EFFECT OF HIGH-TEMPERATURE, DROUGHT, AND NUTRIENT AVAILABILITY ON MORPHO-PHYSIOLOGICAL AND MOLECULAR MECHANISMS OF RAPESEED - AN OVERVIEW

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Abstract

Production of high yielding cereals and oilseed crops is a major challenge for the farming community to meet the increasing global food demands. In many areas, global food production is challenged by abiotic and biotic factors which adversely hamper the growth of plants. Abiotic stresses, for instance, water, temperature, and essential soil nutrients stress affect the plant's growth. Plants have a different mechanism, which enables them to complete their life cycle under stress conditions, for example, some plants use environmental signals to regulate their morpho-physiological and molecular mechanisms. There is a huge gap of knowledge to identify the genetic mechanisms to mitigate the abiotic stresses. Given the current state of understanding; we have consciously focused on the effect of multiple abiotic stress-signals on morpho-physiological and molecular mechanisms of *Brassica napus*. Further, this review emphasizes the link between different abiotic stresses (high temperature, drought, and low nutrients availability) which ultimately regulate the growth and development of *Brassica napus*.

Key words: Brassica napus L.; Stress; Drought; Temperature; Nutrients.

Introduction

Plants are non-motile organisms that suffer from water, temperature, and nutrients stress during their growth and development (Kauser et al., 2006; Ahmad & Prasad, 2011). Environmental conditions are changing very rapidly, which are creating difficulties for the survival of plants. Extreme environmental conditions (high temperature, drought, and low availability of nutrients) affect the morpho-physiological processes of plants ultimately affect the yield of crops. The agricultural sector is one of the key economic components of many agricultural countries in the world. Abiotic stress has the main impact on crop productivity worldwide, decreasing average yields for major crop plants (Kumar, 2013). Abiotic stress conditions enable the plants to develop defensive mechanisms to survive in harsh conditions, which affect the growth and development by affecting the physiological function of plants (Bray, 2000). These environmental stressful conditions affect the expression of particular genes that are responsive under stress conditions at the molecular level (García et al., 2020). Due to the non-motility plants respond to the outer stressful conditions by changing internal physiological functions, the gene expressive activity, and metabolic processes (Macedo, 2012), plants. Abiotic stresses are creating staggering challenges for biologists to solve the alarming conditions, which will be fatal for future cop production. Therefore, there is a dire need to investigate the effect of multiple abiotic stress conditions on morpho-physiological and molecular mechanisms of plants to cope with future environmental challenges (Macedo, 2012).

One of the major oilseed crops of the world is mustard which mainly includes Brassica napus, Brassica juncea, and Brassica rapa. Among these, Brassica napus L is mainly cultivated worldwide (Raman et al., 2014). It has increasingly become an important agro-economic crop that is widely used for vegetative oil, animal feed, alternative fuel, etc., (Lee et al., 2016; Wang et al., 2018). Canola oil is reported as the third most important plant oil in the world after soybean and palm oil. Its seeds contain 40-50% oil (Chew, 2020). After the extraction of oil from the seed, it contains 38-44% protein which is used for animal nutrition in the form of seed cake. The production of B. napus is badly restricted due to the current situation of global climate change, frequent occurrence of extreme weather, water deficit, extreme temperatures, and other natural disasters (Li et al., 2015). Drought is one of the major abiotic stresses, which is limiting crop growth, development and yield. Drought induces excessive ROS production, which damages cell membranes, plant DNA and proteins). Moreover, drought decreases the photosynthesis and biomass production of plants (Cechin et al., 2007). It was studied that the cultivation of Brassica napus under drought conditions decreased the growth, grain yield and seed quality (Wan et al., 2009). Therefore, there is an increased demand for the development of *B. napus* varieties that have a high yield, quality and resistance to adopt abiotic stresses (Wang et al., 2018). Earlier researches provide pieces of evidence that seed yield can be increased by early planting (Degenhardt & Kondra, 1981; Angadi et al., 2000). But it may be restricted due to spring frosts or excess moisture in fields (Koscielny et al., 2018).

Therefore, there is a dire need to investigate the conditions in which yield and quality of Brassica napus are very sensitive to stress conditions so that the cultivation techniques or new varieties can be developed to maintain the productivity and quality of rapeseed crop. Plants also require nutrients for their normal physiological functions to increase productivity. The nutrients unavailability in the soil is the major determinant of Brassica napus production. Though some progress has been made to increase the production of *Brassica napus*, still there is a milestone to achieve in understanding how its productivity and oil quality is challenged by various environmental cues. Moreover, the morpho-physiological and molecular aspects of Brassica napus are still needed to be known in acclimation to numerous environmental conditions. In the present study, an insight into the various morpho-physiological and molecular aspects of Brassica napus with abiotic stresses has been given. Furthermore, we comprehensively elaborated the interlink between a variety of climatic factors that regulate the growth and development of Brassica napus and its adaptability to harsh environmental conditions.

Water availability: Water is a very important factor that influences crop morpho-physiological processes (Deng et al., 2005; Ashraf et al., 2007; Micheletto et al., 2007). Water transportation between leaf and xylem is carried out due to water potential difference, but drought conditions reduce this potential difference, which disturbs the availability of water and nutrients for cell division and and expansion. The low turgor pressure slow photosynthetic rate under water scarcity reduce leaf size and ultimately limit yield due to low availability of assimilates. Water scarcity decreases germination potential, early seedling growth, root and shoot dry weight, hypocotyl length, and vegetative growth and ultimately decreases the yield of crops (Zeid & Shedeed, 2006). Moreover, water scarcity affects root morphology by decreasing the shoot-to-root mass ratio, increasing the growth of minute and thin roots, and decreasing the number of thick roots (Nosalewicz et al., 2016). The effect of water scarcity depends upon the stress intervals, genotypes, stress severity, and growth phases of Brassica napus. Deficiency of water causes a reduction in cell division and expansion which retard plant height, leaf expansion, number of branches, oil content in seeds, chlorophyll content, and rate of photosynthetic activity of canola cultivars (Moaveni et al., 2010). Drought conditions cause a reduction of yield by decreasing the siliquae number per plant, seed number per siliquae, 1000 seed weight, and oil content (Diepenbrock, 2000).

Among phytohormones, abscisic acid produces during water stress conditions and plays their important role in the stomata closure to cope with water scarcity (Hussain *et al.*, 2010). Closure of stomata decreases the availability of CO_2 for photosynthetically active cells. However, the reduction of CO_2 reduces the assimilates availability for vegetative and reproductive parts of the plants due to decline in photosynthesis, which ultimately reduces *Brassica napus* yield (Reddy *et al.*, 2004; Qaderi *et al.*, 2007; Yang *et al.*, 2009; Khan *et al.*, 2012), which is illustrated in (Fig. 1.) Abscisic acid (ABA) limits the production of ethylene and

maintains plant growth (Yang & Hoffman, 1984; Srinivasarao et al., 2003; Hussain et al., 2010). The molecular studies on drought stress in plants have revealed that the DREB is the most important transcriptional factor as their expression increased by drought stress in the Arabidopsis thaliana (Kudo et al., 2019). Another very recent investigation had introduced the new transcriptional factor i.e. CLE25. They proposed that CLE25 was regulated by ABA (Christmann & Grill, 2018; McLachlan et al., 2018). However, more information is required to find out the exact role of CLE25 in water shortage conditions. Moreover, Hsfs and WRKY gene families are of great interest during stressful conditions. Among Hsfs, the HsfA1a, HsfA1, HsfA2, and HsfA3 are the most important ones (SANMIYA et al., 2020). More specifically, the expression of HsfA3 was upregulated under stress in Arabidopsis thaliana (Yan et al., 2020). Interestingly, HsfA3 has a thermotolerance role during water scarcity and its expression is controlled by DREB2A transcriptional factor. HsfA4 plays their interesting role in overcoming reactive oxygen species (ROS) to help the plant to withstand drought conditions (Zhu et al., 2017). Generally, water stress decreases the grain yield, seed oil content (Alamisaied & Gharineh, 2008), and alters the lipids composition (Ullah et al., 2012) and amino acids (glutamate & glutamine) quantity (Lohaus & Moellers, 2000; Norouzi et al., 2008). Plants physiological processes, for example, leaf development, photosynthesis, and stomatal conductance are negatively correlated with water stress (Gammelvind et al., 1996; Qaderi et al., 2006; Müller et al., 2010). It has been reported in Brassica napus that drought stress-induced the leaf wilting (Balestrini & Vartanian, 1983). Plant growth and development is highly influenced by drought as it decreases photosynthesis and osmolyte biosynthesis (Malinowski & Belesky, 2000), which change assimilates portioning (Andersen et al., 1996). Plants close their stomata during drought to overcome transpiration losses and to maintain water potential. Closure of stomata decreases photosynthesis, which decreases yield (Gao et al., 2018).

It was investigated by several researchers that water deficit conditions decreased plant height, branch number, siliqua per plant, seed per siliqua, 1000-seed weight, seed yield, biological yield, and oil yield (Masoud, 2007; Rad & Zandi, 2012; Naderi & Emam, 2014). A 2.4-fold increase in yield of cultivar Hyola 357 Magnum in response to 30 cm irrigation was recorded with an 8% increase in the oil content. Yield for rain-fed treatment (RF) was reported i.e. 1220 kg/ha in contrast to 2905 k/ha for maximum irrigation (HI i.e. 30 cm) (Pavlista et al., 2016). (Rad & Zandi, 2012) reported that rapeseed cultivar (Hyola 401) produced a yield of 4588 kg ha⁻¹ under normal irrigation while the cultivar RGS003 produced a maximum seed yield of 3577 kg ha⁻¹under water deficit conditions. Moreover, in another study, it was reported that 2138 kg/ha oil yield was obtained in control treatment while stress leads to a reduction of 877, 1141, and 1265 kg/ha after stem elongation, flowering, and pod formation stages in canola, respectively (Fard et al., 2018). Increasing the moisture content after the pollination stage can increase seed and oil yield whereas water deficit stress at the end of the growing

season reduces the percentage of seed oil (Masoud, 2007). The most important component of seed yield in rapeseed is the number of siliquae per plant is (Miller et al., 2003). (Rad & Zandi, 2012) reported a 37% decrease in the number of siliquae per plant when exposed to drought stress whereas a maximum of 325 siliquae per plant was obtained from the normal irrigation. Another study on rapeseed cultivars has concluded that a 59% decrease in the number of siliquae per plant in response to drought stress was observed (Rad & Zandi, 2012). The main determinants of the quality of canola oil are the amount of oleic, linoleic, and erucic acids (Masoud, 2007). Previous studies have reported that the amount of saturated fatty acids of seed oil decreased in response to drought stress (Ahmadi and Bahrani, 2009; Shekari et al., 2016). Some previous studies on the molecular regulation of drought stress in brassica plants have reported that among WRKY family WRKY145, 142, BnaWRKY242, 141, 009, and BrWRKY51, 65, 98, and 104 genes showed their expression during the drought in brassica plants (He et al., 2016). Besides, many studies are conducted to exploring the molecular pathways in regulating the association of different environmental factors. But there is still a huge gap of information in elucidating the molecular regulations of brassica plants in response to drought stress.

How to cope with drought stress: Many strategies are adopted in crop production to enhance water conservation and water use efficiency (WUE). Planting methods were also used to cope with drought stress (Ijaz et al., 2015). There are many important water conservation methods adopted by many farmers like drill sowing, raised bed planting, and furrow planting to ensure the availability of water to seeds during water scarcity (Zhang et al., 2007; Kukal et al., 2010; Aiken et al., 2015). Among plant hormones, plant growth regulators are organic compounds that are also used to promote physiological processes in plants under drought conditions. Plant growth regulators promote, roots, stem, and leaves growth, seeds germination, flowering, and fruit ripening, etc. Exogenous applications of different organic compounds can mitigate the adverse effects of droughts on crop plants (Aown et al., 2012). Application of plant growth regulators such as gibberellic acid, cytokinins and salicylic acid modify plant responses under drought conditions (Farooq et al., 2009). The application of putrescine and salicylic on leaves significantly alleviate the harmful effects of water scarcity on rape (Ullah et al., 2012). Brassica napus plants applied with putrescine and salicylic acid had higher relative water contents, carotene contents, chlorophyll contents, and proline contents under drought conditions as compared to untreated plants (Ullah et al., 2012). During the flowering drought increases the concentration of glucosinolates, which produces pungent oil odors (Bouchereau et al., 1996). Application of putrescine to rape plants under water scarcity conditions the accumulation of glucosinolates decreases concentration. Erucic acid produces pungent odors in rape oil, which can be reduced by the application of salicylic acid (Ullah et al., 2012).



Fig. 1. This figure shows the relationship of drought between physiological function that how drought is responsible for the reduction in photosynthesis which ultimately causes a reduction in yield, (A) drought decrease water potential in tissues which reduce, Phosphoenolpyruvate carboxylase (PEPcase), Fructose 1,6-bisphosphatase (FBPase), Nicotinamide adenine dinucleotide phosphatedependent malic enzyme (NADP-ME), Pyruvate phosphate dikinase (PPDK) and Rubisco enzymatic activities and (B) regulation of non-cyclic electron transport, (C) and triggers ABA signaling, plant respond to close their stomata to prevent water loss, close stomata don't allow entry of CO₂, plant increase reactive oxygen species (ROS). Drought decrease photochemical processes which cause the reduction of ATP synthesis, photosynthesis decline, ultimately cause in decrease crop yield.

Among all other nutrients, potassium acts as a primary osmoticum in maintaining low water potential in plant tissues. Potassium ions (K^+) accumulate in plant tissues under drought conditions to increase water uptake along with a soil-plant gradient (Glenn *et al.*, 1996). Another study strengthens this claim and it has been proved that the application of K fertilizer helps in mitigating the adverse effects of drought on plant growth (Andersen *et al.*, 1992; Sangakkara *et al.*, 2001). Moreover, potassium increases plant resistance against drought through its functions in stomatal regulation, osmoregulation, energy status, charge balance, protein synthesis, and homeostasis (Pandey, 2015).

Specifically, the application of potassium sulphate to B. napus under drought conditions helps in boosting seed yield, seed/siliqua and siliqua/plant (Ardestani & Rad, 2012). An increase in seed yield and dry matter of Indian mustard was reported by using higher amounts of potassium under drought stress. Potassium usage under water deficit conditions can prevent losses in seed yield of rapeseed. Increased use of potassium leads to a maximum number of siliquae per plant (Cheema et al., 2012; Neseim et al., 2014). An application of 200 kg/ha of K₂SO₄ in severe moisture conditions (i.e. irrigation after 80% soil water depletion) and normal moisture conditions can help to increase the yield of rapeseed by 59.95% and 21.48%, respectively (Ardestani & Rad, 2012). (Fanaei et al., 2012) reported a maximum seed yield of 2975 kg/ha by the application of 250 kg/ha K₂SO₄, which was 21% more than the control ones (no potassium application). Furthermore, (Jianwei et al., 2007) recorded a significant increase of 17.5 and 31.7% in grain yield of rapeseed by using 150 and 300 kg/ha K₂O, in comparison to control. Despite having these findings, there is still a big gap of information in elucidating the mechanisms of this interlink between drought and nutrients. More specifically, there is a big challenge for future biologists to find out the molecular signaling pathways in controlling the association between drought and nutrients uptake in brassica.

Effect of high temperature: The international report on the Climate in 2017 states that the earth surface temperature has increased $0.7-0.9^{\circ}$ Celsius $(1.3-1.6^{\circ})$ Fahrenheit) in the hundred years since 1901, but from 1975 temperature increased almost in double 1.5-1.8° Celsius (2.7–3.2° Fahrenheit). Temperature affects plant growth and development by various means. High-temperature stress causes a decrease in oil content, and an increase in protein content and saturated fatty acids. The enzyme activity and respiration rates are increased by a rise in temperature (Aksouh-Harradj et al., 2006; Arcus et al., 2016). High leaf temperature increases the photorespiration rate, which decreases the photosynthetic activity (Long, 1991). Furthermore, an increase in temperature decreases the specificity of rubisco for CO2 vs O2 hence the oxygenation reaction will occur (Ku & Edwards, 1978; Jordan & Ogren, 1984). The solubility of CO₂ rises than O₂ by increasing temperature (Ku & Edwards, 1977). The activation state of rubisco decreases as the leaf temperature rises (Crafts-Brandner & Salvucci, 2000; Yamori & von Caemmerer, 2009). At high temperatures, the production of ATP, NADPH, and photosynthetic electron transport rates decreases, which ultimately decreases photosynthesis (Schrader *et al.*, 2004; Wise *et al.*, 2004). Hence yield of crops decreases by decreasing the photosynthetic rate. The effect of temperature stress on crop plants is illustrated in (Fig. 2). Specifically, the previous study on *Brassica napus* observed the effect of heat stress by using hybrids and inbred with two different planting dates and found that heat stress caused a 20% reduction in seed yield of hybrids and 25% reduction within the inbred (Koscielny *et al.*, 2018). This indicated that heterosis can reduce the effect of high-temperature stress on the yield of canola. (Grant & Beversdorf, 1985) reported an increase in seed yield and decrease in protein content, glucosinolate levels under optimal conditions due to heterosis.

Plant growth and development are affected due to an increase in temperature by global warming, certainly at the early stages of plant growth, which is an emerging constrain in several cultivated areas of the world. High temperatures cause a delay in the normal morphophysiological processes of several plant species and increase the probabilities of yield losses. 28°C is the optimum temperature for germination of Brassica napus and any increase in temperature above this level delays growth and development. Tropical areas have high temperatures, which cause scorching of leaves and twigs, leaves senescence, and discoloration of fruits and leaves (Vollenweider & Günthardt-Goerg, 2005). High temperatures cause an increase in proline and a decrease in chlorophyll content (Gupta et al., 2013; Shah et al., 2015). An increase in temperature above optimum temperature leads to floral sterility and reduces flower number which causes a reduction in yield of economically important Brassica napus species, while the high temperature at the flowering and grain filling stage affects pollen variability, seed development, anthesis, and fertilization process.

Plants respond to heat stress at the transcriptional level by showing the functions of heat stress transcriptional factors (HSFs). HSFA1 is a very important protein that regulates the other HSFs (LIU et al., 2011). Interestingly, *HsfA3* has a thermotolerance role during water scarcity and its expression is controlled by DREB2A transcriptional factor. It means that these two proteins have some connections while contributing to the plant's physiology in response to environmental stimuli (Zhu et al., 2017). Moreover, HSP101 belongs to HSPs and is very helpful in thermotolerance during temperature stress (Su & Li, 2008). There are many gene families in Arabidopsis which play their role in abiotic stress conditions. PIF4 is a gene that belongs to the PIF gene family, shows its expression under high-temperature stress and the TCP5 transcriptional factor holds over the plant thermogenesis by controlling the PIF4 activity (Han et al., 2019). Down-Regulation of the BnWRI1 gene occurs during temperature stress conditions. BnWRI1 showed overexpression under temperature stress to maintains the process of photosynthesis and oil accumulation in rapeseed. Although a lot of studies have been done on the morpho-physiological responses of Brassica napus in the regulation of stress signal perception, however, there is a dire need to elucidate the molecular mechanisms in mediating the association between different abiotic stresses. More specifically, how does HsfA3 has crosstalk with DREB2A in mediating the plant responses under drought and heat stress?

Effect of nutrients: Plants require nutrients for carrying out their normal physiological functions. Major nutrients are NPK, which are very important for plants (Khan *et al.*, 2005). There are some nutrients, which play a very important role in the yield of *Brassica napus*, and these are given below.

Nitrogen is a very vital part of a plant as it is a structural component of chlorophyll. It was investigated that high nitrogen application rates increased the glucosinolate content, but oil content decreased (Rathke *et al.*, 2005; Cheema *et al.*, 2010). There should be the use of a recommended dose of N because it increases the economic yield and minimizes environmental pollution (Aufhammer *et al.*, 1994; Mason & Brennan, 1998; Khan *et al.*, 2005). Higher nitrogen rate increased lodging, decreased yield, changed fatty acid composition, enhanced palmitic and stearic acid content, but linoleic and linolenic acid content decreased, hence poorer oil quality by increasing erucic acid and glucosinolate content (Khan *et al.*, 2018).

After nitrogen, phosphorous is the most important nutrient for plants because it is not easily available for plants due to its low solubility in soil (Raghothama & Karthikeyan, 2005). Phosphorous is a very important and essential element of oil because it is a component of proteins that determining its essentiality for *Brassica napus* (Cheema *et al.*, 2010). Due to the low availability of phosphorous, there is a need to develop suitable conditions that would help Brassica napus plants for carrying out their normal morpho-physiological functions. Brassica napus require high availability of phosphorous for optimum yield. Low availability of phosphorous decreases the yield of Brassica napus (Zhang et al., 2009). Phosphorous is present in excess amounts in the upper layer of soil but its availability is not in access for plants due to its unavailable form. Phosphorous deficiency increases the activity of acid Phosphatase (APase) in the soil (Zhu & Smith, 2001). Nutrients deficiency not only affects the plant physiology but also plays a role at a molecular level in mediating the expression of genes like UBC21 and HTB1 which are very responsive in phosphorous deficiency. BnTrx1;1 and BnPht1;3 show their expression under nutrient deficiency. BnPht1;3 shows their expression especially in roots under phosphate deficiency, however, BnTrx1;1 gene expresses itself under high phosphorous deficiency especially in roots and leaves (Wang et al., 2018). A very recent study has reported that *BnaNACED3* is a very important gene in *B*. napus which shows their expression by the regulation of ABA biosynthesis for growth, development, and stress adaptation (Xu and Cai, 2017). Although this study elucidated the role of BnaNACED3, however, there is still a big gap of information for future scientists to study its effect under the combined effect of nutrient and drought stress.



Fig. 2. This figure shows how temperature stress effective and the response of plants by changing their physiological functions, (A) temperature stress inhibits rubisco activity and reduces CO_2 solubility which decreases carboxylation and enhances photorespiration. (B) decrease, Phosphoenolpyruvate carboxylase (PEPcase), Fructose 1,6-bisphosphatase (FBPase), Nicotinamide adenine dinucleotide phosphate-dependent malic enzyme (NADP-ME), Pyruvate phosphate dikinase (PPDK), and Rubisco enzymatic activities. (C) and increase leaf senescence. (D) reactive oxygen species (ROS) production is a good indication of stress situations, which attack the cell membrane and relate to decreasing the process of ATP synthesis and decarboxylation. (E) Downregulation of non-cyclic electron transport causes a reduction of ATP synthesis which ultimately reduces the photosynthetic activity of crop plants.

Magnesium (Mg), another important nutrient for plant growth (Karley & White, 2009). Mg is a central structural atom of chlorophyll molecule which takes part in enzyme activities and chelation to nucleotidyl phosphate forms (Shaul, 2002). Its deficiency causes an increase in antioxidative mechanisms and increases the sugar levels, which is not effective for photosynthesis (Hermans et al., 2005; Hermans et al., 2010). The growth of young leaves decreased by the deficiency of Mg because it disturbs the distribution of carbon towards leaves than that to roots hence the reduction of root growth is less affected than leaves (Hermans & Verbruggen, 2005; Hermans et al., 2006). Mg deficiency causes more Regulation of genes in leaves than in roots because it usually deals with chlorophyll and its related components (Hermans et al., 2010). Though it is a very well-known nutrient to regulate plant growth and development. Plenty of studies have been done to demonstrate Mg role in plants, however, it is one of the most interesting topics to explain its functions in Brassica napus and how it is involved in making the connection between different abiotic stresses. (Kashem & Kawai, 2007), reported that application of Mg to Brassica napus plants growing under hydroponic conditions suffering from Cd toxicity nutrients solution increased the growth and development by detoxifying the Cd toxicity in plants especially shoot.

Another nutrient, cadmium (Cd) is absorbed by roots through plasma membrane-like as iron (Fe), calcium (Ca), and zinc (Zn) (Clemens, 2006; Hermans et al., 2010; Lux et al., 2010). Cd reacts with sulphydryl groups and becomes a source of toxicity. Cd can replace the position of Mg in chlorophyll structure which damage the photosynthetic structure by stopping the function of photosystem II (Faller *et al.*, 2005) or by disturbing the biosynthesis of chlorophyll (Gadallah, 1994), arrangement and construction of light-harvesting complexes (Janik et al., 2010) and chloroplast organization and thickness (Baryla et al., 2001; Carrier et al., 2003). However, this phenomenon still needs to be studied at the molecular level that how do Cd affects the efficiency of chlorophyll? And what are the key genes that are involved in the molecular regulation of the chlorophyll? Higher Cd availability in the growth medium can stop the uptake of Fe in many species (Kovács et al., 2006). Deficiency of Fe occurred due to the Cd introduction, which leads to the development of chlorosis (Kramer et al., 1991; Su & Li, 2008). Cd is one of the most important heavy metals and its increasing toxicity in the world is hazardous for human health. The stress of Cd is harmful to normal morphophysiological functions in plants especially for brassica crop plants because its stress reduces growth, development, and oil content (Ahmad et al., 2015). Brassica spp. is very famous in the world as a phytoremediator because it can accumulate heavy metals (Mourato et al., 2015).

Drought and nutrients uptake: Nutrient uptake is one of the important processes for the proper growth and development of plants. Plants usually uptake nutrients under the presence of water, water scarcity can affect the

nutrients uptake. Water scarcity greatly affects the uptake of essential nutrients. Roots uptake many nutrients under the presence of adequate water like nitrogen, silicon, magnesium, and calcium, drought limits the movement of these nutrients by affecting the process of diffusion and mass flow and ultimately retard the plant growth (Barber, 1995). Drought conditions change the root morphology, for instance, root length and surface area, these alterations in root architecture are helpful to plants for capturing low mobile nutrients (Lynch & Brown, 2001). The deficiency of water retards the growth of plant roots which is a hurdle to plants to uptake low mobile nutrients like phosphorous (Garg, 2003). Microbial activities and their interaction with plant roots play a very important role in nutrients uptake from soil to plants. Drought conditions negatively affect the microbial activities hence reduce the nutrient supplying ability of microbes for plants (Schimel et al., 2007). Different crop species behave differently under water scarcity conditions. Usually, N uptake is enhanced, P uptake is decreased and K uptake remained unaffected under water deficit conditions. Although the relationship among the nutrients is very complicated due to the interactive effect on each other and overall plant physiology. This particular aspect requires more research at a sophisticated molecular level.

Future perspectives: Agriculture is one of the most important economic sectors, it is the economic backbone of all the world and its sustainability depends upon the potent nutrients availability and suitable environmental conditions. The evolution of plants has been influenced by various harsh environmental conditions, which include a shortage of water, extremes of temperature, unavailability of nutrients in the soil profile. As a result of these severe environmental conditions plants have acquired mechanisms to sustain stability among growth, development, and reproduction.

There is a dire need to discuss conditions in which the yield and quality of *Brassica napus* are very sensitive to external factors especially, under stress conditions so that the productivity and quality of this crop could be maintained. Moreover, plants require nutrients for carrying out their normal physiological functions. The nutrients availability in soil is the major determinant of crop productivity of *Brassica napus*. Though many signs of progress have been done to increase the production of *Brassica napus*, still there is a milestone to achieve in understanding how its productivity and oil quality is challenged by various environmental cues.

The morpho-physiological and molecular aspects of *Brassica napus* are still needed to be known in acclimation to numerous hazardous environmental conditions. In the present study, an insight into the various morpho-physiological and molecular aspects of *Brassica napus* with abiotic stresses has been given as illustrated in (Fig. 3) which depicts the questions for future scientists to investigate the multiple abiotic stress factors. However, there is a huge gap of information in elucidating the molecular regulations of *Brassica napus* in response to drought stress especially the newly founded gene *CLE25* (Fig. 3C). How this gene does regulate the molecular regulations of *Brassica napus*? More specifically, how does *HsfA3* has crosstalk with *DREB2A* in mediating the plant responses under drought and heat stress (Fig. 3A)? In context to nutrient availability and their interaction with other abiotic stresses, there would be plenty of questions to be asked for future scientists. For instance, how does Cd affect the efficiency of chlorophyll, when it acts with magnesium in *Brassica napus* (Fig. 3B)? What are the key regulators of this action at the

molecular level? It is discussed that potassium could be helpful to mitigate drought, whereas there is a question arises that how does the potassium interlink with ABA to regulate the phenomenal change in *Brassica napus*? What are the inducible genes and transcriptional factors that are involved in this regulation? Broadly, it one of the most fascinating topics for future studies to investigate the combined effect of abiotic stresses, and to find out the link between multiple environmental stresses in *Brassica napus*.



Fig. 3. This figure shows the relationship of multiple factors on Morpho-Physiological processes of plants, (A) shows how temperature and drought coordinate with each other in terms of HSFs and DREB interaction. (B) what is the key role of Potassium (K) under drought conditions, the role of nutrients on physio-chemical changes in plants? (C) This shows that, how does CLE25 activate under drought stress, and what is the exact role of this gene?

References

- Ahmad, P. and M.N.V. Prasad. 2011. Abiotic stress responses in plants: Metabolism, productivity and sustainability. *Springer Science & Business Media*.
- Ahmad, P., M. Sarwat, N.A. Bhat, M.R. Wani, A.G. Kazi and L.S.P. Tran. 2015. Alleviation of cadmium toxicity in *Brassica juncea* L. (czern. & coss.) by calcium application involves various physiological and biochemical strategies. *PloS One*, 10(1): e0114571.
- Ahmadi, M. and M. Bahrani. 2009. Yield and yield components of rapeseed as influenced by water stress at different growth stages and nitrogen levels. J. Agri. Environ. Sci., 5(6): 755-761.
- Aiken, R., D. Baltensperger, J. Krall, A. Pavlista and J. Johnson. 2015. Planting methods affect emergence, flowering and yield of spring oilseed crops in the us central high plains. *Ind. Crops Prod.*, 69: 273-277.
- Aksouh-Harradj, N., L. Campbell and R. Mailer. 2006. Canola response to high and moderately high temperature stresses during seed maturation. *Can. J. Plant Sci.*, 86(4): 967-980.
- Alamisaied, K. and M. Gharineh. 2008. Optimization of irrigation and nitrogen regimes for rapeseed production under drought stress. J. Agron., 7(4): 321-326.
- Andersen, M.N., C. Jensen and R. Lösch. 1992. The interaction effects of potassium and drought in field-grown barley. I. Yield, water-use efficiency and growth. *Acta Agr Scand B-S P.*, 42(1): 34-44.

- Andersen, M.N., T. Heidmann and F. Plauborg. 1996. The effects of drought and nitrogen on light interception, growth and yield of winter oilseed rape. ACTA AGR SCAND B-S P. 46(1): 55-67.
- Angadi, S., H. Cutforth, P. Miller, B. McConkey, M. Entz, S. Brandt and K. Volkmar. 2000. Response of three brassica species to high temperature stress during reproductive growth. *Can. J. Plant Sci.*, 80(4): 693-701.
- Aown, M., S. Raza, M. Saleem, S. Anjum, T. Khaliq and M. Wahid. 2012. Foliar application of potassium under water deficit conditions improved the growth and yield of wheat (*Triticum aestivum L.*). J. Ani. Plant Sci., 22(2): 431-437.
- Arcus, V.L., E.J. Prentice, J.K. Hobbs, A.J. Mulholland, M.W. Van der Kamp, C.R. Pudney, E.J. Parker and L.A. Schipper. 2016. On the temperature dependence of enzyme-catalyzed rates. *Biochem.*, 55(12): 1681-1688.
- Ardestani, G. and A.H.S. Rad. 2012. Impact of regulated deficit irrigation on the physiological characteristics of two rapeseed varieties as affected by different potassium rates. *Afr. J. Biotechnol.*, 11(24): 6510-6519.
- Ashraf, M., S. Nawazish and H.U.R. Athar. 2007. Are chlorophyll fluorescence and photosynthetic capacity potential physiological determinants of drought tolerance in maize (*Zea mays L.*). *Pak. J. Bot.*, 39(4): 1123-1131.
- Aufhammer, W., E. Kübler and M. Bury. 1994. Nitrogen uptake and nitrogen residuals of winter oil-seed rape and fallout rape. *Agron Crop Sci.*, 172(4): 255-264.
- Balestrini, S. and N. Vartanian. 1983. Rhizogenic activity during water stress-induced senescence in brassica napus var. Oleifera [rape]. *Physiol. Veg.*, 21(2): 269-277.

- Barber, S.A. 1995. Soil nutrient bioavailability: A mechanistic approach. John Wiley & Sons.
- Baryla, A., P. Carrier, F. Franck, C. Coulomb, C. Sahut and M. Havaux. 2001. Leaf chlorosis in oilseed rape plants (*Brassica napus*) grown on cadmium-polluted soil: Causes and consequences for photosynthesis and growth. *Planta*, 212(5-6): 696-709.
- Bouchereau, A., N. Clossais-Besnard, A. Bensaoud, L. Leport and Renard. 1996. Water stress effects on rapeseed quality. *Eur. J. Agron.*, 5(1-2): 19-30.
- Bray, E.A. 2000. Response to abiotic stress. *Biochem. & Mol. Biol. Plants*, 1158-1203.
- Carrier, P., A. Baryla and M. Havaux. 2003. Cadmium distribution and microlocalization in oilseed rape (*Brassica napus*) after long-term growth on cadmium-contaminated soil. *Planta*, 216(6): 939-950.
- Cechin, I., T.D.F. Fumis and A.L. Dokkedal. 2007. Growth and physiological responses of sunflower plants exposed to ultraviolet-b radiation. *Ciên. Rurl.*, 37(1): 85-90.
- Cheema, M., M. Saleem, N. Muhammad, M. Wahid and B. Baber. 2010. Impact of rate and timing of nitrogen application on yield and quality of canola (*Brassica napus* L.). *Pak. J. Bot.*, 42(3): 1723-1731.
- Cheema, M.A., M. Wahid, A. Sattar, F. Rasul and M. Saleem. 2012. Influence of different levels of potassium on growth, yield and quality of canola (*Brassica napus L.*) cultivars. *Pak. J. Agri. Sci.*, 49(2): 163-168.
- Chew, S.C. 2020. Cold-pressed rapeseed (*Brassica napus*) oil: Chemistry and functionality. *Food Res. Int.*, (131): 108997
- Christmann, A. and E. Grill. 2018. Peptide signal alerts plants to drought. *Nature Publishing Group*.
- Clemens, S. 2006. Toxic metal accumulation, responses to exposure and mechanisms of tolerance in plants. *Biochimie*, 88(11): 1707-1719.
- Crafts-Brandner, S.J. and M.E. Salvucci. 2000. Rubisco activase constrains the photosynthetic potential of leaves at high temperature and co2. *Proc. Natl. Acad. Sci.*, 97(24): 13430-13435.
- Degenhardt, D. and Z. Kondra. 1981. The influence of seeding date and seeding rate on seed yield and yield components of five genotypes of brassica napus. *Can. J. Plant Sci.*, 61(2): 175-183.
- Deng, X.P., L. Shan, S. Inanaga and M. Inoue. 2005. Watersaving approaches for improving wheat production. J. Sci. Food Agric., 85(8): 1379-1388.
- Diepenbrock, W. 2000. Yield analysis of winter oilseed rape (brassica napus l.): A review. *Field Crops Res.*, 67(1): 35-49.
- Faller, P., K. Kienzler and A. Krieger-Liszkay. 2005. Mechanism of cd2+ toxicity: Cd2+ inhibits photoactivation of photosystem ii by competitive binding to the essential ca2+ site. *Biochem. Biophys. Acta*, 1706(1-2): 158-164.
- Fanaei, H., M. Galavi, M. Kafi and A. Ghanbari Bonjar. 2012. Amelioration of water stress by potassium fertilizer in two oilseed species. *Int. J. Plant Prod.*, 3(2): 41-54.
- Fard, N.S., H.H.S. Abad, A.S. Rad, E.M. Heravan and J. Daneshian. 2018. Effect of drought stress on qualitative characteristics of canola cultivars in winter cultivation. *Ind. Crops. Prod.*, 114: 87-92.
- Farooq, M., A. Wahid and D.J. Lee. 2009. Exogenously applied polyamines increase drought tolerance of rice by improving leaf water status, photosynthesis and membrane properties. *Acta Physiol. Plant.*, 31(5): 937-945.
- Gadallah, M. 1994. Interactive effect of heavy metals and temperature on the growth, and chlorophyll, saccharides and soluble nitrogen contents inphaseolus plants. *Biol. Plant.*, 36(3): 373.
- Gammelvind, L., J. Schjoerring, V. Mogensen, C. Jensen and J. Bock. 1996. Photosynthesis in leaves and siliques of winter oilseed rape (*Brassica napus* L.). *Plant Soil.*, 186(2): 227-236.

- Gao, L., C.D. Caldwell and Y. Jiang. 2018. Photosynthesis and growth of camelina and canola in response to water deficit and applied nitrogen. *Crop Sci.*, 58(1): 393-401.
- García, P., L. Filippis, A. Gul, M. Hasanuzzaman, M. Ozturk, V. Altay and M.T. Lao. 2020. Oxidative Stress and Antioxidant Metabolism under Adverse Environmental Conditions: a Review. *The Botanical Review*. 1: 1-46.
- Garg, B. 2003. Nutrient uptake and management under drought: Nutrient-moisture interaction. *Curr. Agric.*, 27(1/2): 1-8.
- Glenn, E., R. Pfister, J.J. Brown, T.L. Thompson and J. O'Leary. 1996. Na and k accumulation and salt tolerance of *Atriplex canescens* (Chenopodiaceae) genotypes. *Amer. J. Bot.*, 83(8): 997-1005.
- Grant, I. and W.D. Beversdorf. 1985. Agronomic performance of triazine-resistant single-cross hybrid oilseed rape (*Brassica napus* L.). Can. J. Plant Sci., 65(4): 889-892.
- Gupta, N., S. Agarwal, V. Agarwal, N. Nathawat, S. Gupta and G. Singh. 2013. Effect of short-term heat stress on growth, physiology and antioxidative defence system in wheat seedlings. *Acta Physiol. Plant.*, 35(6): 1837-1842.
- Han, X., H. Yu, R. Yuan, Y. Yang, F. An and G. Qin. 2019. Arabidopsis transcription factor tcp5 controls plant thermomorphogenesis by positively regulating pif4 activity. *Sci.*, 15: 611-622.
- He, Y., S. Mao, Y. Gao, L. Zhu, D. Wu, Y. Cui, J. Li and W. Qian. 2016. Genome-wide identification and expression analysis of wrky transcription factors under multiple stresses in *Brassica napus*. *PLoS One*, 11(6): e0157558.
- Hermans, C. and N. Verbruggen. 2005. Physiological characterization of mg deficiency in arabidopsis thaliana. J *Exp. Bot.*, 56(418): 2153-2161.
- Hermans, C., F. Bourgis, M. Faucher, R.J. Strasser, S. Delrot and N. Verbruggen. 2005. Magnesium deficiency in sugar beets alters sugar partitioning and phloem loading in young mature leaves. *Planta*, 220(4): 541-549.
- Hermans, C., J.P. Hammond, P.J. White and N. Verbruggen. 2006. How do plants respond to nutrient shortage by biomass allocation? *Trends Plant Sci.*, 11(12): 610-617.
- Hermans, C., M. Vuylsteke, F. Coppens, A. Craciun, D. Inzé and N. Verbruggen. 2010. Early transcriptomic changes induced by magnesium deficiency in arabidopsis thaliana reveal the alteration of circadian clock gene expression in roots and the triggering of abscisic acid-responsive genes. *New Phytol.*, 187(1): 119-131.
- Hermans, C., M. Vuylsteke, F. Coppens, S.M. Cristescu, F.J. Harren, D. Inzé and N. Verbruggen. 2010. Systems analysis of the responses to long-term magnesium deficiency and restoration in arabidopsis thaliana. *New Phytol.*, 187(1): 132-144.
- Hussain, S., M. Saleem, M. Ashraf, M. Cheema and M. Haq. 2010. Abscisic acid, a stress hormone helps in improving water relations and yield of sunflower (*Helianthus annuus* L.) hybrids under drought. *Pak. J. Bot.*, 42(3): 2177-2189.
- Ijaz, M., M.A.S. Raza, S. Ali, K. Ghazi, T.A. Yasir, M. Saqib and M. Naeem. 2015. Differential planting density influences growth and yield of hybrid maize (*Zea mays L.*). *J. Agri. & Environ. Sci.*, 2(3): 1-5.
- Janik, E., W. Maksymiec, R. Mazur, M. Garstka and W.I. Gruszecki. 2010. Structural and functional modifications of the major light-harvesting complex ii in cadmium-or copper-treated secale cereale. *Plant Cell Physiol.*, 51(8): 1330-1340.
- Jianwei, L., C. Fang and W.B. Garden. 2007. Effect of phosphorus and potassium application on rapeseed yield and nutrients use efficiency. In: (The 12th International Rapeseed Congress). Huazhong Agricultural University: pp: 1029-1032.

- Jordan, D.B. and W.L. Ogren. 1984. The co 2/o 2 specificity of ribulose 1, 5-bisphosphate carboxylase/oxygenase. *Planta.*, 161(4): 308-313.
- Karley, A.J. and P.J. White. 2009. Moving cationic minerals to edible tissues: Potassium, magnesium, calcium. *Curr. Opin. Plant Biol.*, 12(3): 291-298.
- Kashem, M.A. and S. Kawai. 2007. Alleviation of cadmium phytotoxicity by magnesium in japanese mustard spinach. *Soil Sci. Plant Nutr.*, 53(3): 246-251.
- Kauser, R., H.U.R. Athar and M. Ashraf. 2006. Chlorophyll fluorescence: A potential indicator for rapid assessment of water stress tolerance in canola (*Brassica napus* L.). *Pak. J. Bot.*, 38(5 Spec. Iss.): 1501-1509.
- Khan, I.A., A. Khatri, G.S. Nizamani, M.A. Siddiqui, S. Raza and N.A. Dahar. 2005. Effect of npk fertilizers on the growth of sugarcane clone aec86-347 developed at nia, tando jam, pakistan. *Pak. J. Bot.*, 37(2): 355.
- Khan, S., S. Anwar, J. Kuai, A. Noman, M. Shahid, M. Din, A. Ali and G. Zhou. 2018. Alteration in yield and oil quality traits of winter rapeseed by lodging at different planting density and nitrogen rates. *Sci. Rep.*, 8(1): 634.
- Khan, S.U., B. Asghari and A. Gurmani. 2012. Abscisic acid and salicylic acid seed treatment as potent inducer of drought tolerance in wheat (*Triticum aestivum* L.). *Pak. J. Bot.*, 44(1): 43-49.
- Koscielny, C., S. Gardner and R. Duncan. 2018. Impact of high temperature on heterosis and general combining ability in spring canola (*Brassica napus* L.). *Field Crops Res.*, 221: 61-70.
- Kovács, K., E. Kuzmann, F. Fodor, A. Vértes and A. Kamnev. 2006. Mössbauer study of iron uptake in cucumber root. In: Icame 2005. *Springer*. pp: 289-294.
- Kramer, G.F., H.A. Norman, D.T. Krizek and R.M. Mirecki. 1991. Influence of uv-b radiation on polyamines, lipid peroxidation and membrane lipids in cucumber. *Phytochemistry*, 30(7): 2101-2108.
- Ku, S.B. and G.E. Edwards. 1977. Oxygen inhibition of photosynthesis: Ii. Kinetic characteristics as affected by temperature. *Plant Physiol.*, 59(5): 991-999.
- Ku, S.B. and G.E. Edwards. 1978. Oxygen inhibition of photosynthesis. *Planta*, 140(1): 1-6.
- Kudo, M., S. Kidokoro, T. Yoshida, J. Mizoi, M. Kojima, Y. Takebayashi, H. Sakakibara, A.R. Fernie, K. Shinozaki and K. Yamaguchi-Shinozaki. 2019. A gene-stacking approach to overcome the trade-off between drought stress tolerance and growth in arabidopsis. *Plant J.*, 97(2): 240-256.
- Kukal, S., E. Humphreys, S. Thaman, B. Singh and J. Timsina. 2010. Factors affecting irrigation water savings in raised beds in rice and wheat. *Field Crops Res.*, 118(1): 43-50.
- Kumar, M., 2013. Crop plants and abiotic stresses. J. Biomol. Res. Ther., 3(1):
- Lee, B.R., Q. Zhang, D.W. Bae and T.H. Kim. 2016. Pod removal responsive change in phytohormones and its impact on protein degradation and amino acid transport in source leaves of brassica napus. *Plant Physiol. Biochem.*, 106: 159-164.
- Li, Q., M. Yin, Y. Li, C. Fan, Q. Yang, J. Wu, C. Zhang, H. Wang and Y. Zhou. 2015. Expression of brassica napus ttg2, a regulator of trichome development, increases plant sensitivity to salt stress by suppressing the expression of auxin biosynthesis genes. J. Exp. Bot., 66(19): 5821-5836.
- Liu, H.C., H.T. Liao and Y.Y. Charng. 2011. The role of class al heat shock factors (hsfa1s) in response to heat and other stresses in arabidopsis. *Plant Cell Environ*. 34(5): 738-751.
- Lohaus, G. and C. Moellers. 2000. Phloem transport of amino acids in two *Brassica napus* L. Genotypes and one b. Carinata genotype in relation to their seed protein content. *Planta*, 211(6): 833-840.

- Long, S.P. 1991. Modification of the response of photosynthetic productivity to rising temperature by atmospheric co2 concentrations: Has its importance been underestimated? *Plant Cell Environ.*, 14(8): 729-739.
- Lux, A., M. Martinka, M. Vaculík and P.J. White. 2010. Root responses to cadmium in the rhizosphere: A review. J. Exp. Bot., 62(1): 21-37.
- Lynch, J.P. and K.M. Brown. 2001. Topsoil foraging–an architectural adaptation of plants to low phosphorus availability. *Plant Soil.*, 237(2): 225-237.
- Macedo, A.F. 2012. Abiotic stress responses in plants: Metabolism to productivity. In: Abiotic stress responses in plants. *Springer*. pp: 41-61.
- Malinowski, D.P. and D.P. Belesky. 2000. Adaptations of endophyte-infected cool-season grasses to environmental stresses: Mechanisms of drought and mineral stress tolerance. *Crop Sci.*, 40(4): 923-940.
- Mason, M. and R. Brennan. 1998. Comparison of growth response and nitrogen uptake by canola and wheat following application of nitrogen fertilizer. J. Plant Nutr., 21(7): 1483-1499.
- Masoud, S.M. 2007. The effects of water deficit during growth stages of canola (*Brassica napus* L.). AEJAES. 2: 417-422.
- McLachlan, D.H., A.J. Pridgeon and A.M. Hetherington. 2018. How arabidopsis talks to itself about its water supply. *Mol. Cell.*, 70(6): 991-992.
- Micheletto, S., L. Rodriguez-Uribe, R. Hernandez, R.D. Richins, J. Curry and M.A. O'Connell. 2007. Comparative transcript profiling in roots of phaseolus acutifolius and p. Vulgaris under water deficit stress. *Plant Sci.*, 173(5): 510-520.
- Miller, P., S. Angadi, G. Androsoff, B. McConkey, C. McDonald, S. Brandt, H. Cutforth, M. Entz and K. Volkmar. 2003. Comparing brassica oilseed crop productivity under contrasting n fertility regimes in the semiarid northern great plains. *Can. J. Plant Sci.*, 83(3): 489-497.
- Moaveni, P., A. Ebrahimi and H.A. Farahani. 2010. Studying of oil yield variations in winter rapeseed (*Brassica napus* L.) cultivars under drought stress conditions. J. Agri. Biotech. Sustain. Dev., 2(5): 71-75.
- Mourato, M., I. Moreira, I. Leitão, F. Pinto, J. Sales and L. Martins. 2015. Effect of heavy metals in plants of the genus brassica. *Int. J. Mol. Sci.*, 16(8): 17975-17998.
- Müller, T., D. Lüttschwager and P. Lentzsch. 2010. Recovery from drought stress at the shooting stage in oilseed rape (*Brassica napus*). J. Agron. Crop Sci., 196(2): 81-89.
- Naderi, R. and Y. Emam. 2014. Evaluation of rapeseed (*Brassica napus* L.) cultivars performance under drought stress. *Aust. J. Crop Sci.*, 8(9): 1319-1323.
- Neseim, M., A. Amin and M. El-Mohammady. 2014. Effect of potassium applied with foliar spray of yeast on sugar beet growth and yield under drought stress. *GARJAS*, 3(8): 211-222.
- Norouzi, M., M. Toorchi, G.H. Salekdeh, S. Mohammadi, M. Neyshabouri and S. Aharizad. 2008. Effect of water deficit on growth, grain yield and osmotic adjustment in rapeseed. *J. Food Agric. Environ.*, 6(2): 312.
- Nosalewicz, A., J. Siecińska, M. Śmiech, M. Nosalewicz, D. Wiącek, A. Pecio and D. Wach. 2016. Transgenerational effects of temporal drought stress on spring barley morphology and functioning. *Environ. Exp. Bot.*, 131: 120-127.
- Pandey, R. 2015. Mineral nutrition of plants. In: Plant biology and biotechnology. *Springer*. pp: 499-538.
- Pavlista, A., G. Hergert, J. Margheim and T. Isbell. 2016. Growth of spring canola (*Brassica napus*) under deficit irrigation in western nebraska. *Ind. Crops Prod.*, 83: 635-640.
- Qaderi, M.M., D.M. Reid and E.C. Yeung. 2007. Morphological and physiological responses of canola (*Brassica napus*) siliquas and seeds to uvb and co2 under controlled environment conditions. *Environ. Exp. Bot.*, 60(3): 428-437.

- Qaderi, M.M., L.V. Kurepin and D.M. Reid. 2006. Growth and physiological responses of canola (*Brassica napus*) to three components of global climate change: Temperature, carbon dioxide and drought. *Physiol. Plant.* 128(4): 710-721.
- Rad, A.H.S. and P. Zandi. 2012. The effect of drought stress on qualitative and quantitative traits of spring rapeseed (*Brassica* napus L.) cultivars. Zemdirbyste-Agriculture, 99: 47-54.
- Raghothama, K. and A. Karthikeyan. 2005. Phosphate acquisition. *Plant Soil.*, 274(1-2): 37.
- Raman, H., J. Dalton-Morgan, S. Diffey, R. Raman, S. Alamery, D. Edwards and J. Batley. 2014. Snp markers-based map construction and genome-wide linkage analysis in brassica napus. *Plant Biotechnol. J.*, 12(7): 851-860.
- Rathke, G.-W., O. Christen and W. Diepenbrock. 2005. Effects of nitrogen source and rate on productivity and quality of winter oilseed rape (*Brassica napus* L.) grown in different crop rotations. *Field Crops Res.*, 94(2-3): 103-113.
- Reddy, A.R., K.V. Chaitanya and M. Vivekanandan. 2004. Drought-induced responses of photosynthesis and antioxidant metabolism in higher plants. J. Plant Physiol., 161(11): 1189-1202.
- Sangakkara, U., M. Frehner and J. Nösberger. 2001. Influence of soil moisture and fertilizer potassium on the vegetative growth of mungbean (*Vigna radiata* L. Wilczek) and cowpea (*Vigna unguiculata* L. Walp). J. Agron. Crop Sci., 186(2): 73-81.
- Sanmiya, K.K., Y. Koja and A. Iguchi. 2020. The Cloning of cDNAs for the Heat-shock Transcription Factors HSFA1, HSFA2 and HSFA3 from Tobacco. *Trop. Agric.*, 64(1): 34-40.
- Schimel, J., T.C. Balser and M. Wallenstein. 2007. Microbial stress-response physiology and its implications for ecosystem function. *Ecol.*, 88(6): 1386-1394.
- Schrader, S., R. Wise, W. Wacholtz, D.R. Ort and T. Sharkey. 2004. Thylakoid membrane responses to moderately high leaf temperature in pima cotton. *Plant Cell Environ.*, 27(6): 725-735.
- Shah, S.H., S. Ali, S.A. Jan and G.M. Ali. 2015. Piercing and incubation method of in planta transformation producing stable transgenic plants by overexpressing dreb1a gene in tomato (*Solanum lycopersicum Mill.*). *PCTOC.*, 120(3): 1139-1157.
- Shaul, O. 2002. Magnesium transport and function in plants: The tip of the iceberg. *Biometals*, 15(3): 307-321.
- Shekari, F., V. Soltaniband, A. Javanmard and A. Abbasi. 2016. The impact of drought stress at different stages of development on water relations, stomatal density and quality changes of rapeseed (*Brassica napus* L.). *Iran Agri. Res.*, 34(2): 81-90.
- Srinivasarao, C., M. Ali, A. Ganeshamurthy and K. Singh. 2003. Potassium requirements of pulse crops. *BCI*., 17(1): 8.
- Su, P.H. and H.M. Li. 2008. Arabidopsis stromal 70-kd heat shock proteins are essential for plant development and important for thermotolerance of germinating seeds. *Plant Physiol.*, 146(3): 1231-1241.
- Ullah, F., A. Bano and A. Nosheen. 2012. Effects of plant growth regulators on growth and oil quality of canola

(Brassica napus L.) under drought stress. Pak. J. Bot., 44(6): 1873-1880.

- Vollenweider, P. and M.S. Günthardt-Goerg. 2005. Diagnosis of abiotic and biotic stress factors using the visible symptoms in foliage. *Environ. Pollut.*, 137(3): 455-465.
- Wan, J., R. Griffiths, J. Ying, P. McCourt and Y. Huang. 2009. Development of drought-tolerant canola (*Brassica napus* L.) through genetic modulation of aba-mediated stomatal responses. *Crop Sci.*, 49(5): 1539-1554.
- Wang, Z., C. Yang, H. Chen, P. Wang, P. Wang, C. Song, X. Zhang and D. Wang. 2018. Multi-gene co-expression can improve comprehensive resistance to multiple abiotic stresses in *Brassica napus* L. *Plant Sci.*, 274: 410-419.
- Wise, R., A. Olson, S. Schrader and T. Sharkey. 2004. Electron transport is the functional limitation of photosynthesis in field grown pima cotton plants at high temperature. *Plant Cell Environ.*, 27(6): 717-724.
- Xu, P. and W. Cai. 2017. Functional characterization of the bnnced3 gene in brassica napus. *Plant Sci.*, 256: 16-24.
- Yamori, W. and S. von Caemmerer. 2009. Effect of rubisco activase deficiency on the temperature response of co2 assimilation rate and rubisco activation state: Insights from transgenic tobacco with reduced amounts of rubisco activase. *Plant Physiol.*, 151(4): 2073-2082.
- Yan, Q.Z., F. Wu, J. Li, T. Ma, Y. Zhao, Q. Ma, P. Wang, Y. Wang and J. Zhang. 2020. Integrated analysis of coexpression, conserved genes and gene families reveal core regulatory network of heat stress response in *Cleistogenes songorica*, a xerophyte perennial desert plant. *BMC Genom.*, 21(1): 1-18.
- Yang, C.W., H.H. Xu, L.L. Wang, J. Liu, D.C. Shi and D.L. Wang. 2009. Comparative effects of salt-stress and alkalistress on the growth, photosynthesis, solute accumulation, and ion balance of barley plants. *Photosyn.*, 47(1): 79-86.
- Yang, S.F. and N.E. Hoffman. 1984. Ethylene biosynthesis and its regulation in higher plants. Ann. Rev. Plant Physiol., 35(1): 155-189.
- Zeid, I. and Z. Shedeed. 2006. Response of alfalfa to putrescine treatment under drought stress. *Biol. Plant.*, 50(4): 635.
- Zhang, H., Y. Huang, X. Ye, L. Shi and F. Xu. 2009. Genotypic differences in phosphorus acquisition and the rhizosphere properties of brassica napus in response to low phosphorus stress. *Plant Soil.*, 320(1-2): 91-102.
- Zhang, J., J. Sun, A. Duan, J. Wang, X. Shen and X. Liu. 2007. Effects of different planting patterns on water use and yield performance of winter wheat in the huang-huai-hai plain of china. *Agri. Water Manag.*, 92(1-2): 41-47.
- Zhu, X., C. Huang, L. Zhang, H. Liu, J. Yu, Z. Hu and W. Hua. 2017. Systematic analysis of hsf family genes in the *Brassica napus* genome reveals novel responses to heat, drought and high co2 stresses. *Front. Plant Sci.*, 8: 1174.
- Zhu, Y. and S. Smith. 2001. Seed phosphorus (p) content affects growth, and p uptake of wheat plants and their association with arbuscular mycorrhizal (am) fungi. *Plant Soil*, 231(1): 105-112.

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