EFFECT OF GIBBERELLIC ACID ON ENHANCING FLOWERING TIME IN CHRYSANTHEMUM MORIFOLIUM

MUHAMMAD SAJID^{1*} NOORUL AMIN², HABIB AHMAD¹AND KHALID KHAN³

¹Department of Genetics, Hazara University Mansehra, Pakistan ²Department of Horticulture, The University of Agriculture, Peshawar, Pakistan ³Sawabi Women University Sawabi, Pakistan *Corresponding author's email: drsajid_1@yahoo.com

Abstract

The study was conducted to evaluate the effect of various concentrations of GA₃ on plant height, number of branches, leaves, flowers plant⁻¹, leaf area, days to flowering, blooming period, flower size and flower fresh weight. There were total six treatments of GA₃ concentrations at 0, 50, 100, 150, 200 and 250 mg L⁻¹. Data regarding vegetative and flowering attributes indicated that flowering in *Chrysanthemum morifolium* varied significantly for most of the studied parameters. Application of GA₃ at the rate of 250 mg L⁻¹ with plant height (62.0 cm), number of branches (8.65), suckers (8.05), leaves (55.05) plant⁻¹, leaf area (122.35 cm²), days to flower (110.8 days) and number of flowers (31.9) were significantly different as compared to rest of the treatments and was followed by GA₃ at the rate of 200 mg L⁻¹ in flower size (5.49 cm) and flower fresh weight (4.01g). Untreated Chrysanthemum plants had least plant height (47.5 cm), number of branches (6.4), suckers (6.75), leaves (39.2) plant⁻¹, leaf area (96.85 cm²), days to flower (131.05 days), number of flowers (21.91) plant⁻¹, flower size (4.88 cm) and flower fresh weight (3.44 g). Application of GA₃@ 250 mg L⁻¹ produced flowers earlier (28th September) than the normal season and extended the flowering season up to 22 days.

Key words: Chrysanthemum flowering, Gibberellic acid, Early flowering.

Introduction

Chrysanthemum (Chrysanthemum morifolium) is commonly known as Gul-e-Daudi or Queen of the East. It belongs to the family Asteraceae. It is highly attractive short day plant, which behaves both as an annual as well as perennial flowering herb (Ghafoor & Khan, 2002). This Attractive flowering plant is extremely popular all over the world. It starts to bloom during autumn season, while in Pakistan, its peak blooming period is month of November and December. In Hazara Division, it blooms from 15th to 25th October. Flowers are attractive, showy and their popularity has increased not only due to their outstanding aesthetic beauty but also due to their excellent potential of export as cut flowers to many countries of world (Erler & Seigmund, 1986). The plant growth regulators are compounds that in minor amounts modify the physiological processes of plants and ultimately alter the yield and quality. Numerous plant growth regulators have been widely used in many flowering plants and their efficacy have been demonstrated for nursery production, foliage plants and many other ornamental plants (Sanap et al., 2000). Among the major groups are auxins, cytokinins, abscisic acid, gibberellins, polyamines and ethylene (Salisbury & Ross, 1992). Gibberellin was first recognized in 1926 by a Japanese scientist, Eiichi Kurosawa who was studying bakanae or "foolish seedling" disease in rice (Salisbury & Ross, 1992). Three gibberellins are known as gibberellic acid $(C_{19}H_{22}O_6)$, gibberellin A₁ ($C_{19}H_{24}O_6$) and gibberellin A₂ ($C_{19}H_{26}O_6$). The biological activity of all three gibberellins is qualitatively similar. Young and growing meristematic tissues, apical root cells, young fruits and germinating seeds are rich in gibberellins (Brian, 2008).

The most characteristic effects of GA₃ on shoot growth are increase in inter-node extension, increase in leaf-growth, increase in diameter of plant, increase in number of flowers, induce flowering and enhanced apical dominance (Medina & Saavedra, 1999; Taiz & Zeiger, 2004). In its effects on leaf expansion and on some forms of dormancy, GA_3 simulates light. In most photoperiodically sensitive plants, particularly in the form of long-day photoperiod, induces increased shoot growth. GA_3 has a similar effect. There is thus a fundamental unity in the effects of GA_3 on plant development, in which GA_3 closely simulates effects usually induced in nature either by exposure to light or by vernalization (Brian, 2008). It also has stimulant effect on germination (Shohani *et al.*, 2014).

It has been observed that gibberellins cause early flowering in those biennial plants that need a cold period (chilling requirement) before they flower (Hartmann *et al.*, 1981). Gibberellins function as plant growth regulators influencing a range of developmental processes in plants life like stem elongation, germination, breaking dormancy, flowering, sex expression, enzyme induction and leaf and fruit senescence. Spraying of GA₃ recorded maximum plant height, plant spread and more number of leaves and branches in chrysanthemum and other flowering plants (Lal & Mishra, 1986; Nagarjuna *et al.*, 1988; Sujatha *et al.*, 2002; Kumar *et al.*, 2003; Rana *et al.*, 2005).

Gibberellins initiates early flowering in many ornamental plants and increases the number of flowers. Spraying of GA₃ induced early flowering, increased size of the flowers, fresh weight and dry weight of flowers in Chrysanthemum (Nagarjuna *et al.*, 1988); (Koriesh *et al.*, 1989). Spraying of GA₃ gave maximum number of flowers per plant, flower weight and flower yield (Kumar *et al.*, 2003), while stalk length and spathe length increase with foliar application of GA₃ in anthurium (Dhaduk *et al.*, 2007). Whereas, Devadanam *et al.* (2007) observed minimum number of days required for spike emergence maximum spike length with foliar spray of GA₃. Lower concentrations of GA₃ decreases number of days to 50% flowering, increases number of leaves, spike girth, spike length, rachis length, floret length and floret diameter in marigold and tuberose (Panwar *et al.*, 2006; Samruban & Karuppaiah, 2007; Devadanam *et al.*, 2007). Keeping in view the above mentioned important properties of the gibberellic acid, this experiment was designed to study the influence of various doses of GA₃ on regulation of flowering time in Chrysanthemum.

Materials and Methods

The studies were carried out during 2010-11 at Hazara Agriculture Research Station Abbottabad Khyber Pakhtunkhwa, Pakistan. The experiment was laid-out in Completely Randomized Design with six doses of GA₃. Terminal cuttings of chrysanthemum cv. Fanfare were taken from the stock and were planted in 7 cm plastic pots separately on 10th June. The clay pots 28 cm size were filled with 2 parts leaf mould and one part silt and plants were transplanted to clay pots separately on 10th July. Fertilizer NPK was applied at the rate of 1.5 g to each pot. GA₃ solutions were sprayed on plants in the morning at 15 and 30 days after transplanting. There were total six treatments including; a control with no GA₃ spray. Other treatments included 50, 100, 150, 200 and 250 mg L^{-1} of GA₃ were designated as T1, T2, T3, T4, T5 and T6 respectively.

All the cultural practices were kept uniform for all the treatments in the experiment. The experiment with all same inputs and treatments was repeated in 2011 and average data of both years were analyzed at the end using computer statistical software "Statistix 9.0".

The physical traits considered included plant height which was the measure of stem length from the crown to the top of the stem. The number of branches plant ⁻¹ grown on plant were counted and recoded after the last flower harvested. All the leaves grown on plant were counted and recorded after the last flower harvested. Leaf area was measured with the help of an automatic Leaf Area meter. Days taken to flowering were counted from

date of transplanting to small pots (10th June) till the date of flower bud opening. The number of flowers, flower size (diameter) and fresh weight were then taken after harvest. All flowers grown on the main stalk and the side branches were counted up to the last flower harvested. Number of days from flower bud break till its petal fadding were counted. The flower size was recorded by measuring the diameter of the flower in cm. Full bloomed flowers were excised and weighed on electronic balance individually.

Results

Vegetative attributes

Plant height (cm): The data regarding plant height revealed that GA₃ had profound effect on plant height while the year wise as well as GA₃ interaction with year did not affect it. Plant height was (62.0 cm) when GA₃ @ 250 mg L⁻¹ was applied, while it was lower (47.5 cm) in untreated plants. Untreated plants were significantly different from those treated with GA₃ @ 50, 100, 150, 200 and 250 mg L⁻¹, while treatments GA₃ @ 150 and 200 mg L⁻¹ were non significantly different from each other. Plants treated with GA₃ @ 200 and 250 mg L⁻¹ also had non significant difference regarding plant height (Table 1).

Number of branches plant⁻¹: The data in table 1 indicated that GA₃ had pronounced effect on number of branches in Chrysanthemum. Number of branches were more (8.65) when plants were subjected to GA₃ @ 250 mg L⁻¹ followed by those plants that were treated with GA₃ @ 200 mg L⁻¹ (8.05), while branches were less (6.4) in untreated plants followed by (6.45) GA₃ @ 50 mg L⁻¹. Branches produced in plants treated with control, GA₃ @ 50 and 100 mg L⁻¹ were non significantly different from each other. Statistically similar effect was recorded between and GA₃ @ 200 and 250 mg L⁻¹. There was significant difference in branches between untreated plants and plants treated with GA₃ @ 150, 200 and 250 mg L⁻¹.

 Table 1. Effect of Gibberellic Acid on plant height, number of branches/plant and number of suckers/

 plant in Chrysanthemum for year 1 (2010) and year 2 (2011).

| GA ₃ (ppm) | Plant height (cm) | | | No. of branches plant ⁻¹ | | | No. of suckers plant ⁻¹ | | |
|------------------------|-------------------|---------|---------|-------------------------------------|---------|---------|------------------------------------|---------|---------|
| | 2010 | 2011 | Mean | 2010 | 2011 | Mean | 2010 | 2011 | Mean |
| Control | 46.2 d | 48.8 c | 47.5 e | 7.1 bc | 5.7 c | 6.40 c | 6.8 d | 6.7 d | 6.75 c |
| 50 | 54.3 c | 49.8 c | 52.1 d | 6.3 c | 6.6 c | 6.45 c | 6.9 d | 6.8 cd | 6.85 c |
| 100 | 54.5 bc | 58.2 b | 56.1 c | 6.7 c | 7.3 c | 7.00 bc | 7.3 cd | 7.3 bc | 7.30 b |
| 150 | 58.4 abc | 57.7 b | 58.2 bc | 7.3 bc | 7.2 c | 7.25 b | 7.5 bc | 7.6 ab | 7.55 b |
| 200 | 60.2 ab | 59.7 b | 60.0 ab | 7.9 ab | 8.2 b | 8.05 a | 7.9 ab | 8.0 a | 7.91 a |
| 250 | 60.5 a | 63.5 a | 62.0 a | 8.8 a | 8.5 a | 8.65 a | 8.1 a | 8.0 a | 8.05 a |
| LSD (p=0.05) | 5.8(**) | 3.6(**) | 3.2(**) | 1.1(**) | 0.8(**) | 0.7(**) | 0.5(**) | 0.5(**) | 0.4(**) |
| Year | | | | | | | | | |
| 2010 | | | 55.69 | | | 7.35 | | | 7.39 |
| 2011 | | | 56.24 | | | 7.24 | | | 7.38 |
| LSD (<i>p</i> =0.05) | | | NS | | | NS | | | NS |
| GA ₃ x year | | | | | | | | | |
| LSD(p=0.05) | | | NS | | | NS | | | NS |

Means followed by similar letters in a column are non significantly different from each other at a 0.05

NS = Non significant

* = Significant at 5% level of probability

Number of suckers plant⁻¹: Table 1 showed pronouncedly increasing trend in number of suckers plant⁻¹ with the increase in concentration of GA₃. More number of suckers (8.05) were recorded in plants treated with GA₃@ 250 mg L⁻¹ closely followed by (7.91) in the plants treated with GA₃@ 200 mg L⁻¹, while less number of suckers were produced in untreated plants. Number of suckers produced in untreated plants and plants treated with GA₃@ 50 mg L⁻¹ were statistically same. Similar trend was observed between GA₃@ 100 mg L⁻¹ and GA₃@ 150 mg L⁻¹. GA₃@ 200 mg L⁻¹ and GA₃@ 250 mg L⁻¹ were also at par regarding number of suckers plant⁻¹.

Number of leaves plant⁻¹: There was substantial difference in number of leaves in Chrysanthemum as affected by various concentrations of GA₃. Higher number of leaves (55.05) were recorded in plants treated with GA₃ @ 250 mg L⁻¹, while it was less (39.20) in control. Profound difference was recorded between control and GA₃ @ 100, 150 and 250 mg L⁻¹. The effects of untreated plants and GA₃ @ 50 mg L⁻¹ were non-significantly different. Plants treated with GA₃ @ 200 and 250 mg L⁻¹ also showed non significant difference with each other (Table 2).

Leaf area (cm²): Data regarding leaf area indicates considerable difference among various treatments of GA₃ (Table 2). Broader area (122.35 cm²) was recorded when plants were treated with GA₃ @ 250 mg L⁻¹, while it was smaller (96.85 cm²) in control. The effect of (control and GA₃ @ 50 mg L⁻¹), (GA₃ @ 200 and 250 mg L⁻¹) were statistically similar in their effects while rest of the treatments were found significantly different from each other.

The interactive effect of GA₃ and year was significant on leaf area (Fig. 1). Broader leaves (125.3 cm²) were recorded at interaction of year 1 with GA₃@ 250 mg L⁻¹ application non significantly followed by (122.3 cm²) year 1 with GA₃@ 200 mg L⁻¹ application, while less leaf area (93.7 cm²) was recorded at year 1 with control followed non significantly by year 1 (97.9 cm²) with GA₃ (a) 50 mg L^{-1} application. The difference might be attributed to plant height.

Flowering attributes

Number of days to flowering: Foliar application of GA₃ extensively affected number of days to flowering in *Chrysanthemum* (Table 3). Untreated plants took significantly more days to flower (131.05 days) as compared to the plants that were sprayed with 250 mg L⁻¹ GA₃ that took less days (110.8 days) to flower. Days to flower were statistically unchanged at control, GA₃ @ 50 and 100 mg L⁻¹ treatments on plants. Significant difference was recorded between control and GA₃ @ 150, 200 and 250 mg L⁻¹. Plants treated with GA₃ @ 150 and 200 mg L⁻¹ were non significantly different from each other regarding number of days to flowering. Plants treated with GA₃ @ 100 and 150 mg L⁻¹ were non significantly different from each other.

Number of flowers plant⁻¹: The data in table 3 indicated that GA₃ deeply affected the number of flowers plant⁻¹. Plants produced more number of flowers (31.9) when treated by GA₃ @ 250 mg L⁻¹, followed by (30.0) GA₃ @ 200 mg L⁻¹ as compared to control which produced less (21.91) flowers. Effects of untreated plants were significantly different from GA₃ @ 150, 200 and 250 mg L⁻¹. GA₃ @ 200 and 250 mg L⁻¹ had statistically same effect on number of flower. Likewise control and GA₃ @ 50 and 100 mg L⁻¹ had similar effect regarding number of flowers plant⁻¹.

The interactive effect of GA₃ and year was also significant on number of flowers plant⁻¹ (Fig. 2). Higher number of flowers (32.7) were recorded at interaction of year 1 with GA₃@ 250 mg L⁻¹ application closely followed by same year with GA₃@ 200 mg L⁻¹ application (32.05), while less (19.5) number of flowers were recorded at year 1 with control followed significantly by same year with GA₃ @ 50 mg L⁻¹ application (23.17). The difference might be attributed to seasonal variations.

| $C \wedge (nnm)$ | 1 | Number of leave | es plant ⁻¹ | Average leaf area (cm ²) | | | |
|------------------------|---------|-----------------|------------------------|--------------------------------------|----------|----------------|--|
| GA ₃ (ppm) | 2010 | 2011 | Mean | 2010 | 2011 | Mean | |
| Control | 36 e | 43 b | 39.20 e | 93.7 d | 100.0 c | 96.85 d | |
| 50 | 43 cd | 37 c | 40.00 de | 97.9 cd | 100.9 c | 99.40 d | |
| 100 | 40 de | 45 b | 42.50 d | 102.7 c | 110.7 b | 106.70 c | |
| 150 | 48 bc | 45 b | 46.50 c | 113.7 b | 109.0 b | 111.40 b | |
| 200 | 49 b | 53 a | 51.00 b | 127.3 a | 113.1 ab | 120.20 a | |
| 250 | 56 a | 54 a | 55.05 a | 125.3 a | 119.4 a | 122.35 a | |
| LSD(<i>p</i> =0.05) | 4.9(**) | 3.8(**) | 3.0(**) | 5.1(**) | 6.9(**) | 4.2(**) | |
| Year | | | | | | | |
| 2010 | | | 45.9 | | | 109.28 | |
| 2011 | | | 46.5 | | | 109.69 | |
| LSD (<i>p</i> =0.05) | | | NS | | | NS | |
| GA ₃ x Year | | | | | | | |
| LSD(p=0.05) | | | NS | | | *5.87 (Fig. 1) | |

Table 2. Effect of Gibberellic Acid on number of leaves plant⁻¹ and leaf area in *Chrysanthemum*.

Means followed by similar letters in a column are non significantly different from each other at $\alpha 0.05$

NS = Non significant

* = Significant at 5% level of probability



Fig. 1. Interactive effect of GA₃ and Year on leaf area.

Fig. 2. Interactive effect of GA₃ and year on number of flowers plant⁻¹.

Table 3. Effect of Gibberellic Acid on number of days to flowering, number of flowers/ plant and blooming period in *Chrysanthemum*.

| plant and blooming period in <i>Chrysanthemum</i> . | | | | | | | | | | | |
|---|-----------------------------|---------|-----------|---------------------------------------|--------|----------------|-----------------|------|-------|--|--|
| | Number of days to flowering | | | Number of flowers plant ⁻¹ | | | Blooming period | | | | |
| GA ₃ (ppm) | 2010 | 2011 | Mean | 2010 | 2011 | Mean | 2010 | 2011 | Mean | | |
| Control | 132 a | 130 a | 131.05 a | 20 c | 24 b | 21.91 c | 35 | 38 | 36.90 | | |
| 50 | 130 a | 125 ab | 127.68 a | 21 c | 27 b | 23.95 c | 37 | 36 | 36.50 | | |
| 100 | 123 b | 118 c | 120.40 b | 25 b | 24 b | 24.35 c | 38 | 36 | 37.00 | | |
| 150 | 115 c | 120 bc | 117.48 bc | 28 b | 26 b | 27.00 b | 39 | 36 | 37.75 | | |
| 200 | 112 c | 116 cd | 114.20 cd | 35 a | 25 b | 30.00 a | 39 | 37 | 37.70 | | |
| 250 | 111 c | 111 d | 110.80 d | 33 a | 31 a | 31.90 a | 39 | 37 | 37.95 | | |
| LSD (<i>p</i> =0.05) | 6.3(**) | 5.8(**) | 4.1(**) | 4.0(**) | 3.9(*) | 4.5(**) | NS | NS | NS | | |
| Year | | | | | | | | | | | |
| 2010 | | | 120.6 | | | 26.8 | | | 37.9 | | |
| 2011 | | | 119.9 | | | 26.2 | | | 36.7 | | |
| LSD (p=0.05) | | | NS | | | NS | | | NS | | |
| GA ₃ x Year | | | | | | | | | | | |
| LSD(<i>p</i> =0.05) | | | NS | | | *3.47 (Fig. 2) | | | NS | | |
| | | | | | | | | | | | |

Means followed by similar letters in a column are non significantly different from each other at $\alpha 0.05$

NS = Non significant

* = Significant at 5% level of probability

Blooming period: The difference regarding flower blooming period against various treatments of GA₃ application was non significant. The parameter was not affected by year and GA₃ interaction. The blooming period was higher (37.95 days) in plants treated with GA₃ @ 250 mg L⁻¹ whereas it was lower (36.5 days) in GA₃ @ 50 mg L⁻¹. All concentrations were non-significantly different from each other at 5% level of significance (Table 3).

Flower size (cm): The flower size was remarkably affected by various concentrations of GA₃ (Table 4). Bigger size (5.55 cm) was recorded in GA₃ @ 200 mg L⁻¹ followed (5.49 cm) by GA₃ @ 250 mg L⁻¹. Smaller size was recorded in control (4.88 cm). The effects of GA₃ @ 150, 200 and 250 mg L⁻¹ were statistically same. Control and GA₃ @ 50 mg L⁻¹ exhibited non significant difference regarding flower size. Plants sprayed with GA₃ @ 100 mg L⁻¹ recorded non significant difference with those plants treated with GA₃ @ 50 mg L⁻¹ but significant difference with rest of the treatments (Table 4).

Flower fresh weight (g): Flower fresh weight was strongly affected by various concentrations of GA₃, while it was not affected by year and interactive effected of year and GA₃ (Table 4). Significantly higher fresh weight (4.16 g) was recorded when GA₃ @ 200 mg L⁻¹ was sprayed on plants, while it was less (3.44 g) in control. The effects of GA₃ @ 50, 100 and 150 mg L⁻¹ were statistically same. Plants treated with GA₃ @ 200 mgl⁻¹ were significantly different from control, GA₃ @ 50, 100 and 150 mg L⁻¹, while they were non significantly different from the plants treated with GA₃ @ 250 mg L⁻¹ regarding flower fresh weight.

Discussion

GA₃ treated plants showed remarkable increase in plant height. Taller plants were observed when treated with GA₃ @ 250 mg L⁻¹ closely followed by the application of GA₃ @ 200 mg l⁻¹. Significantly less height (47.55 cm) was recorded in control. Plant

height increased with the increasing doses of GA₃. Foliar application of GA₃ might have influenced the stem elongation by stimulating cell division and elongation. These findings are in line with those reported by Talukdar & Paswan (1994) who noticed an increase in plant height with the increase in doses of GA₃ concentrations in chrysanthemum. Schmidt et al. (2003) found 16.78% increase in plant height when applied GA_3 at 300 mg L⁻¹ on chrysanthemum c.v Viking. Kumar et al. (2012) also recorded same trend in carnation.

Number of branches increased with the increasing doses of GA₃ as compared to the control. Foliar application of GA₃ might have influenced the stem elongation by stimulating cell division and elongation that resulted in enhanced branches and vegetative growth. Nagarjuna et al. (1988) recorded more number of branches with 200 ppm GA₃ as compared to control in Chrysanthemum. These findings are also in agreement with those of Dabas et al. (2001) in marigold and Krishnamoorthy & Madalagery (2000) recorded increased number of branches in Ajovan.

| | I | Flower size (cm | ı) | Flower fresh weight (g) | | | |
|-----------------------|---------|-----------------|---------|-------------------------|--------|---------------|--|
| GA ₃ (ppm) | 2010 | 2011 | Mean | 2010 | 2011 | Mean | |
| Control | 4.8 cd | 5.0 c | 4.88 c | 3.4 c | 3.5 b | 3.44 c | |
| 50 | 4.7 d | 5.3 b | 4.99 bc | 3.7 b | 3.9 a | 3.84 b | |
| 100 | 4.9 c | 5.4 ab | 5.19 b | 3.8 b | 3.9 a | 3.85 b | |
| 150 | 5.4 b | 5.5 a | 5.42 a | 3.9 b | 4.0 a | 3.95 b | |
| 200 | 5.6 a | 5.5 a | 5.55 a | 4.2 a | 4.1 a | 4.16 a | |
| 250 | 5.5 ab | 5.4 ab | 5.49 a | 4.1 a | 3.9 a | 4.01 ab | |
| LSD (<i>p</i> =0.05) | 0.2(**) | 0.2(**) | 0.2(**) | 0.2(**) | 0.4(*) | 0.2(**) | |
| Year | | | | | | | |
| 2010 | | | 5.18 | | | 3.84 | |
| 2011 | | | 5.31 | | | 3.90 | |
| LSD (<i>p</i> =0.05) | | | NS | | | NS | |
| GA ₃ xYear | | | | | | | |
| LSD (<i>p</i> =0.05) | | | NS | | | *5.87 (Fig.2) | |

Table 4. Effect of gibberellic acid on flower size and flower fresh weight in Chrysanthemum.

NS = Non significant

* = Significant at 5% level of probability

The number of suckers increased with the increase in concentration. Higher number of suckers produced by GA₃ application might be attributed to the increase in number of branches, number of leaves and leaf area, which ultimately increased the translocation of photosynthates to produce more suckers. These findings are in agreement with the findings of Sharifuzzaman et al. (2011) who recorded an increase in number of suckers with GA₃ application in Chrysanthemum.

Greater number of leaves were recorded at GA₃ @ 250 mg L^{-1} . It might be attributed to the effect of GA_3 which increases cell division, cell elongation and tissue differentiation that resulted in the initiation of more number of leaves. The number of branches were also higher with the application of GA₃ which helped to initiate more leaves. More number of leaves plant⁻¹ as compared to control in Chrysanthemum has been reported by Talukdar & Paswan (1988). Sharifuzzaman et al. (2011) recorded maximum number of leaves at 150 ppm GA3 treatment in Chrysanthemum. These results are also supported by those of Radives et al. (1992) in gladiolus.

Plants showed a significant increase in leaf area. This might be attributed to the fact that GA₃ stimulates the cell division and cell elongation, which ultimately results in an increase in leaf area and also the number of leaves. These results are also in agreement with those of Sharifuzzaman et al. (2011) recorded significant increase

in leaf length by the application of different concentrations of GA₃ in Chrysanthemum Singh (2003) recorded broader leaves in calendula by GA₃ application.

Chrysanthemum plants produced early flowers in all GA treatments as compared to control. It is the effect of GA₃ that causes flower initiation and early flowering by decreasing the concentration of ABA in plant shoot (Phengphachanh et al., 2012). Moreover as the number of leaves was increased, it resulted in more photosynthates to initiate early flowering and complete life cycle of the plant. The findings are in line with those reported by Dutta et al. (1993) who noticed early flowering in chrysanthemum when treated plants with GA₃. Sharifuzzaman et al. (2011) recorded early flowering with the application of 150 ppm GA₃ as compared to control in chrysanthemum. Kumar et al. (2012) noticed earliness in flowering in carnation as affected by all concentrations of GA3.

The increase in number of flowers by GA₃ application might be due to increase in number of leaves as well as leaf area as compared to control, which might have enhanced the production and accumulation of increased photosynthates that were diverted to the sink and produced more flowers (Sharifuzzaman et al., 2011). These results are confirmed from those reported by Kumar et al. (2012) who recorded a substantial increase in number of flowers when plants were treated with GA3 at 150 ppm in carnation. Double application with GA₃

accelerated flower bud development of Ajania pacifica (Zalewska & Antkowiak, 2013)

GA₃ treatment of chrysanthemum plants resulted an increase in flower size. The increase in size of flower might be attributed to the increase in number of leaves and leaf area that produced more photosynthates which in turn might have increased the flower size. Spraying of GA₃ at 200 ppm (Nagarjuna *et al.*, 1988) or twice at 100 ppm (Koriesh *et al.*, 1989) induced early flowering and increased size of the flowers in *Chrysanthemum*. Bigger sized flowers as a result of GA₃ application have also been reported by Sharifuzzaman *et al.* (2011) in *Chrysanthemum*. Kumar *et al.* (2012) recorded increase in flower size in all GA₃treatments as compared to control in carnation. The findings of all these researchers confirm the findings of the current study.

Application of various concentrations of GA_3 increased the flower fresh weight in Chrysanthemum. The increase in fresh flower weight might be due to an increase in leaf area and number of leaves that resulted in production of more photosynthates which diverted to flowers to increase fresh weight. Kumar *et al.* (2012) recorded significant increase in flower weight in carnation affected by all concentrations of GA^3 . Application of GA_3 at 200 ppm or twice at 100 ppm increased the flower fresh weight and dry weight of *Chrysanthemum* flowers (Nagarjuna *et al.*, 1988; Koriesh *et al.*, 1989). These findings are also similar with those reported by Singh (2003b) who recorded increase in weight by GA_3 treatment in calendula.

Conclusion

From the results obtained during the study it can be concluded that among various gibberellic acid concentrations, treatment GA₃ @ 250 mg L⁻¹ was superior with 22 days early flowering (28^{th} September). This concentration not only helped to increase the flowering time rather it increased the number of branches, leaves and leaf area was also higher. It is further concluded that GA₃ @ 200 mg L⁻¹ also showed better performance with higher flower size and flower fresh weight, while it performed close to GA₃ @ 250 mg L⁻¹ regarding plant height, number of branches, days to flowering (114 days), number of leaves, leaf area, number of flowers and blooming period.

References

- Brian, P.V. 2008. Effects of Gibberellins on plant growth and development. *Biol. Rev.*, 34(1): 37-77.
- Dabas, H.K., L. Mitra and S. Dabas. 2001. Effect of different concentrations of GA₃ MH and NAA on primary branches of marigold (*Tagetes erecta* L.). *Indian Agric.*, 45(3-4): 265-267.
- Devadanam, A., B.N. Shinde, P.B. Sable and S.G. Vedpathak. 2007. Effect of foliar application of plant growth regulators on flowering and vase life of tuberose (*Polianthes tuberosa* L.). J. Soil and Crops, 17(1): 86-88.
- Dhaduk, B.K., S. Kumara, A. Singh and J.R. Desai. 2007. Response of gibberellic acid on growth and flowering attributes in anthurium (*Anthurium andreanum* Lind.). J. Orn. Hort., 10(3): 187-189.

- Dutta, J.P., R. Seemanthini, M. A. Khader and S. Ramdas. 1993. Regulation of flowering by growth regulator in Chrysanthemum (*Chrysanthemum indicum* L.) cv.CO-1. *South Indian Horti.*, 41(5): 293-299.
- Erler, R. and I. Siegmund 1986. Year Book of the International Horticultural Statistics. p: 44. USA.
- Ghafoor, A. and R. Khan. 2002. Anthemideae. In: Flora of Pakistan, (Eds.): S.I Ali & M. Qaiser. University of Karachi, Missouri Botanical Garden, Press, 207: 45-47.
- Hartmann, H.T., W.J. Flocker and A.M. Kofranek. 1981. Plant Science, growth, development and utilization of cultivated plants. pp. 125-126.
- Koriesh, E.M., A.M. A. Dahab and E.W.M. Ali. 1989. Physiological studies on *Chrysanthemum morifolium* Ramat. Effect of cycocel, gibberellic acid and nucleic acids on vegetative growth and some chemical components. Assiut J. Agric. Sci., 20 (1): 27-41.
- Krishnamoorthy, V. and M.B. Madalageri. 2000. Influence of plant growth regulating on growth and seed yield and oil content in ajowan (*Trachyspermum ammi L.*). *Indian Perfumer*, 44(4): 255-259.
- Kumar, P., S.P.S. Raghava, R.L. Mishra and K.P. Singh. 2003. Effect of GA₃ on growth and yield of China aster. J. Orn. Hort., 6(2): 110-112.
- Kumar, V., V. Kumar, V. Umrao and M. Singh. 2012. Effect of GA₃ and IAA on growth and flowering of carnation. *Hort. Flora Res. Spect.*, 1(1): 69-72.
- Lal, H. and S.D. Mishra. 1986. Effect of gibberellic acid and Maleic hydrazide on growth and flowering of marigold and aster. *Prog. Hort.*, 18(1-2): 151-152.
- Medina, E.O. and A.L. Saavedra .1999. The use of regulators in floriculture Mexican. *Sci. Desar, Bogota.*, 148: 1-17.
- Nagarjuna, B., V.P. Reddy, M.R. Rao and E.N. Reddy. 1988. Effect of growth regulators and potassium nitrate on growth, flowering and yield of chrysanthemum (*Chrysanthemum indicum* L.). South Indian Hort., 36: 136-140.
- Panwar, R. D., S.S. Sindhu, J.R. Sharma and R.S. Saini. 2006. Effect of gibberellic acid spray on growth, flowering, quality and yield of bulbs in tuberose. *Haryana J. Hort. Sci.*, 35(3/4): 253-255.
- Phengphachanh, B., D. Naphrom, W. Bundithya and N. Potapohn. 2012. Effects of Day-length and Gibberellic Acid (GA3) on Flowering and Endogenous Hormone Levels in Rhynchostylis gigantea (Lindl.) Ridl. J. of Agric Sci., 4(4): 217-222.
- Radives, L., P.K. Rajeevan and P.K. Valasalakumari. 1992. Effect of foliar application of growth regulators on the growth, flowering and corn yield of gladiolus cv. Friendship. South Indian Hort., 40: 329-335.
- Rana, P., J. Kumar and M. Kumar. 2005. Response of GA3, plant spacing and planting depth on growth, flowering and corm production in gladiolus. J. Orn. Hort., 8(1): 41-44.
- Salisbury, F.B. and C.W. Ross.1992. *Plant Physiology*. 4th ed. Belmont, Company. pp. 682.
- Samruban, J. and P. Karuppaiah. 2007. Effect of plant growth regulators on growth and yield of French marigold (*Tagetes paluta* L.). J. Asian Hort., 3(3): 162-165.
- Sanap, P.B., B.A. Patil and B.V. Gondhali. 2000. Effect of growth regulators on quality and yield of flowers in tuberose (*Polianthes tuberosa* L.) cv. Single. Orissa J. Hort., 28 (1): 68-72.
- Schmidt, C., A.B. Belle, C. Nardi and A.K. Toledo. 2003. The gibberellic acid (GA₃) in the cut chrysanthemum (*Dedranthema grandiflora* Tzevelev.) viking: planting summer/autumn. *Revista Sci. Rural.*, 33(2): 1451-1455.

- Sharifuzzaman, S.M., K.A. Ara, M.H. Rahman, K. Kabir and M.B. Talukdar. 2011. Effect of GA₃, CCC and MH on vegetative growth, flower yield and quality of chrysanthemum. *Int. J. Expt. Agric.*, 2(1): 17-20.
- Shohani, F., A.A. Mehrabi, R.A. Khawarinegad, Z. Safari and S. Kian. 2014. The effect of Gibberellic acid (GA₃) on seed germination and early growth of lentil seedlings under salinity stress. *Middle-East J. Sci. Res.*, 19(7): 995-1000.
- Singh, A.K. 2003b. Effect of growth regulators on growth and flowering in calendula (*Calendula officinalis*). *Indian Perfumer*, 47(3): 275-278.
- Singh, K. 2003. Effect of growth regulators and shoot tip pinching on calendula. J. Orn. Horti., 6(2): 134-136.
- Sujatha, A. Nair, V. Singh and T.V.R.S. Sharma. 2002. Effect of plant growth regulators and quality of gerbera under by island conditions. *Indian J. Hort.*, 59(1): 100-105.
- Taiz, L. and E. Zeiger. 2004. Plant Physiology. (3rd Ed.) p. 720.
- Talukdar, M.C. and L. Paswan. 1988. Effect of GA₃ and CCC on growth and flowering of standard chrysanthemum. *J. Orna. Hort.*, 1: 11-16.
- Talukdar, M.C. and L. Paswan. 1994. Effect of gibberellic acid and cycocel on growth and flowering of chrysanthemum. *Hort. J.*, 7: 141-144.
- Zalewska, M. and M. Antkowiak. 2013. Gibberellic acid effect on growth and flowering of *Ajania pacifica* /nakai/ bremer et Humphries. J. Hort. Res., 21(1): 21-27.

(Received for publication 8 May 2014)