ISOLATION OF FIVE FRANKIA STRAINS FROM ACTINORHIZAL NODULES OF CASUARINA GLAUCA

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Abstract

Five strains of Frankia isolated from root nodules of Casuarina glauca collected from different localities of Islamabad were characterized by the occurrence of branched and septate hyphae bearing sporangia and vesicles typical of Frankia. The strains showed marked differences in their growth requirements, growth rate, nitrogen fixing ability, production of pigments and infectivity of host seedlings, CAQP1 and CAQP2 strains infected C. glauca seedlings while CAQP3, CAQP4 and CAQP5 failed to induce nodules on the host.

Introduction

Microsymbionts that are responsible for the formation of actinorhizae have been classified in the genus Frankia of actinomycetes. The first success in isolation of Frankia and its cultivation in vitro was achieved in Comptonia peregrina by Callaham, et al., (1978). Since then a number of Frankia strains have been isolated from the actinorhizae of Alnus (Baker, et al., 1979. Hafeez et al., 1984), Casuarina (Diem et al., 1982), Colletia (Akkermans et al., 1984), Hippophae (Burggraff, et al., 1981) and Myrica (Baker, et al., 1981). Diem, et al., (1982) have isolated 5 Frankia strains from root nodules of Casuarina equisetifolia by serial dilution technique. Gauthier, et al., (1981) reported the isolation of 2 actinomycete strains from C. equisetifolia which reduced C₂H₄ in vitro but were unable to nodulate the host seedlings. Later Diem, et al., (1983) reported successful isolation of an effecive nodulating strain of Frankia from Casuarina spp. In Pakistan Casuarina glauca and C. equisetifolia are common species that have been introduced of which C. glauca is quite common in Islamabad. The present studies report the isolation and characterization of 5 Frankia strains from C. glauca.

Materials and Methods

C. glauca root nodules collected from different localities of Islamabad were used. Nodules surface sterilized in 0.1% HgCl₂ were crushed into suspension, 1 g nodule/10 ml sterile distilled water from which serial dilution of $10^{-2} - 10^{-4}$ were spread over QMOD solid medium, 0.2 ml/Petri plate. The dishes were incubated at 28-30°C. The colonies of Frankia were visible in 3 weeks. The isolates were purified by repeated subculturing from isolated colonies by dilution and plating. Five pure strains viz., CAQP₁, CAQP₂, CAQP₃, CAQP₄ and CAQP₅ were seperated and propagated in liquid and solid media.

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Surface sterilized nodules were cut into small pieces and inoculated on QMOD agar slant. After 2 weeks of incubation the apparently axenic nodule parts were transferred into tubes containing 10 ml of liquid QMOD medium and incubated at 29°C. Frankia colonies attached with the cut exposed surfaces of the nodule pieces were visible within 3 weeks. Flocks of Frankia were disintegrated by passing through a syringe needle (26G x 11mm) and purified by subculturing onto QMOD and NH₄-propionate medium (Lalonde & Calvert, 1979).

Isolates cultured on QMOD, P + N (Propionate + NH_4Cl) and P - N (Propionate - NH_4Cl) medium (Hafeez, et al., 1984) were harvested by centrifugation and washed once with phosphate buffer (50 mM, pH 7.0). One month old cultures of the isolates were inoculated with 0.1 ml (base soluble protein) of washed cells in Erlenmeyer flasks, containing 50 ml of medium. The content of base soluble protein was determined according to Moss & Bond (1957).

One month old culture of each strain was washed twice by centrifugation at 2700 rpm for 5 min with 50 mM phosphate buffer, pH 7.0 and homogenized by forcing through a syringe needle, 26G x 11 mm. Eight week old seedlings of *C. glauca* were inoculated by immersing the roots in nitrogen free solution containing *Frankia* isolates and grown in test tubes (Gibson, 1963) filled with ½ strength nitrogen free Hoagland solution (Hoagland & Arnon, 1938) and modified leonard's bottle jar assemblies (Vincent, 1970). In sand culture experiment the seedlings were inoculated with suspension of washed, homogenized 1 month old culture by adding it in sand near the roots of the seedlings. Nodulation was recorded 3 months after inoculation.

Nitrogenase activity of pure cultures of *Frankia* and of the nodulated plants was determined by acetylene reduction assay. Cells of *Frankia* isolates grown in stationary Erlenmeyer flasks with P — N medium were harvested and 5 ml of concentrated cells taken in 16.6 ml air tight tubes with rubber stopper and incubated with 10% acetylene at 29°C. After 1 h 0.1 ml of gas samples were taken from each tube and analyzed chromatographically on Hitachi, 163 gas chromatograph (Hardy, et al., 1968).

Results

Morphology: Strains CAQP₁, CAQP₂ and CAQP₄ were pigmented with distinctly pinkish red orange, red and pink colonies respectively, while strains CAQP₃ and CAQP₅ were non-pigmented. Growth of the colonies on solid media was slower than in liquid media. Colonies of Frankia did not cause turbidity in the liquid medium and remained undisturbed even when shaken. The strains grew and remained at the bottom of the test tube in QMOD medium, while in NH₄-Propionate and Tween-80 medium the colonies were sticky and adhered to the walls of the test tube. The hyphae were septate and branched. The vesicles were spherical, formed terminally on short parental hyphae. The sporangia of different sizes with irregular shape were always formed terminally.

| C Carefug Tallando e American commente (1922) 2000 | | Nitrogenase activity** | | | |
|--|-----------------|---|------------------|--|--|
| Frankia isolates P - N | | (mg protein 1 ⁻¹)* P + N | QMOD | (nmoles C_2H_4 mg ⁻¹ protein hr ⁻¹) P - N | |
| CAQP ₁ | 4.19 ± 0.52 | 8.95 ± 0.86 | 16.84 ± 1.54 | 323 (278-380) | |
| $CAQP_2$ | 3.72 ± 0.31 | 6.59 ± 0.55 | 14.70 ± 1.33 | 166 (150-179) | |
| $CAQP_3$ | 6.00 ± 0.70 | 9.16 ± 1.05 | 22.50 ± 1.81 | 60 (42- 90) | |
| CAQP ₄ | 5.40 ± 1.10 | 7.44 ± 0.81 | 17.70 ± 1.21 | 35 (29- 45) | |
| CAQP ₅ | 3.20 ± 0.58 | 1.90 ± 0.48 | 12.88 ± 1.29 | 200 (180-230) | |

Table 1. Yield* and nitrogenase activity of Frankia isolates.

Growth yield on various media: All the 5 isolates showed growth on Propionate medium that lacked combined nitrogen (Table 1). Maximum growth of P-N medium was observed in strain $CAQP_3$ whereas $CAQP_5$ showed minimum growth. Growth rate of all the 5 isolates was higher in QMOD medium than on P-N and P+N media.

Nitrogenase activity: All the 5 strains grew on nitrogen free media with propionate as carbon source and were able to fix nitrogen in vitro (Table 1). Strain CAQP₁ exhibited high acetylene reduction rate of 323 n mole C₂H₄ mg protein⁻¹ h⁻¹ while strain CAQP₄ showed the lowest rate (Table 1).

Infectivity tests: Infectivity of Frankis strains tested on host seedlings showed nodule formation after 40 days of inoculation. Strains CAQP₁ and CAQP₂ were infective (Table 2) in water as well as in sand culture experiments while strains CAQP₃, CAQP₄ and CAQP₅ failed to nodulate the host seedlings.

Discussion

The morphological characteristics of all the 5 strains isolated from Casuarina glauca root nodules are similar to the Frankia strains isolated from other types of actinorhizae in having prokaryotic septate and branched hypahae, irregular sporangia and spherical vesicles. Strains isolated from other actinorhizal plants (Callaham et al., 1978; Diem et al., 1982) always exhibited hyphae and sporangia but vesicles appeared only on QMOD and N free medium.

^{*}After 21 days of growth. Values are mean of 5 determinations.

^{**}Values are mean of 5 determinations. Maximum and minimum values are given in parentheses.

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| Frankia | Inoculated/ | Nodulation | L | Inoculated/ | Nodulation | |
| isolates | Nodulated | % | Effectivity* | Nodulated | % | Effectivity |
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| CAQP ₁ | 20 (11) | 55 | + | 20 (17) | 85 | + |

20 (16)

20 (0)

20 (0)

20 (0)

80

Table 2. Nodulation of Casuarina glauca by Frankia isolates.

25

20 (5)

20 (0)

20(0)

20(0)

20 (0)

CAQP₂ CAQP₃

CAQP4

CAOP₅

Control

Production of insoluble pigments have been reported for the isolates from *Elaeagnus* (Baker et al., 1979), Casuarina (Gauthier et al., 1981), Alnus (Hafeez et al., 1984). The strains from C. equisetifolia isolated by Diem et al., (1983) were non-pigmented. During the present investigation isolates CAQP₁, CAQP₂ and CAQP₄ produced pigments while CAQP₃ and CAQP₅ were non-pigmented. Pigmented strains did not cause turbidity in the medium indicating the insoluble nature of the pigments.

All the 5 strains formed spherical vesicles on various media. They were able to grow in nitrogen free medium, though at different rates. The measurement of their nitrogenase activity (Table 1) confirmed that these strains were free living N₂-fixers like various other Frankia strains (Gauthier et al., 1981; Diem et al., 1982). Non formation of vesicles in the medium with combined nitrogen supports the view that vesicles are involved in nitrogen fixation like heterocysts in cyanobacteria (Tjepkema et al., 1980).

Theoretically the 3 strains unable to nodulate the host seedlings could not be considered as endophytes. Isolates of Frankia from C. equisetifolia did not nodulate their own host (Diem et al., 1982). Lechevalier & Lechevalier (1983) considered free-living actinomycetes which produce vesicles and sporangia as members of Frankia even if they had no known nodule forming capacity. Failure of Frankia to nodulate host plant may be linked to the use of inappropriate isolation procedures (Diem et al., 1983). The inability of CAQP₃, CAQP₄ and CAQP₅ strains to infect host plant may not necessarily be related to the isolation techniques since similar techniques were used as has been successfully used by Lalonde et al., (1981). Out of the 5 isolates strain CAQP₄ is pigmented and non-infective. It falls under the category "A" according to the classification proposed by Lechevalier (1984) which is based upon morphology, physiology, cell chemistry and infectivity of the host plant. However, the morphological characteristics and infectivity

^{*}As determined by acetylene reduction assay.

of other strains are not in conformity with the categorization of Lechevalier (1984). There is therefore a need for more critical investigations for the categorization of *Frankia* isolates.

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