-1}) in May and minimum (70 mg L^{-1}) in September. Brown (1993) reported that total hardness acts as limiting factor for alkalinity. Calcareous water with alkalinity more than 50 mg L^{-1} is most productive, zero to 20 mg L^{-1} for low production, 20 to 40 mg L^{-1} for medium production and 40 to 90 mg L^{-1} for higher production. The maximum value (0.61 mg L^{-1}) of TS was observed in March and minimum (0.20 mg L^{-1}) in December. The maximum value of TDS (0.57 mg L^{-1}) and TVDS (0.06 mg L^{-1}) was observed in March and minimum (0.22 mg L^{-1}) and (0.01 mg L^{-1}) in August, respectively. The SAR ranged from 1.36 (December) to 7.42 (June). Irrigation water with high SAR precipitates soil solution calcium and increase solution sodium, resulting in soil dispersion (Pervaiz, 2005). RSC was observed only in May (0.2 meq L^{-1}) and June (2.3 meq L^{-1}).

Phytoplankton were most abundant as compared to Zooplankton during the whole study period. Total number of organisms was observed 616, out of which 523 were Phytoplankton with relative abundance (R.A) 84.9% and 93 were Zooplankton with R.A 15.1%. Total 53 genera were observed in which 39 were of Phytoplankton and 14 of Zooplankton. Phytoplankton belong to Cyanophyta (8 genera), Chlorophyta (12 genera), Chrysophyta (11 genera), Euglenophyta (4 genera), Pyrrophyta (2 genera) and Cryptophyta (2 genera) while Zooplankton including Protozoan (9 genera), Rotifers (4 genera) and Cladoceran (one genus). Among the Phytoplankton, the members of Cyanophyta, Chlorophyta and Chrysophyta were present throughout the study period. The members of Euglenophyta were present in all months except April. Minimum frequency of occurrence was found in

Pyrrophyta and Cryptophyta. Among the Zooplankton, Protozoan and Rotifers were present in all months while Cladocerans were present in six months (Tables 2 & 3).

In March, *Chlorella* was most abundant genus among Phytoplankton with R.A 17.3% while in Zooplankton, *Diffugia* was most abundant genus with R.A 3.50%. In April, *Cymbella* was most abundant genus among Phytoplankton with R.A 10.6% while in Zooplankton, *Epiphanes* was most abundant genus with R.A 7.57%. In May, *Natrium* and *Treubaria* were most abundant genera among Phytoplankton with R.A 11.8% while in Zooplankton, *Tintinnopsis* was most abundant genus with R.A 7.35%. In June, *Navicula* was most abundant genus among Phytoplankton with R.A 12.3% while in Zooplankton, *Hemiphrys* was most abundant genus with R.A 5.10%. In July, *Chlorella* was most abundant genus among Phytoplankton with R.A 11.1% while in Zooplankton, *Pseudodiffugia* was most abundant genus with R.A 8.45%. In August, *Navicula* and *Tetrastrum* were most abundant genera among Phytoplankton with R.A 9.87% while in Zooplankton, *Daphnia* was most abundant genus with R.A 6.17%. In September, *Closterium* was most abundant genus among Phytoplankton with R.A 11.5% while in Zooplankton, *Paramecium* was most abundant genus with R.A 7.13%. In October, *Melosira* was most abundant genus among Phytoplankton with R.A 12.1% while in Zooplankton, *Colurella* was most abundant genus with R.A 10.3%. In November, *Chodatella* was most abundant genus among Phytoplankton with R.A 11.3% while in Zooplankton, *Centropyxis* was most abundant genus with R.A 5.27%. In December, *Tetrastrum* and *Tabellaria* were most abundant genera among Phytoplankton with R.A 12.3% while in Zooplankton, *Pseudodiffugia* was most abundant genus with R.A 5.26% (Tables 2 & 3).

Meshram (2003) stated that macrophytes stimulate the growth of Phytoplankton which helps in the recycling of organic matter; this can be positively correlated with high Phytoplankton density. Biodiversity is fluctuated with different factors like water level, temperature and nutrient level. Changes in aquatic environment due to pollution are a cause of growing concern, and require monitoring of the surface water and organisms inhabiting them (Vandysh, 2004).

Schabetsberger *et al.*, (2004) concluded that in a freshwater ecosystem, Phytoplankton were dominated by green algae while the Zooplankton were cladocerans. Similarly in present study, the maximum numbers of genera were observed of Chlorophyta in accordance with Ali *et al.*, (2010) while studying the algal flora in fresh waters of Swat valley in Pakistan.

Diversity index of Phytoplankton was found to be highest in September (2.99) and lowest in March (2.53), showed an increasing trend up to September and then decreased in rest of months. Diversity index of Zooplankton was observed maximum (1.68) in December and minimum (1.08) in July and October (Table 5). There was highly significant ($p < 0.001$) inverse correlation between relative abundance of phytoplankton and Zooplankton (Table 4). Pathania *et al.*, (2010) reported that the diversity index of Phytoplankton and Zooplankton of freshwater pond ranged 1.92 to 2.37 and 0.64 to 0.83, respectively.

Table 2. Relative abundance (%) of Phytoplankton in D.G. Khan Canal water affected with domestic sewage.

| Phytoplankton | Mar | Apr | May | Jun | Jul | Aug | Sep | Oct | Nov | Dec |
|-------------------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|
| Cyanophyta | 1.92 | 6.06 | 11.1 | 16.9 | 7.92 | 7.40 | 6.55 | 1.72 | 14.0 | 5.26 |
| <i>Anabaenopsis</i> | - | 6.06 | - | - | - | - | - | - | - | - |
| <i>Lyngbya</i> | - | - | 7.35 | - | - | 6.17 | - | - | - | - |
| <i>Coelosphaerium</i> | - | - | 3.75 | - | - | - | - | - | - | - |
| <i>Gloeocapsa</i> | - | - | - | 7.69 | - | - | - | - | 5.26 | - |
| <i>Dactylococcopsis</i> | - | - | - | 9.23 | - | - | - | 1.72 | - | - |
| <i>Anabaena</i> | 1.92 | - | - | - | 6.34 | - | - | - | - | - |
| <i>Aphanizomenon</i> | - | - | - | - | 1.58 | - | - | - | - | 5.26 |
| <i>Microcystis</i> | - | - | - | - | - | 1.23 | 6.55 | - | 8.77 | - |
| Chlorophyta | 44.2 | 25.7 | 35.4 | 24.6 | 31.7 | 29.6 | 36.1 | 29.3 | 40.3 | 42.1 |
| <i>Closterium</i> | 11.5 | - | - | 1.53 | - | 8.64 | 11.5 | - | - | - |
| <i>Chlorella</i> | 17.3 | - | - | - | 11.1 | 4.93 | - | - | - | 8.77 |
| <i>Oocystis</i> | 5.76 | 7.57 | 10.3 | 4.61 | - | 6.17 | - | - | - | - |
| <i>Gonatozygon</i> | - | 7.57 | - | - | - | - | - | 1.72 | - | 7.01 |
| <i>Netrium</i> | - | 9.09 | 11.8 | - | - | - | - | 12.1 | 8.06 | - |
| <i>Cosmarium</i> | - | - | 1.47 | - | 7.93 | - | - | - | 4.83 | - |
| <i>Treubaria</i> | - | - | 11.8 | - | - | - | 6.55 | - | 6.45 | - |
| <i>Asterococcus</i> | 9.61 | - | - | 7.69 | - | - | - | - | - | 10.5 |
| <i>Chodatella</i> | - | - | - | - | 6.34 | - | - | 5.17 | 11.3 | - |
| <i>Tetrastrum</i> | - | - | - | 10.8 | - | 9.87 | 1.63 | - | - | 12.3 |
| <i>Staurostrum</i> | - | 1.51 | - | - | 6.34 | - | 8.19 | - | - | 3.50 |
| <i>Coelastrum</i> | - | - | - | - | - | - | 8.19 | 10.3 | 9.67 | - |
| Chrysophyta | 40.4 | 31.8 | 22.9 | 33.8 | 20.6 | 33.3 | 24.6 | 34.5 | 16.1 | 26.3 |
| <i>Navicula</i> | 7.69 | - | 5.88 | 12.3 | - | 9.87 | 9.83 | 6.89 | - | - |
| <i>Cyclotella</i> | 13.5 | - | - | - | 11.1 | - | - | - | - | 8.77 |
| <i>Mallomonas</i> | 9.61 | - | 4.41 | - | - | - | 4.91 | - | - | - |
| <i>Tabellaria</i> | - | 9.09 | - | 9.23 | - | 8.63 | - | - | - | 12.3 |
| <i>Cocconeis</i> | - | 4.54 | 3.75 | - | - | - | 8.19 | - | - | - |
| <i>Melosira</i> | 3.84 | - | 8.82 | 1.53 | - | - | - | 12.1 | 1.61 | - |
| <i>Stephanodiscus</i> | - | - | - | 4.61 | - | - | - | - | - | 5.26 |
| <i>Fragilaria</i> | - | 7.57 | - | - | - | 6.17 | - | 8.62 | 4.83 | - |
| <i>Synedra</i> | - | - | - | 6.15 | - | 8.64 | - | - | - | - |
| <i>Nitzschia</i> | 5.76 | - | - | - | 9.52 | - | - | - | 9.67 | - |
| <i>Cymbella</i> | - | 10.6 | - | - | - | - | 1.63 | 6.89 | - | - |
| Englenophyta | 7.69 | - | 8.82 | 6.15 | 7.92 | 12.9 | 19.7 | 1.71 | 8.06 | 3.00 |
| <i>Euglenopsis</i> | 7.69 | - | - | - | 1.58 | 8.64 | - | - | - | - |
| <i>Phacus</i> | - | - | - | 6.15 | - | - | 8.19 | - | - | - |
| <i>Euglena</i> | - | - | - | - | 6.34 | - | - | 1.71 | 8.06 | - |
| <i>Lepocinclis</i> | - | - | 8.82 | - | - | 4.32 | 11.5 | - | - | 3.00 |
| Pyrrophyta | - | 9.09 | - | 7.69 | - | 4.32 | 3.27 | - | - | 7.60 |
| <i>Peridinium</i> | - | 9.09 | - | - | - | - | 3.27 | - | - | 7.60 |
| <i>Glenodinium</i> | - | - | - | 7.69 | - | 4.32 | - | - | - | - |
| Cryptophyta | - | 6.06 | 1.47 | - | 6.34 | - | - | 6.89 | 1.61 | 2.40 |
| <i>Cryptomonas</i> | - | 6.06 | 1.47 | - | - | - | - | - | 1.61 | - |
| <i>Nephroselmis</i> | - | - | - | - | 6.34 | - | - | 6.89 | - | 2.40 |

Table 3. Relative abundance (%) of Zooplankton in D.G. Khan Canal water, affected with domestic sewage.

| Zooplankton | Mar | Apr | May | Jun | Jul | Aug | Sep | Oct | Nov | Dec |
|-----------------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|
| Protozoan | 4.74 | 4.54 | 13.2 | 8.60 | 16.4 | 2.23 | 7.13 | 13.2 | 10.5 | 7.01 |
| <i>Holophrya</i> | 1.24 | - | - | - | 7.93 | - | - | - | - | - |
| <i>Didinium</i> | - | 4.54 | - | - | - | - | - | - | - | 1.75 |
| <i>Tintinnopsis</i> | - | - | 7.35 | - | - | 2.23 | - | - | - | - |
| <i>Hemiophrys</i> | - | - | - | 5.10 | - | - | - | 3.44 | - | - |
| <i>Centropyxis</i> | - | - | - | 3.50 | - | - | - | - | 5.27 | - |
| <i>Pseudodiffugia</i> | - | - | - | - | 8.45 | - | - | - | - | 5.26 |
| <i>Paramecium</i> | - | - | - | - | - | - | 7.13 | 9.80 | - | - |
| <i>Diffugia</i> | 3.50 | - | - | - | - | - | - | - | 1.15 | - |
| <i>Cyphoderia</i> | - | - | 5.86 | - | - | - | - | - | 4.04 | - |
| Rotifers | 1.05 | 9.66 | 2.16 | 2.26 | 9.12 | 4.08 | 2.65 | 10.3 | 4.31 | 2.34 |
| <i>Asplanchna</i> | - | 2.09 | - | - | 2.78 | - | - | - | - | - |
| <i>Epiphanes</i> | - | 7.57 | - | 1.11 | - | - | - | - | - | 2.34 |
| <i>Colurella</i> | - | - | - | - | 6.34 | 1.23 | 2.65 | 10.3 | - | - |
| <i>Dicranophorus</i> | 1.05 | - | 2.16 | 1.15 | - | 2.85 | - | - | 4.31 | - |
| Cladocerans | - | 7.09 | 4.95 | - | - | 6.17 | - | 2.39 | 5.12 | 3.99 |
| <i>Daphnia</i> | - | 7.09 | 4.95 | - | - | 6.17 | - | 2.39 | 5.12 | 3.99 |

Table 4. Relationship among limnological parameters of D.G. Khan Canal water affected with domestic sewage.

| Correlation matrix | WT | LP | Turbidity | pH | EC | DO ₂ | Free CO ₂ | Acidity | Hardness | Alkalinity | TS | NPP |
|----------------------|----------------------|----------------------|----------------------|----------------------|----------------------|----------------------|----------------------|----------------------|----------------------|----------------------|----------------------|-----------------|
| LP | 0.795** | | | | | | | | | | | |
| Turbidity | 0.563 ^{ns} | 0.590 ^{ns} | | | | | | | | | | |
| pH | 0.083 ^{ns} | 0.278 ^{ns} | 0.167 ^{ns} | | | | | | | | | |
| EC | -0.803** | -0.708* | -0.447 ^{ns} | -0.450 ^{ns} | | | | | | | | |
| DO ₂ | -0.577 ^{ns} | -0.905** | -0.565 ^{ns} | -0.549 ^{ns} | 0.630* | | | | | | | |
| Free CO ₂ | 0.187 ^{ns} | 0.465 ^{ns} | 0.369 ^{ns} | 0.779** | -0.466 ^{ns} | -0.732* | | | | | | |
| Acidity | -0.008 ^{ns} | 0.199 ^{ns} | -0.345 ^{ns} | 0.525 ^{ns} | -0.165 ^{ns} | -0.413 ^{ns} | 0.579 ^{ns} | | | | | |
| Hardness | -0.581 ^{ns} | -0.541 ^{ns} | -0.159 ^{ns} | -0.324 ^{ns} | 0.583 ^{ns} | 0.504 ^{ns} | -0.291* | -0.576 ^{ns} | | | | |
| Alkalinity | -0.103 ^{ns} | 0.068 ^{ns} | -0.061 ^{ns} | 0.694* | -0.201 ^{ns} | -0.407 ^{ns} | 0.795** | 0.846 ^{ns} | -0.405 ^{ns} | | | |
| TS | 0.280 ^{ns} | 0.487 ^{ns} | 0.341 ^{ns} | -0.198 ^{ns} | -0.273 ^{ns} | -0.391 ^{ns} | -0.036 ^{ns} | -0.421 ^{ns} | 0.288 ^{ns} | -0.448 ^{ns} | | |
| NPP | -0.313 ^{ns} | 0.079 ^{ns} | 0.327 ^{ns} | -0.105 ^{ns} | 0.425 ^{ns} | 0.085 ^{ns} | -0.048 ^{ns} | -0.448 ^{ns} | 0.477 ^{ns} | -0.393 ^{ns} | 0.360 ^{ns} | |
| NZP | 0.313 ^{ns} | -0.079 ^{ns} | -0.327 ^{ns} | 0.105 ^{ns} | -0.425 ^{ns} | -0.085 ^{ns} | 0.048 ^{ns} | 0.448 ^{ns} | -0.477 ^{ns} | 0.393 ^{ns} | -0.360 ^{ns} | -1.000** |

ns = non significant ($p \geq 0.05$), * = significant ($p < 0.05$), ** = highly significant ($p < 0.001$)

WT: Water Temperature, LP: Light Penetration, DO₂: Dissolved Oxygen, TS: Total Solids, NPP: Net Phytoplankton, NZP: Net Zooplankton

Table 5. Diversity indices of Phyto and Zooplankton in D.G. Khan Canal water affected with domestic sewage.

| Months | Phytoplankton | | | | Zooplankton | | | |
|-----------|---------------|----|------|-----------------|-------------|----|------|-----------------|
| | S | N | In N | Diversity Index | S | N | In N | Diversity Index |
| March | 11 | 52 | 3.95 | 2.53 | 3 | 04 | 1.39 | 1.44 |
| April | 11 | 49 | 3.89 | 2.57 | 4 | 14 | 2.63 | 1.14 |
| May | 12 | 59 | 4.07 | 2.70 | 4 | 09 | 2.20 | 1.36 |
| June | 13 | 58 | 4.06 | 2.95 | 4 | 07 | 1.94 | 1.55 |
| July | 11 | 47 | 3.85 | 2.59 | 4 | 16 | 2.77 | 1.08 |
| August | 13 | 67 | 4.20 | 2.86 | 4 | 14 | 2.64 | 1.14 |
| September | 13 | 55 | 4.01 | 2.99 | 3 | 06 | 1.79 | 1.12 |
| October | 11 | 42 | 3.74 | 2.67 | 4 | 16 | 2.77 | 1.08 |
| November | 12 | 41 | 3.71 | 2.96 | 5 | 21 | 3.04 | 1.32 |
| December | 12 | 53 | 3.97 | 2.81 | 4 | 06 | 1.79 | 1.68 |

S= Number of genera, N= Total number of individuals, In = Natural logarithm

Mason (1998) reported that diversity index is a good pollution indicator in aquatic ecosystem. Diversity index greater than three indicates the clean water; range from one to three is the characteristic of moderately polluted water and values less than three characterize the heavily polluted water. Chughtai *et al.*, (2011) reported the diversity index of Phytoplankton more than three while of Zooplankton less than three throughout the study period in river Chenab water at Multan and indicated its water quality marginally fit for aquatic life. Similarly, El-Sheekh *et al.*, (2010) also studied the water quality of river Nile at different locations and declared as moderately polluted on the basis of biological assessment through diversity and saprobic indices that were less than three throughout the study period.

Conclusions

In an aquatic ecosystem, limnological characteristics can affect both on fauna and flora. The water quality parameters of D.G. Khan were compared with water quality standards. Most of the parameters were found to be in permissible level throughout the study period. However, diversity index of Phytoplankton and Zooplankton was found less than three throughout the study period which indicates the quality of D.G. Khan Canal water as moderately polluted. It is, therefore, necessary to add sewage water after proper treatment to protect this water resource and also the aquatic life.

References

- Abbasi, S.A. 1998. *Water quality sampling and analysis*. (1st Edn.). Discovery Publishing House, New Delhi.
- Achionye-Nzeh, C.G. and A. Isimaikaiye. 2010. Fauna and flora composition and water quality of a reservoir in Ilorin, Nigeria. *Int. J. Lakes & Rivers*, 3(1): 7-15.
- Agbaire, P.O. and C.G. Obi. 2009. Seasonal variations of some physico-chemical properties of River Ethiope water in Abraka, Nigeria. *J. Appl. Sci. Environ. Manage.*, 13(1): 55-57.
- Ali, A., Z.K. Shinwari and F.M. Sarim. 2010. Contribution to the algal flora (Chlorophyta) of fresh waters of District Swat, N.W.F.P., Pakistan. *Pak. J. Bot.*, 42(5): 3457-3462.
- Anonymous. 2006. Status Quo Report Dera Ghazi Khan (SQR): Urban water supply and sewerage reform strategy. World Bank - Government of Punjab, Pakistan.
- Anonymous. 2007. Government of Pakistan (GoP). Pakistan economic survey 2006-2007. Islamabad: Ministry of Finance.
- Anonymous. 2008. Food and Agriculture Organization (FAO). State of food insecurity in the world. Rome.
- Barnabe, G. 1990. *Aquaculture*. Vol. 1, Ellis Harwood, New York.
- Basu, M., N. Roy and A. Barik. 2010. Seasonal abundance of net zooplankton correlated with physico-chemical parameters in a freshwater ecosystem. *Int. J. Lakes & Rivers*, 3(1): 67-77.
- Battish, S.K. 1992. *Freshwater zooplankton of India*. Oxford and IBH Publishing Co. Ltd., New Delhi. pp. 233.
- Bos, M.G., M.A. Burton and D.J. Molden. 2005. Irrigation and drainage performance assessment: Practical Guidelines. CABI Publishing, Cromwell Press: Trowbridge, UK.
- Boyd, C.E. 1981. *Water quality in warm water fish ponds*. Craftmaster Printers, Inc. Opelika, Alabama.
- Boyd, C.E. and C.S. Tucker. 1998. *Pond aquaculture water quality management*. Kluwer Academic Publishers, London.
- Brown, L. 1993. *Aquaculture for veterinarians fish husbandry and medicine*. Pergamon Press, Oxford. pp. 242-245.
- Chattopadhyay, C. and T.C. Banerjee. 2007. Temporal changes in environmental characteristics and diversity of net-phytoplankton in a freshwater lake. *Turk. J. Bot.*, 31: 287-296.
- Chughtai, M.I., K. Mahmood and A.R. Awan. 2011. Assessment of planktonic diversity in river Chenab as affected by sewage of Multan city. *Pak. J. Bot.*, 43(5): 2551-2555.
- El-Sheekh, M.M., M.A.I. Deyab, S.S. Desouki and M. Eladl. 2010. Phytoplankton compositions as a response of water quality in El- Salam canal Hadous drain and Damietta branch of river Nile Egypt. *Pak. J. Bot.*, 42(4): 2621-2633.
- Gany, H. 2006. An outlook of the recent development of irrigation and propensity for transformation into an efficient engine of growth. International seminar on the occasion of 70th anniversary of the RCWR, Bandung, Indonesia.
- Ghafoor, A., M. Azam and M. Shoaib. 1993. Characterization of tube well and hand pump waters in the Faisalabad tehsil. *Pakistan J. Soil Sci.*, 8: 1-2.
- Iqbal, J., M.W. Mumtaz, H. Mukhtar, T. Iqbal, S. Mahboob and A. Razak. 2010. Particle size distribution analysis and physico-chemical characterization of Chenab river water at Marala Headworks. *Pak. J. Bot.*, 42(2): 1153-1161.
- Khangerot, B.S. and S. Das. 2009. Acute toxicity of metals and reference toxicants to a freshwater ostracod, *Cypris subglobosa* Sowerby, 1840 and correlation to EC50 values of other test models, *J. Hazard. Mat.*, 172(2-3): 641-9.
- Laghari, K.Q., B.K. Lashari and H.M. Memon. 2008. Perceptive research on wheat evapotranspiration in Pakistan. *Irrigation and Drainage*, 57: 571-584.
- Mason, C.F. 1998. *Biology of freshwater pollution*. Longman scientific and technical, New York, USA. 351pp.
- Meshram, C.B. 2003. Macro invertebrate fauna of Lake Wadali; Amravati Maharashtra. *J. Aqu. Boil.*, 18(2): 47-50.
- Nabi, G., M.N. Akhtar and B.A. Khokhar. 1995. *Physical Chemistry*. Ilmi Kitab Khana, Lahore.
- Pathania, D., M. Sabesan and S. Kumari. 2010. Seasonal variation in physico-chemical parameters and planktons population of fish pond in Jalandhar, Punjab. *Asian J. Water Environ. Pollut.*, 7(1): 123-128.
- Pervaiz, Z. 2005. Characterization of irrigation quality of ground water in union council Ghakhra Kalan, District Gujrat (Pakistan). *Int. J. Agri. Biol.*, 7(2): 269-271.
- Prabhakar, C., K. Saleshrani and K. Tharmaraj. 2011. Hydrobiological investigations on the planktonic diversity of Vellar River, Vellar Estuary and Portonovo coastal waters, South East Coast of India. *Int. J. Pharmaceutical & Biological Archives*, 2(6): 1699-1704.
- Rahman, S. and A.F. Hussain. 2008. A study on the abundance of zooplankton of a culture and non-culture pond of the Rajshahi University campus. *Univ. J. Zool. Rajshahi. Univ.*, 27: 35-41.
- Rather, G.M., M.S. Bhat and T.A. Kanth. 2010. Impact of urban waste of Srinagar city on the quality of water of river Jehlum. *Int. J. Lakes & Rivers*, 3(1): 17-24.
- Revenga, C. and G. Mock. 2000. Pilot analysis of global ecosystems: Freshwater systems. World Resources Institute, Washington, DC.
- Schabetsberger, R., G. Drozdowski, I. Drozdowski, C.D. Jersabek and E. Rott. 2004. Limnological aspects of two tropical crater lakes (Lago Biao and Lago Loreto) on the island of Bioko (Equatorial Guinea). *Hydrobiologia*, 524: 79-90.
- Tonapi, G.T. 1980. *Freshwater animals of India*. Oxford and IBH Publishing Co. Ltd., New Delhi.
- Vandysh, O.I. 2004. Zooplankton as an indicator of the state of lake ecosystem polluted with mining waste water in the Kola Peninsula. *Russian J. Ecol.*, 35(2): 110-116.
- Ward, H.B. and G.C. Whipple. 1959. *Freshwater biology*. (2nd Edn.) John Wiley and Sons Inc., New York. pp. 248.