

PHOTOSYNTHETIC SYSTEM OF *LEPTOCHLOA FUSCA* (L.) KUNTH.

YUSUF ZAFAR AND KAUSER A. MALIK

*Soil Biology Division,
Nuclear Institute for Agriculture and Biology, Faisalabad, Pakistan.*

Abstract

Photosynthetic system of *Leptochloa fusca* (L.) Kunth., was determined by observing its leaf anatomy, distribution of starch grains, measurement of CO₂-compensation point and ¹³C/¹²C ratios. All diagnostic characters revealed that this grass has a C-4 pathway of photosynthesis.

Introduction

It has been known that many grasses possess the C-4 dicarboxylic acid pathway of photosynthesis instead of reductive pentose pathway (C-3) found generally in plants (Kortschak *et al.*, 1965; Hatch & Slack, 1970). Grasses possessing C-4 pathway utilize their available nitrogen more efficiently in producing biomass than grasses having C-3 pathway (Black *et al.*, 1978). The fact that all the grasses in which associative N₂-fixation has been reported except rice and wheat possess C-4 pathway, lead Dobreiner *et al.*, (1972) to propose a relationship between C-4 photosynthetic pathway and dinitrogen fixation in the root zones of the plant. Malik *et al.*, (1980, 1981) has already reported nitrogenase activity based on acetylene reduction assays (ARA) in the excised roots of Kallar grass (*Leptochloa fusca* (L.) Kunth.), growing in saline areas. Present investigations were carried out to know the photosynthetic system of this grass.

Materials and Methods

Collection: Plant material of *Leptochloa fusca* (L.) Kunth. (Synonym *Diplachne fusca* (L.) Beauv.) commonly known as Kallar grass was collected from field of NIAB campus, Faisalabad.

Leaf anatomy: Free hand sections of leaves of Kallar grass were placed in 40% commercial sodium hypochlorite solution (v/v) in water for one hour in order to decolourise the plant material. Water mounts were made and examined under the Microlux-11 photomicroscope. Distribution of starch grains in the parenchymatous cells was determined according to the method of Downton & Tregunna (1968). For staining purpose, leaves were collected in the afternoon of a sunny day. Sections were dipped for one hour in small vial containing Gram's iodine (Iodine-1 gm, KI-2 gm, distl. H₂O, 100 ml). Sections were examined under microscope for the distribution of stained starch grains and photographed.

CO₂-compensation point: CO₂-compensation point for Kallar grass was measured by an infra red gas analyser (IRGA-120). A quantity of fresh green leaves were enclosed in a growth chamber having light intensity of 30,000 Lux and temperature of 30 ± 2°C. The decrease in CO₂ concentration was recorded. In addition to Kallar grass, known C-3 plants namely *Vicia faba* and *Triticum aestivum* were also studied.

Carbon isotope ratios: Leaf tissue was collected from the field and dried in a forced air oven at 80°C for 24 hrs. The dried tissue (5-10 mg) was combusted at 750°C in an excess of oxygen and isotopic ratio (¹³C/¹²C) of the CO₂ evolved was measured on a mass spectrometer as described by Osmond *et al.*, (1978).

This method is based on the observation that plants discriminate against ¹³C during photosynthesis in ways which reflect plant metabolism and environment (Benedict, 1978; O'Leary, 1981). Atmospheric CO₂ contains about 1.1% of the heavier isotope ¹³C and 98.9% of the lighter isotope ¹²C. The discrimination of ¹³C in favour of ¹²C has been highly correlated with the C₃ and C₄ pathways of photosynthetic metabolism. This characteristic when considered in relation to leaf anatomy, provides the most reliable criterion for distinguishing these two photosynthetic pathways (Smith & Brown, 1973). A theory is developed by Farguhar *et al.*, (1982) to explain the carbon isotope composition of plants which is based on diffusion of gaseous CO₂ and carboxylation.

The isotopic composition is specified as Δ¹³C values.

$$\Delta^{13}\text{C}\text{‰} = \left(\frac{R_{\text{sample}}}{R_{\text{standard}}} - 1 \right) \times 1000$$

where R sample and R standard are the ¹³C/¹²C ratios of the sample and the standard. As absolute isotope ratios are troublesome to obtain and for most purposes it is adequate

to give $\Delta^{13}\text{C}_{\text{‰}}$ values in relation to some standard. The standard in general use is PDB (belemnite from Pee Dee Formation in South Carolina, USA) with $^{13}\text{C}/^{12}\text{C}$ ratio = 0.01124 (Craig, 1958) and can be obtained from U.S. National Bureau of Standards. In addition to Kallar grass a known C-4, *Saccharum officinarum* and a C-3 *Rosa indica* were also analysed for $^{13}\text{C}/^{12}\text{C}$ ratios.

Results and Discussions

Anatomical sections of leaves of *Leptochloa fusca* were found to be of Kranz type (Fig. 1a & b). This name was given by Meser (1934) to such leaf structural arrangement in which a chlorenchymatous sheath of large, thick walled cells surrounds vascular bundles. There are dense concentration of chloroplasts, mitochondria and peroxisomes in the bundle sheath cells as compared to mesophyll cells which are often radially arranged (Laetsch, 1971). The Kranz syndrome is a remarkable example of structure related to function with the functional aspects of photosynthetic carbon fixation being correlated with the anatomy of the leaf. Well developed bundle sheath cells were observed around vascular bundle. Adaxial bundle sheath cells of primary lateral bundles are enlarged and devoid of chloroplasts (Fig. 2b). Earlier Kranz syndrome in this grass growing in Southern Africa have been reported by Ellis (1977). Smith & Brown (1973) surveyed the family Gramineae for the occurrence of Kranze syndrome which was based on $^{13}\text{C}/^{12}\text{C}$ ratios

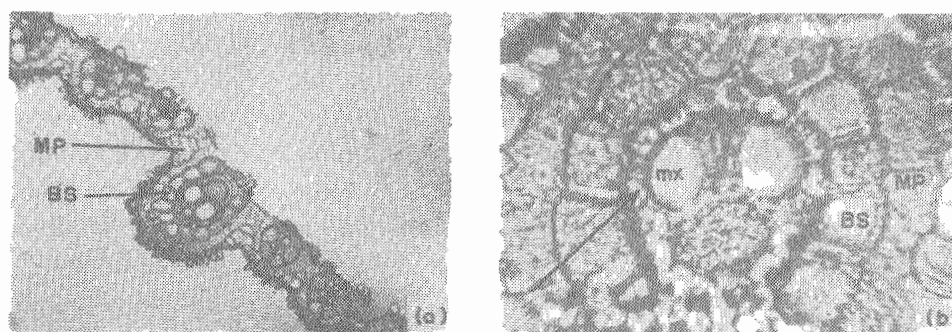


Fig. 1a. Hand cut sections of leaf of Kallar grass showing bundle sheath (BS) and Mesophyll (MP) chlorenchymatous cells ($\times 25.0$).

b. A portion at higher magnification ($\times 100$). Intervening cells between metaxylem vessel (mx) and bundle sheath of primary vascular bundle are indicated by arrow.

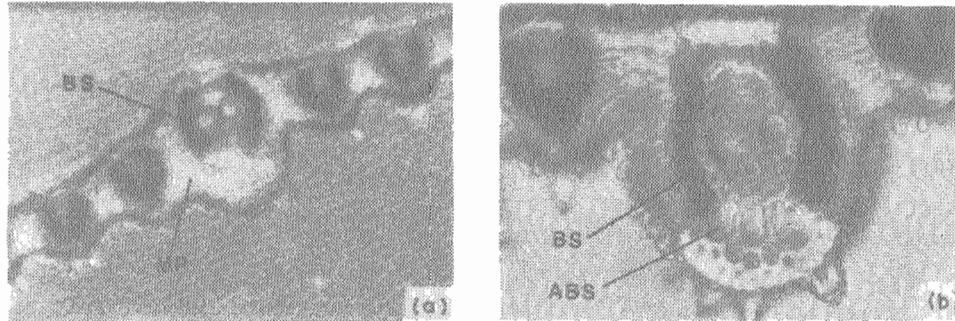


Fig. 2a. IKI-stained dark coloured bundle sheath cells (BS) of Kallar grass. Mesophyll (MP) cells are light in colour as they do not contain starch (x 25.0).

b. Same section at higher magnification (x 100). Empty adaxial cells (ABS) of primary lateral bundle are also evident.

and reported *L. fusca* to be of Kranze type. They did not measure the carbon isotopic ratio of this specie and cited the reference of Chen *et al.*, (1971). A search of literature revealed that no reference of *L. fusca* or its synonym *Diplachne fusca* exists in that report. Our observations confirmed the report of Ellis (1977).

Three sub groups of C-4 photosynthesis are currently recognized as NADP-ME type (NADP-malic enzyme species), NAD-ME type (NAD-malic enzyme species), and PCK type (PEP-carboxykinase species), depending on the reation sequence for C-4 acid decarboxylation in the bundle sheath cells of leaf blades (Hatch *et al.*, 1975). Presence (Xy MS+) or absence (Xy MS-) of cells intervening between metaxylem vessel elements and laterally adjacent chlorenchymatous bundle sheath cells of primary lateral vascular bundle is related to sub grouping in C-4 species by Hatterslay & Watson (1976). They observed a perfect correlation; NADP-Malic enzyme species being Xy MS- while Xy MS+ species were found to be NAD-Malic enzyme or PCK type. Anatomical sections of Kallar grass were found to be Xy MS+ (Fig. 1b) and thus belongs to NAD-Malic enzyme or PCK type. From its anatomy Ellis (1977) categorised it as NAD-Malic enzyme species, however, biochemical studies are needed to confirm the exact C-4 sub group of this grass.

IKI-staining: Clear, well defined bundle sheath cells turned dark after staining with IKI solution as shown in Fig. 2a & b. Rhoades & Carvalho (1944) have shown that for maize and sorghum (C₄ plants) the sheath cells contain specialized plastids concerned with starch formation and no starch was formed in the outer chlorenchymatous cells.

In C_3 species starch is accumulated in the mesophyll cells during the day (Rhoades & Carvalho, 1944) whereas in C_4 species it is only formed in the Kranz sheath cells and not in the mesophyll cells. Downton & Tregunna (1968) determined the correlation between CO_2 -compensation and leaf anatomy alongwith starch distribution. They observed that plants having high CO_2 compensation point (C_3) have starch granules present in whole mesophyll. On the other hand the plant species with low CO_2 compensation point (<10 ppm, C_4) have starch granules restricted to bundle sheath.

CO₂-compensation point: The result of the study made on Kallar grass regarding its compensation point is presented in Fig. 3. CO_2 concentration in the chamber containing Kallar grass leaves decreased from 300 ppm to about 4 ppm in an hour as compared to 70 ppm in case of *Vicia faba* (C_3 -dicot) and 45 ppm in wheat (C_3 -monocot). Chen *et al.*, (1970) included *L. fusca* in the list of plants having low CO_2 -compensation point and thus has C-4 photosynthesis. Our results reconfirm the earlier report.

Carbon isotope ratio: The difference in isotopic composition has become one of the standard methods by which C-4 plants can be distinguished from C-3 plants.

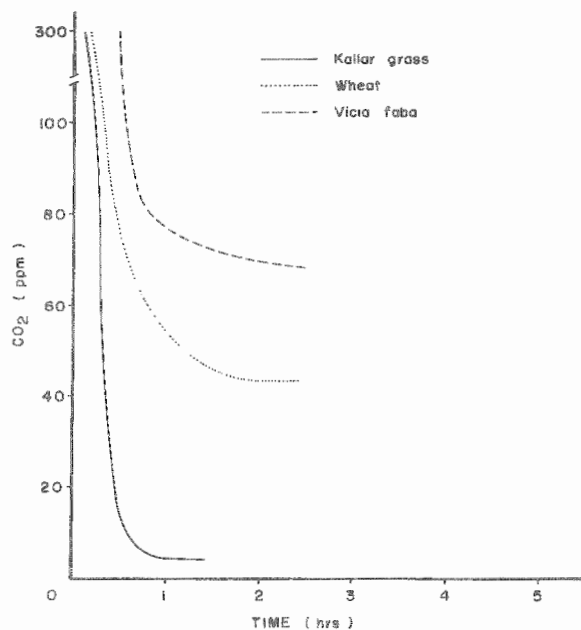


Fig. 3. CO_2 concentration in an enclosed vessel (ul/L) is plotted against time (h). Sealed vessels containing plants were placed in an illuminated growth chamber fitted with an automatic gas sampler for infra red gas analyser and a recorder. Chart speed was 2 mm/min and initial conc. of CO_2 was 300 ppm.

Table 1. $\Delta^{13}\text{C}\text{‰}$ values of plants collected from NIAB-campus.
Standard deviation is $\pm 0.2\%$.

Name of plant	$\Delta^{13}\text{C}\text{‰}$
<i>Saccharum officinarum</i> (Sugarcane C-4)	-12.5
<i>Rosa indica</i> (Wild rose-C-3)	-26.7
<i>Leptochloa fusca</i> (Kallar grass)	-15.9

O'Leary, (1981) reported mean $\Delta^{13}\text{C}\text{‰}$ values -13.5 ± 1.5 and -28.1 ± 2.5 for C-4 and C-3 plants respectively. The $\Delta^{13}\text{C}\text{‰}$ values of Kallar grass and other plants are presented in Table 1. The $\Delta^{13}\text{C}\text{‰}$ value for *L. fusca* ($15.9 \pm 0.2\%$) is higher but it is known that $\Delta^{13}\text{C}$ value is influenced by salinity, dry and hot weather and CO_2 composition in the environment (O'Leary, 1978). Bender (1971) has reported $\Delta^{13}\text{C}\text{‰}$ value of -15.3 for *Cynodon dactylon* (L.) Pers. which is a C-4 grass and this figure is quite close to the observed value of Kallar grass. $\Delta^{13}\text{C}\text{‰}$ readings for sugarcane and wild rose are well in the range of already reported values from other locations. The $\Delta^{13}\text{C}\text{‰}$ value long with other observations are sufficient to conclude that *Leptochloa fusca* which is commonly known as Kallar grass is a C-4 grass.

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References

- Bender, M.M. 1971. Variations in the $^{13}\text{C}/^{12}\text{C}$ ratios of plants in relation to the pathway of photosynthetic carbon dioxide fixation. *Phytochemistry*, 10: 1239-1244.
- Benedict, C.R. 1978. Nature of obligate photoautotrophy. *Ann. Rev. Plant. Physiol.*, 29: 67-93.
- Black, C.C., R.H. Brown and R.C. Moore. 1978. Plant photosynthesis. *Basic Life Sci.*, 10: 95-110.

- Chen, T.M., R.H. Brown and C.C. Black Jr. 1970. CO₂-compensation concentration, rate of photosynthesis and carbonic anhydrase activity of plants. *Weed Sci.*, 18: 399-403.
- Chen, T.M., R.H. Brown and C.C. Black Jr. 1971. Photosynthetic ¹⁴CO₂ fixation products and activities of enzymes related to photosynthesis in Bermuda grass and other plants. *Plant Physiol.*, 47: 199-203.
- Craig, H. 1957. Isotopic standards for carbon and oxygen and correction factors for mass spectrometric analysis of carbon dioxide. *Geochim. Cosmochim. Acta.*, 12: 133-149.
- Dobereiner, J., J.M. Day and P.J. Dart 1972. Nitrogenase activity and oxygen sensitivity of the *Paspalum notatum* - *Azotobacter paspali* association. *J. Gen. Microbiol.*, 71: 103-116.
- Downton, W.J.S. and E.B. Tregunna. 1968. Carbon dioxide compensation - its relation to photosynthetic carboxylation reactions, systematics of the Gramineae and leaf anatomy. *Can. J. Bot.*, 46: 207-215.
- Ellis, R.P. 1977. Distribution of the Kranz syndrome in the Southern African Eragrostoideae and Panicoideae according to bundle sheath anatomy and cytology. *Agroplanta*, 9: 73-110.
- Farquhar, G.D., M.H. O'Leary and J.A. Berry. 1982. On the relationship between carbon isotope discrimination and the intercellular carbon dioxide concentration in leaves. *Aust. J. Plant Physiol.*, 9: 121-137.
- Hatch, M.D. and C.R. Slack. 1970. Photosynthetic CO₂-fixation pathways. *Ann. Rev. Plant Physiol.*, 21: 141-162.
- Hatch, M.D., T. Kagawa and S. Craig. 1975. Subdivision of C-4 pathway species based on differing C-4 acid decarboxylating systems and ultrastructural features. *Aust. J. Plant Physiol.*, 2: 111-128.
- Hattersley, P.W. and L. Watson. 1976. C-4 grasses: an anatomical criterion for distinguishing between NADP-Malic enzyme species and PCK or NAD-Malic species. *Aust. J. Bot.*, 24: 297-213.
- Laetsch, W.M. 1971. Chloroplast structural relationships in leaves of C-4 plants. In: *Photosynthesis and photorespiration*. M.D. Hatch, C.B. Osmond and R.O. Slatyer (Eds.) pp. 323-349, Wiley-Interscience, New York.
- Malik, K.A., Y. Zafar and A. Hussain. 1980. Nitrogenase activity in rhizosphere of Kallar grass. *Biologia*, 26: 107-112.
- Malik, K.A., Y. Zafar and A. Hussain. 1981. Associative dinitrogen fixation in *Diplachne fusca* (Kallar grass). In: *BNF technology for Tropical Agriculture*. P.H. Graham & S.C. Harris (Eds.) pp. 503-307. CIAT, Cali Colombia.

- Moser, H. 1934. Untersuchungen über die Blattstruktur von *Atriplex*. *Beih. Bot. Zentralbl.*, 52: 378-388.
- O'Leary, M.H. 1978. Heavy atom isotope effect in enzyme-catalyzed reactions. In: *Transition States of Biochemical Processes*. R. Gandour and R.L. Schowen (Eds.) pp. 285-316. Plenum Press, New York.
- O'Leary, M.H. 1981. Carbon isotope fractionation in plants. *Phytochemistry*, 20: 533-567.
- Osmond, C.B., H. Ziegler, W. Stichler and P. Trimbon 1975. Carbon isotope discrimination in alpine succulent plants supposed to be capable of crassulacean acid metabolism (CAM). *Oecologia* (Berl.), 18: 209-217.
- Rhoades, M.M. and A. Corvalho. 1944. The function and structure of the parenchyma sheath plastids of the maize leaf. *Bull. Torrey Bot. Club.*, 71: 335-340.
- Smith, B.N. and N.V. Brown. 1973. The kranz syndrome in the gramineae as indicated by carbon isotopic ratios. *Amer. J. Bot.*, 60: 505-513.