

PHYTOPLANKTON ASSEMBLAGES IN THE COASTAL ZONE OF THE GULF OF İSKENDERUN - NORTH EASTERN MEDITERRANEAN

AYŞE NESLİHAN ÖZMAN-SAY¹ AND NESLİHAN BALKİS^{2*}

¹Gebze Institute of Technology, Department of Environmental Engineering, Gebze 41400, Kocaeli, Turkey

²Istanbul University, Faculty of Science, Department of Biology, 34134-Vezneciler, Istanbul, Turkey

*Corresponding author's e-mail: neslbalk@istanbul.edu.tr

Abstract

In order to determine the phytoplankton species in the Gulf of İskenderun and the environmental factors that affect the distributions of the species, samples were collected horizontally with a plankton net at 9 stations in November 2005 and June 2006. As a result of the examination of the samples, 95 phytoplankton species belonging to 3 classes were identified. 12 species were new records for the Gulf of İskenderun, one (*Navicula transitans* Cleve, 1883) of them was a new report for all the Turkish seas. Dinoflagellates were the majority group of the species composition (53.7%), followed by diatoms (45.3%). Most of the species identified in this study consist of phytoplanktonic algae reported in the Eastern Mediterranean earlier. These species were neritic, oceanic, temperate and subtropic. In this study, 18 species defined as potentially harmful algal species and three of them were also toxic. Primary hydrographic conditions, such as temperature (20.2-28.7 °C), salinity (31.0-39 ppt) and dissolved oxygen (7.21-8.60 mg l⁻¹), were recorded on each sampling occasion. In addition, a check-list of the phytoplankton species recorded up to this date was given and totally 269 species were recorded from the Gulf of İskenderun.

Introduction

The eastern part of the Mediterranean is one of the most oligotrophic area in the world in terms of nutrient concentrations and productivity (Azov, 1986; Krom *et al.*, 1991) and the oligotrophy increases from the west to the east of the Mediterranean Sea (Ignatiades *et al.*, 2002). The coastal zone receives considerable amounts of freshwater, nutrients, dissolved and particulate organic matter, sediment and contaminants. It is a transition area between land and the open ocean (Gazeau, 2004). There are also limited upwelling areas and few rivers discharging nutrients to the Mediterranean (Dugdale & Wilkerson, 1988). The nutrient distribution and phytoplankton production in the eastern Mediterranean is principally determined by the durations and intensity of deep winter mixing, which provides transport of nutrients from deeper layer to the surface (Yılmaz & Tuğrul, 1998).

There are many local studies on phytoplankton communities, including information regarding their spatial and temporal variations along the eastern Mediterranean coastline (Lakkis & Lakkis, 1980; Azov, 1986; Aktan, 2011; Drira *et al.*, 2010). Gulf of İskenderun an important area for fisheries is located in the northeast of the eastern Mediterranean. Primary production in the gulf is about 2-4 times higher than that in the north-eastern Mediterranean; average production is 115gC/m²/year. The average depth is 70m and the depth of 1% light transmission ranges between 40-70m (Yılmaz *et al.*, 1992). Especially, pollutants coming from highly populated cities, agricultural areas, industrial facilities and the inputs from the regional rivers are the main factors affecting the structure of the water column in the area (Polat *et al.*, 2006). As oligotrophic areas are very sensitive to environmental variations, their monitoring is essential for the evaluation of the long-term changes in the community structure. Gulf of İskenderun, regularly sampled since 1972 (Gökalp, 1972), is especially

interesting due to several studies conducted in this area which facilitates comparison. Detailed studies on phytoplankton and their ecological features were carried out in the Gulf of İskenderun (Polat *et al.*, 2000, 2005, 2006; Polat & Işık, 2002; Polat & Piner, 2002; Polat, 2002, 2004, 2007a, 2007b, 2007c, 2008; Aka & Polat 2006; Polat & Koray 2007).

The aim of this study was to determine the composition of microphytoplankton species in this area and to report, if any, species that would be new for this region and existing harmful algal species (HAS). A list of phytoplankton species, of Gulf of İskenderun along with nomenclatural changes if any, is also provided.

Materials and Methods

This research was carried out in neritic waters of the Gulf of İskenderun in November 2005 and June 2006. The samples were gathered from the subsurface using a 40cm diameter plankton net with a 55µm mesh size at 9 stations (Fig. 1). Material was fixed with neutral 4% formaldehyde solution and observed by using an Olympus CK2 inverted phase-contrast microscope equipped with a microphotosystem at a magnification of 400x. References used to identify the phytoplankton species can be found in previous work (Balkis, 2003, 2005). The recent revisions of phytoplankton species were presented according to MarBEF Data System-European of Marine Species (ERMS) (<http://www.marbef.org/data/erms.php>), Hasle & Syvertsen (1997) and Gomez (2005).

Water samples for physical and chemical analyses were collected using a 3 l Ruttner bottle with a thermometer. Temperature (°C), salinity (ppt) and dissolved oxygen (mg l⁻¹) were measured at each sampling point (Fig. 2). The Mohr-Knudsen method (Ivanoff, 1972) was used for measuring the salinity values, and the Winkler (1888) method was used for determining dissolved oxygen (DO).

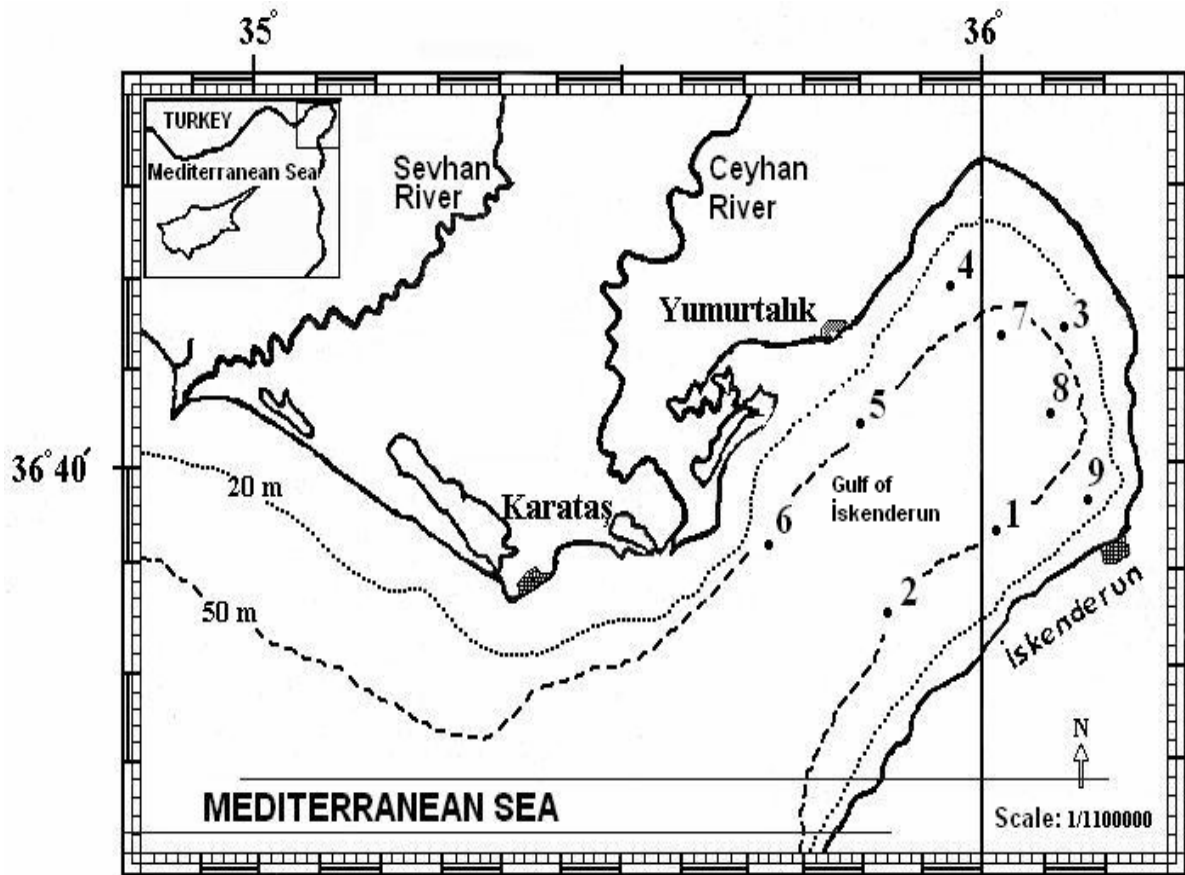


Fig. 1. Research stations in neritic waters of the Gulf of İskenderun.

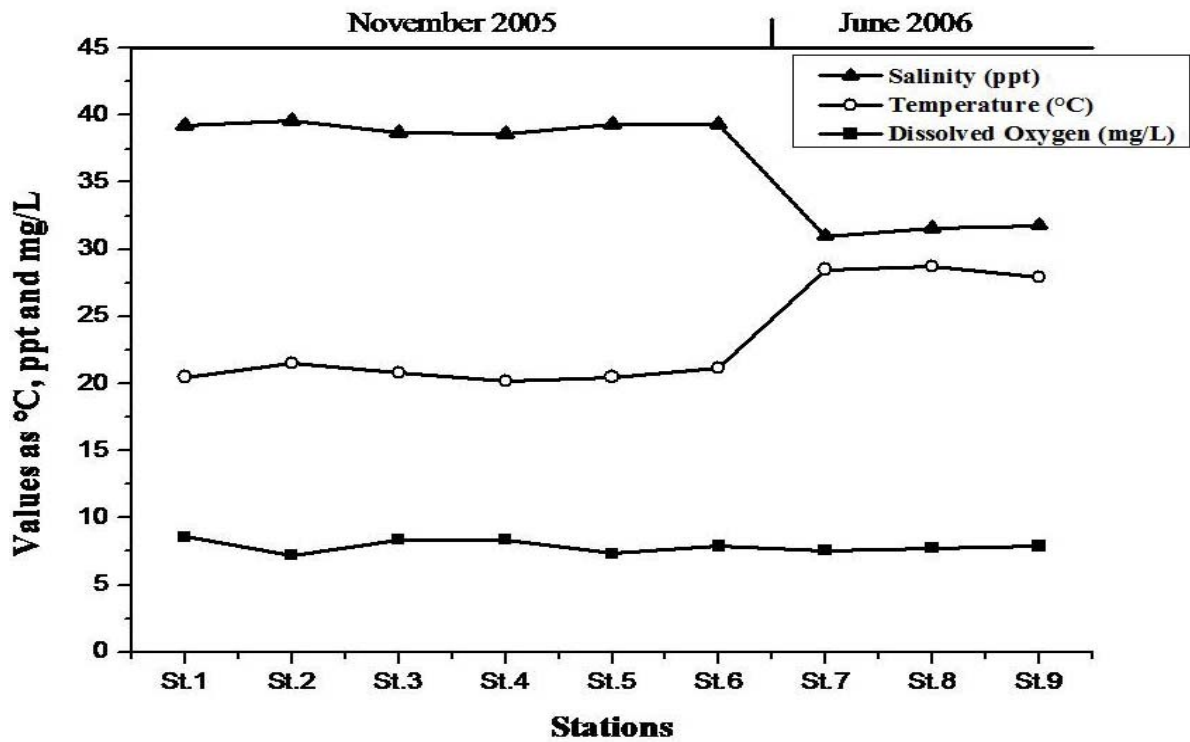


Fig. 2. Variations of temperature (°C), salinity (ppt) and dissolved oxygen (mg l⁻¹) at the research stations.

The occurrence of each species at the sampling stations were categorized into frequency groups as followed; V: very abundant (81-100%), A: abundant (61-80%), C: common (41-60%), R: rare (21-40%) and X:

present sporadically (1-20%) (Fig. 3). The Sorensen similarity matrix (Sorensen, 1948) was performed in order to group the sampling stations according to the species involved (Fig. 4).

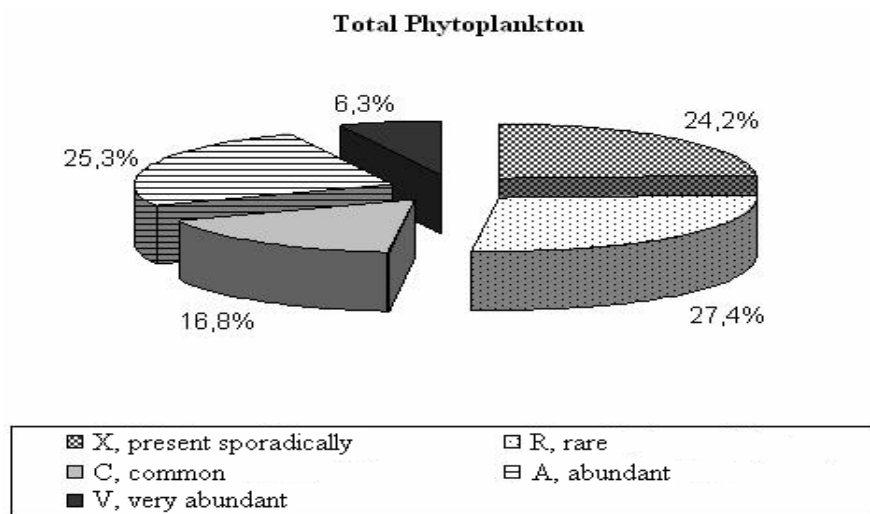


Fig. 3. Distribution of phytoplankton species according to frequency index.

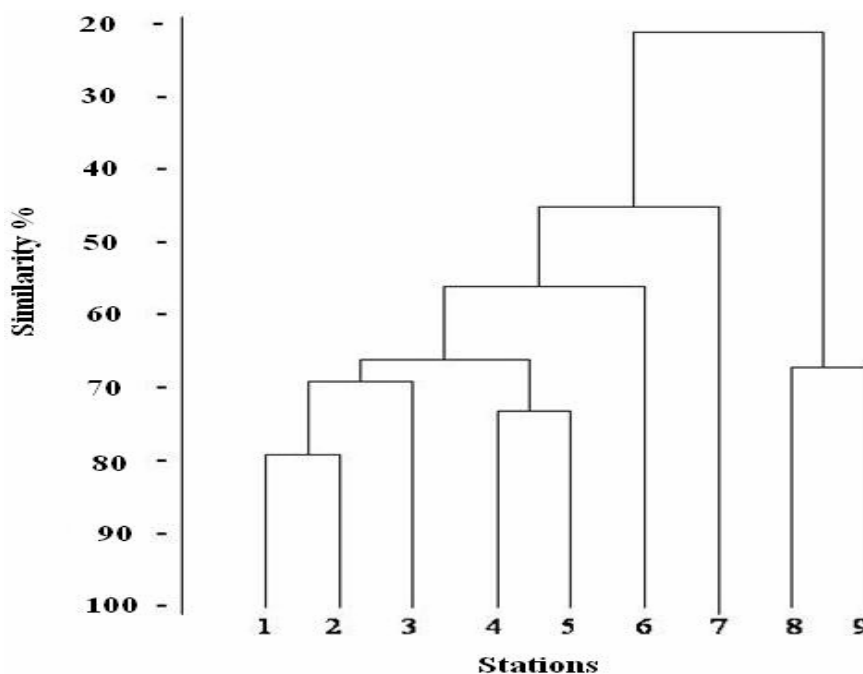


Fig. 4. Sorensen similarity dendrogram of the sampling stations in neritic waters of the Gulf of İskenderun.

Results

From the analysis of phytoplankton community composition, 95 species of three different algal groups were identified: 51 dinoflagellates (53.7%), 43 diatoms (45.3%) and 1 dictyochophyceans (1%) (Table 1). Of these, 1 species (*Navicula transitans* Cleve) was new to Turkish seas, and so were 11 species (*Dinophysis diegensis* Kofoid, *D. fortii* Pavillard, *Prorocentrum gracile* Schütt, *P. priestinum* Schiller from dinoflagellates,

Amphora ovalis (Kützing) Kützing, *Dactyliosolen antarcticus* Castracene, *Diploneis bombus* (Ehrenberg) Cleve, *Nitzschia lorenziana* Grunow in Cleve & Grunow, *Petrodictyon gemma* (Ehrenberg) D.G. Mann, *Pseudonitzschia pseudodelicatissima* (Hasle) Hasle and *Thalassiothrix longissima* Cleve & Grunow from diatoms) to the Gulf of İskenderun. The most common genera were as follows: *Ceratium* Schrank (22 species), *Chaetoceros* Ehrenberg (9 species) and *Protoberidium* Bergh (6 species).

Table 1. List and frequency distribution of phytoplanktonic taxa found during the study period in the Gulf of İskenderun. V, very abundant (81-100%); A, abundant (61-80%); C, common (41-60%); R, rare (21-40%); X, present sporadically (1-20%).

Species	Stations									f (%)	
	November 2005						June 2006				
	1	2	3	4	5	6	7	8	9		
Dinophyceae											
<i>Amphisolenia bidentata</i>			+			+					R
<i>Ceratium arietinum</i>			+						+		R
<i>Ceratium candelabrum</i>	+	+	+	+	+	+	+	+	+		V
<i>Ceratium carriense</i>	+		+			+					R
<i>Ceratium concilians</i>	+	+	+	+	+	+					A
<i>Ceratium contortum</i>	+	+	+			+					C
<i>Ceratium contrarium</i>	+	+	+	+	+	+					A
<i>Ceratium declinatum</i>	+	+	+			+					C
<i>Ceratium euarquatium</i>	+	+	+	+							C
<i>Ceratium extensum</i>	+	+				+	+	+	+		A
<i>Ceratium falcatum</i>	+			+							R
<i>Ceratium furca</i>	+	+		+	+		+	+	+		A
<i>Ceratium fusus</i>	+	+		+			+	+	+		A
<i>Ceratium hexacanthum</i>		+				+					R
<i>Ceratium horridum</i>	+	+	+	+	+	+					A
<i>Ceratium kofoidii</i>	+	+	+	+	+	+	+				A
<i>Ceratium macroceros</i>	+	+	+	+	+						C
<i>Ceratium pentagonum</i>				+		+					R
<i>Ceratium ranipes</i>		+									X
<i>Ceratium symmetricum</i>		+		+	+	+					C
<i>Ceratium teres</i>	+	+	+	+		+					C
<i>Ceratium trichoceros</i>	+	+	+	+	+	+					A
<i>Ceratium tripos</i>	+	+	+	+	+	+		+	+		V
<i>Ceratocorys horrida</i>	+		+			+	+	+	+		A
<i>Dinophysis caudata</i>	+	+			+	+	+	+	+		A
<i>Dinophysis diegensis</i>							+				X
<i>Dinophysis fortii</i>									+		X
<i>Dinophysis hastata</i>		+									X
<i>Dinophysis ovum</i>	+	+	+		+			+	+		A
<i>Diplopsalis lenticula</i>								+	+		R
<i>Goniodoma polyedricum</i>	+	+	+	+	+	+	+	+			V
<i>Gonyaulax polygramma</i>									+		X
<i>Ornithocercus heteroporus</i>						+					X
<i>Ornithocercus magnificus</i>	+	+	+	+	+	+					A
<i>Ornithocercus quadratus</i>		+									X
<i>Ornithocercus steinii</i>	+	+	+			+			+		C
<i>Phalacroma doryphorum</i>	+		+	+				+	+		R
<i>Phalacroma rapa</i>			+					+	+		R
<i>Podolampas bipes</i>	+	+	+		+						C
<i>Prorocentrum gracile</i>	+						+				R
<i>Prorocentrum triestinum</i>								+			X
<i>Protoperidinium conicum</i>			+								X
<i>Protoperidinium divergens</i>								+	+		R
<i>Protoperidinium globulus</i>								+	+		R
<i>Protoperidinium mediterraneum</i>	+										X
<i>Protoperidinium oceanicum</i>			+								X
<i>Protoperidinium steinii</i>	+							+	+		R

Table 1. (Contd.)

Species	Stations									f (%)
	November 2005						June 2006			
	1	2	3	4	5	6	7	8	9	
<i>Pyrophacus horologium</i>			+				+			R
<i>Pyrocystis elegans</i>			+							X
<i>Pyrocystis hamulus</i>		+				+				R
<i>Spiraulax Jolliffei</i>						+				X
Dictyochophyceae										
<i>Dictyocha fibula</i>	+		+	+	+	+				C
Bacillariophyceae										
<i>Amphora ovalis</i>			+							X
<i>Asterolampra marylandica</i>	+	+				+				R
<i>Bacillaria paxillifera</i>	+									X
<i>Bacteriastrium delicatulum</i>	+	+	+	+	+	+	+		+	V
<i>Bacteriastrium elegans</i>	+	+	+	+	+	+				A
<i>Cerataulina pelagica</i>	+	+			+					R
<i>Chaetoceros affinis</i>	+	+	+	+						C
<i>Chaetoceros dadayi</i>	+	+	+							R
<i>Chaetoceros danicus</i>	+	+	+	+	+		+			A
<i>Chaetoceros decipiens</i>	+	+	+	+	+	+			+	A
<i>Chaetoceros didymus</i>	+	+	+	+	+					C
<i>Chaetoceros diversus</i>	+	+							+	R
<i>Chaetoceros lorenzianus</i>	+	+	+	+	+	+	+			A
<i>Chaetoceros peruvianus</i>	+	+	+	+	+					C
<i>Chaetoceros rostratus</i>	+	+		+	+		+	+		A
<i>Coscinodiscus radiatus</i>							+		+	R
<i>Cylindrotheca closterium</i>		+	+		+		+			C
<i>Dactyliosolen antarcticus</i>	+	+	+		+		+			C
<i>Dactyliosolen mediterraneus</i>				+			+			R
<i>Diploneis bombus</i>			+							X
<i>Eucamphia zodiacus</i>	+									X
<i>Guinardia striata</i>	+	+	+	+	+	+	+			A
<i>Hemiaulus hauckii</i>	+	+	+	+	+	+	+			A
<i>Leptocylindrus minimus</i>	+	+	+	+	+	+	+			A
<i>Lithodesmium undulatum</i>				+						X
<i>Navicula transitans</i>					+	+	+			R
<i>Nitzschia longissima</i>	+		+				+			R
<i>Nitzschia lorenziana</i>			+							X
<i>Nitzschia sigma</i>						+				X
<i>Odontella aurita</i>	+									X
<i>Odontella mobiliensis</i>				+						X
<i>Petrodictyon gemma</i>		+							+	R
<i>Pleurosigma normanii</i>	+		+	+	+	+		+		A
<i>Proboscia alata</i>	+	+	+		+	+	+	+	+	V
<i>Pseudo-nitzschia pseudodelicatissima</i>	+		+	+			+			C
<i>Pseudosolenia calcar-avis</i>	+	+	+	+	+	+				A
<i>Rhabdonema adriaticum</i>			+							X
<i>Rhizosolenia imbricata</i>	+	+	+	+	+	+				A
<i>Rhizosolenia styliformis</i>	+	+	+	+						C
<i>Striatella unipunctata</i>	+		+		+					R
<i>Thalassionema frauenfeldii</i>	+	+	+	+	+	+	+			A
<i>Thalassionema nitzschioides</i>	+	+	+	+	+	+	+	+	+	V
<i>Thalassiothrix longissima</i>					+	+	+			R

Of the 95 species that belong to the classes Dinophyceae, Dictyochophyceae and Bacillariophyceae, the number of present sporadically species is 23 (24.2%), rare species is 26 (27.4%), common species is 16 (16.8%), abundant species is 24 (25.3%) and very abundant species is 6 (6.3%) (Fig. 3). Of the very abundant species, the frequency for the species *Ceratium candelabrum* and *Thalassionema nitzschioides* is 100%.

When the dendrogram was analysed the similarity between the stations in terms of identified species was produced (Fig. 4) using Sorensen's (1948) similarity formula and 8 of the stations showed 50% and more similarity. Among these stations, the similarity between

stations 1 and 2 is 79%, stations 4 and 5 is 73%, and stations 8 and 9 is 67%. It was also found that especially the stations that belong to the same sampling period are grouped noticeably among each other.

In the present study, it was determined that out of 95 species 18 are potentially harmful phytoplankton species, and the diet and dangerous effects of these species were shown in Table 2. While three of these species (*Dinophysis caudata*, *D. fortii*, *Pseudo-nitzschia pseudodelicatissima*) are toxin-producing species at the same time, It was seen that the highest number of HAB species belong to diatoms.

Table 2. Potentially harmful species (HAS) observed in the coastal zone of the Gulf of İskenderun during the sampling dates which have had harmful impacts indicated elsewhere.

Species	Trophic state	Harmful
Dinophyceae		
<i>Ceratium furca</i>	Mixo	Bloom-forming, fish-killing, anoxia
<i>Ceratium fusus</i>	Mixo	Fish-killing
<i>Dinophysis caudata</i>	Auto	Bloom-forming, DSP toxin, ichthyotoxins
<i>D. fortii</i>	Auto	Bloom-forming, DSP toxin
<i>Gonyaulax polygramma</i>	Auto	Fish and shellfish kills due to anoxia after red tide
<i>Prorocentrum triestinum</i>	Auto	Bloom-forming, fish-killing
<i>Pyrophacus steinii</i>	Auto	Bloom-forming
Dictyochophyceae		
<i>Dictyocha fibula</i>	Auto	Fish-killing
Bacillariophyceae		
<i>Bacillaria paxillifera</i>	Auto	Bloom-forming
<i>Cerataulina pelagica</i>	Auto	Bloom-forming
<i>Chaetoceros didymus</i>	Auto	Bloom-forming
<i>Cylindrotheca closterium</i>	Auto	Bloom-forming
<i>Eucampia zodiacus</i>	Auto	Bloom-forming
<i>Nitzschia longissima</i>	Auto	Bloom-forming
<i>Odontella aurita</i>	Auto	Bloom-forming
<i>Proboscia alata</i>	Auto	Bloom-forming
<i>Pseudo-nitzschia pseudodelicatissima</i>	Auto	ASP toxin
<i>Thalassionema nitzschioides</i>	Auto	Bloom-forming

Trophic state (auto-, hetero- or mixotrophic), DSP: diarrhetic shellfish poisoning, ASP: amnesic shellfish poisoning. The effects of HAS were summarized according to Hallegraeff (1993), Faust and Gullede (2002), Lu and Hodgkiss (2004), and Vila and Masó (2005).

The list of phytoplankton species recorded up to this date were given in Table 3 and a total of 269 microalgae, 11 of them were identified at the generic level: 1 Cyanophyceae, 148 Dinophyceae, 2 Dictyochophyceae, 115 Bacillariophyceae, 1 Prasinophyceae and 2 Prymnesiophyceae (Table 3). Dinoflagellates represented the majority of the population (55%), followed by diatoms (42.8%). The genera *Ceratium* (40 species), *Chaetoceros* (28 species) and *Protoperdinium* (27 species) were dominant in terms of diversity.

Discussion

So far, a total of 269 taxa have been identified from the Gulf of İskenderun. Gökalp (1972) reported 23 species, Polat *et al.*, (2000) 153 species, Polat & Işık (2002) 129 species, Polat & Piner (2002) 90 species, Polat (2004) 1 species, Polat *et al.*, (2005) 11 species, Aka & Polat (2006) 71 species, Polat *et al.*, (2006) 36 species, Polat (2007a) 1 species, Polat (2007b) 6 species, Polat (2007c) 160 species, Polat & Koray (2007) 140 species, Polat (2008) 30 species and in the present research 95 species.

Table 3. List of phytoplanktonic taxa recorded up to this date in neritic waters of the Gulf of İskenderun.

Species	Gökakın (1972)	Polat <i>et al.</i> , (2000)	Polat & Işık (2002)	Polat & Piner (2002)	Polat (2004)	Polat <i>et al.</i> , (2005)	Aka & Polat (2006)	Polat <i>et al.</i> , (2006)	Polat (2007a)	Polat (2007b)	Polat (2007c)	Polat & Koray (2007)	Polat (2008)	This research
Cyanophyceae														
<i>Oscillatoria</i> sp.		+	+					+			+			
Dinophyceae														
<i>Akashiwo sanguinea</i> (Hirasaka) G. Hansen & Moestrup								+			+	+		
<i>Amphisolenia bidentata</i> Schröder		+	+								+	+		+
<i>Amphisolenia palaeotheroides</i> Kofoid												+		
<i>Amphisolenia truncata</i> Kofoid & Michener		+										+		
<i>Centrodinium pavillardii</i> F.J.R. Taylor												+		
<i>Ceratium arietinum</i> Cleve		+	+	+							+	+		+
<i>Ceratium azoricum</i> Cleve												+		
<i>Ceratium belone</i> Cleve												+		
<i>Ceratium breve</i> (Ostenfeld & Schmidt) Schröder		+									+	+		
<i>Ceratium candelabrum</i> (Ehrenberg) Stein		+	+	+							+	+	+	+
<i>Ceratium carriense</i> Gourret	+	+	+								+	+		+
<i>Ceratium concilians</i> Jørgensen	+	+	+								+	+		+
<i>Ceratium contortum</i> (Gourret) Cleve		+	+	+							+	+	+	+
<i>Ceratium contrarium</i> (Gourret) Pavillard	+	+	+								+	+		+
<i>Ceratium declinatum</i> (Karsten) Jørgensen		+	+								+	+		+
<i>Ceratium digitatum</i> Schütt												+		
<i>Ceratium euarcuatum</i> Jørgensen		+	+								+	+		+
<i>Ceratium extensum</i> (Gourret) Cleve		+	+	+		+					+	+		+
<i>Ceratium falcatum</i> (Kofoid) Jørgensen	+											+		+
<i>Ceratium furca</i> (Ehrenberg) Clapparède & Lachmann		+	+	+				+			+	+	+	+
<i>Ceratium fusus</i> (Ehrenberg) Dujardin	+	+	+	+				+			+	+	+	+
<i>Ceratium gibberum</i> Gourret		+									+	+		+
<i>Ceratium gravidum</i> Gourret												+		
<i>Ceratium hexacanthum</i> Gourret	+	+	+	+							+	+		+
<i>Ceratium horridum</i> (Cleve) Gran		+	+	+							+	+	+	+
<i>Ceratium incisum</i> (Karsten) Jørgensen												+		
<i>Ceratium inflatum</i> (Kofoid) Jørgensen	+	+	+	+							+	+		
<i>Ceratium karstenii</i> Pavillard	+													
<i>Ceratium kofoidii</i> Jørgensen		+	+	+		+				+	+	+	+	+
<i>Ceratium limulus</i> Gourret		+									+	+		
<i>Ceratium lineatum</i> (Ehrenberg) Cleve												+		
<i>Ceratium longirostrum</i> Gourret	+	+	+									+		
<i>Ceratium longissimum</i> (Schröder) Kofoid											+	+		
<i>Ceratium macroceros</i> (Ehrenberg) Vanhöffen	+	+	+	+							+	+		+
<i>Ceratium massiliense</i> (Gourret) Jørgensen	+	+	+	+							+	+	+	
<i>Ceratium paradoxides</i> Cleve											+	+		
<i>Ceratium pavillardii</i> Jørgensen	+													
<i>Ceratium pentagonum</i> Gourret		+	+								+	+		+
<i>Ceratium platycorne</i> Daday												+		
<i>Ceratium ranipes</i> Cleve		+	+								+	+		+
<i>Ceratium schroeteri</i> Schröder												+		
<i>Ceratium symmetricum</i> Pavillard		+	+								+	+	+	+
<i>Ceratium teres</i> Kofoid		+	+	+							+	+	+	+
<i>Ceratium trichoceros</i> (Ehrenberg) Kofoid	+	+	+	+		+					+	+	+	+
<i>Ceratium tripos</i> (O.F. Müller) Nitzsch		+	+	+							+	+		+
<i>Ceratocorys armata</i> (Schütt) Kofoid		+										+		
<i>Ceratocorys gourreti</i> Paulsen		+	+								+	+		
<i>Ceratocorys horrida</i> Stein	+	+	+	+							+	+		+
<i>Citharistes regius</i> Stein					+							+		
<i>Cladopyxis caryophyllum</i> (Kofoid) Pavillard											+	+		

Table 3. (Contd.).

Species	Gökalp (1972)	Polat <i>et al.</i> , (2000)	Polat & Işık (2002)	Polat & Piner (2002)	Polat (2004)	Polat <i>et al.</i> , (2005)	Aka & Polat (2006)	Polat <i>et al.</i> , (2006)	Polat (2007a)	Polat (2007b)	Polat (2007c)	Polat & Koray (2007)	Polat (2008)	This research
<i>Dinophysis amandula</i> (Balech) Sournia														+
<i>Dinophysis caudata</i> Saville-Kent		+	+	+				+				+	+	+
<i>Dinophysis diegensis</i> Kofoid														+
<i>Dinophysis doryphorum</i> (Stein) Abé		+										+	+	+
<i>Dinophysis fava</i> (Kofoid & Michener) Abé												+	+	
<i>Dinophysis fortii</i> Pavillard														+
<i>Dinophysis hastata</i> Stein		+										+	+	+
<i>Dinophysis mitra</i> (Schütt) Abé		+		+								+	+	
<i>Dinophysis odiosa</i> (Pavillard) Tai & Skogsberg				+								+	+	
<i>Dinophysis ovum</i> Schütt												+	+	+
<i>Dinophysis parvula</i> (Schütt) Balech				+								+	+	
<i>Dinophysis rapa</i> (Stein) Balech												+	+	+
<i>Dinophysis recurva</i> Kofoid & Skogsberg				+								+	+	
<i>Dinophysis rotundata</i> Claparède & Lachmann					+			+				+	+	
<i>Dinophysis schuettii</i> Murray & Whitting												+	+	
<i>Dinophysis sphaerica</i> Stein												+	+	
<i>Dinophysis tripos</i> Gourret								+				+	+	
<i>Diplosalis lenticula</i> Bergh		+	+									+	+	+
<i>Diplosalopsis bomba</i> (Stein) Dodge & Toriumi												+	+	
<i>Goniodoma acuminatum</i> (Ehrenberg) Stein												+	+	+
<i>Goniodoma polyedricum</i> (Pouchet) Jørgensen		+	+	+										+
<i>Goniodoma sphaericum</i> Murray & Whitting		+										+	+	
<i>Gonyaulax birostris</i> Stein		+										+	+	
<i>Gonyaulax diegensis</i> Kofoid		+	+									+	+	
<i>Gonyaulax digitalis</i> (Pouchet) Kofoid												+	+	
<i>Gonyaulax monacantha</i> Pavillard		+										+	+	
<i>Gonyaulax pacifica</i> Kofoid									+			+	+	
<i>Gonyaulax polygramma</i> Stein		+	+	+				+				+	+	+
<i>Gonyaulax spinifera</i> (Claparède & Lachmann) Diesing		+	+	+				+				+	+	
<i>Gonyaulax turbynei</i> Murray & Whitting		+						+				+	+	
<i>Gyrodinium fusus</i> (Meunier) Akselman												+	+	
<i>Gyrodinium lachryma</i> (Meunier) Kofoid & Swezy												+	+	
<i>Heterodinium milneri</i> (Murray & Whitting) Kofoid												+	+	
<i>Histioneis elongata</i> Kofoid & Michener												+	+	
<i>Histioneis oxypteris</i> Schiller												+	+	
<i>Kofoidinium velelloides</i> Pavillard		+	+	+								+	+	+
<i>Lingulodinium polyedra</i> (Stein) Dodge		+	+					+				+	+	
<i>Noctiluca scintillans</i> (Macartney) Kofoid								+				+	+	
<i>Ornithocercus francescae</i> (Murray) Balech												+	+	
<i>Ornithocercus heteroporus</i> Kofoid												+	+	+
<i>Ornithocercus magnificus</i> Stein				+	+							+	+	+
<i>Ornithocercus quadratus</i> Schütt		+	+									+	+	+
<i>Ornithocercus splendidus</i> Schütt												+	+	
<i>Ornithocercus steinii</i> Schütt												+	+	+
<i>Ornithocercus thumii</i> (Schmidt) Kofoid & Skogsberg												+	+	
<i>Oxytoxum compressum</i> Kofoid												+	+	
<i>Oxytoxum constrictum</i> (Stein) Bütschli												+	+	
<i>Oxytoxum milneri</i> Murray & Whitting												+	+	
<i>Oxytoxum reticulatum</i> (Stein) Schütt												+	+	
<i>Oxytoxum sceptrum</i> (Stein) Schröder												+	+	
<i>Oxytoxum scolopax</i> Stein		+	+	+								+	+	
<i>Phalacroma argus</i> Stein												+	+	
<i>Phalacroma minutum</i> Cleve												+	+	

Table 3. (Contd.).

Species	Gökalp (1972)	Polat <i>et al.</i> , (2000)	Polat & Işık (2002)	Polat & Piner (2002)	Polat (2004)	Polat <i>et al.</i> , (2005)	Aka & Polat (2006)	Polat <i>et al.</i> , (2006)	Polat (2007a)	Polat (2007b)	Polat (2007c)	Polat & Koray (2007)	Polat (2008)	This research
<i>Podolampas bipes</i> Stein		+	+	+							+	+		+
<i>Podolampas elegans</i> Schütt												+		
<i>Podolampas palmipes</i> Stein											+	+		
<i>Podolampas spinifera</i> Okamura		+	+	+							+	+		
<i>Prorocentrum compressum</i> (Bailey) Abé <i>ex</i> Dodge		+	+					+			+	+	+	
<i>Prorocentrum gracile</i> Schütt														+
<i>Prorocentrum micans</i> Ehrenberg		+	+	+		+		+			+	+	+	
<i>Prorocentrum triestinum</i> Schiller														+
<i>Protoceratium areolatum</i> Kofoid												+		
<i>Protoperdinium brochii</i> (Kofoid & Swezy) Balech		+	+								+	+		
<i>Protoperdinium claudicans</i> (Paulsen) Balech		+									+	+		
<i>Protoperdinium conicoides</i> (Paulsen) Balech												+		
<i>Protoperdinium conicum</i> (Gran) Balech		+	+	+						+	+	+		+
<i>Protoperdinium crassipes</i> (Kofoid) Balech											+	+		
<i>Protoperdinium depressum</i> (Bailey) Balech		+	+	+				+			+	+	+	
<i>Protoperdinium diabolus</i> (Cleve) Balech											+	+		
<i>Protoperdinium divergens</i> (Ehrenberg) Balech		+	+	+							+	+	+	+
<i>Protoperdinium globulus</i> (Stein) Balech		+	+	+							+	+		+
<i>Protoperdinium grande</i> (Kofoid) Balech		+										+		
<i>Protoperdinium granii</i> (Ostenfeld) Balech											+	+		
<i>Protoperdinium leonis</i> (Pavillard) Balech		+										+		
<i>Protoperdinium mediterraneum</i> (Kofoid) Balech		+	+	+							+	+		+
<i>Protoperdinium mite</i> (Pavillard) Balech												+		
<i>Protoperdinium oblongum</i> (Aurivillius) Parke & Dodge												+		
<i>Protoperdinium oceanicum</i> (Vanhöffen) Balech		+	+	+								+		+
<i>Protoperdinium oviforme</i> (P. Dangeard) Balech												+		
<i>Protoperdinium ovum</i> (Schiller) Balech											+	+	+	
<i>Protoperdinium pallidum</i> (Ostenfeld) Balech		+									+	+		
<i>Protoperdinium pedunculatum</i> (Schütt) Balech		+	+	+								+		
<i>Protoperdinium pellucidum</i> Bergh				+	+			+			+	+		
<i>Protoperdinium pentagonum</i> (Gran) Balech												+		
<i>Protoperdinium punctulatum</i> (Paulsen) Balech												+		
<i>Protoperdinium pyriforme</i> (Paulsen) Balech		+	+								+	+		
<i>Protoperdinium quarnerense</i> (Schröder) Balech		+	+								+	+		
<i>Protoperdinium steinii</i> (Jørgensen) Balech		+	+	+		+					+	+	+	+
<i>Protoperdinium subinermis</i> (Paulsen) Loeblich III				+							+	+		
<i>Pyrocystis elegans</i> Pavillard												+		+
<i>Pyrocystis fusiformis</i> (Wyville Thomson <i>ex</i> Haeckel) Blackman		+										+		+
<i>Pyrocystis hamulus</i> Cleve												+		+
<i>Pyrocystis noctiluca</i> J. Murray <i>ex</i> Haeckel		+												
<i>Pyrocystis robusta</i> Kofoid												+		
<i>Pyrophacus horologium</i> Stein		+										+		+
<i>Pyrophacus steinii</i> (Schiller) Wall & Dale		+	+	+				+			+	+		
<i>Scropsiella trochoidea</i> (Stein) Loeblich III			+	+		+		+		+	+	+	+	
<i>Spiraulax jolliffei</i> (Murray & Whitting) Kofoid		+	+								+	+		+
Dictyochophyceae														
<i>Dictyocha fibula</i> Ehrenberg		+	+	+				+			+			+
<i>Dictyocha speculum</i> Ehrenberg				+										
Bacillariophyceae														
<i>Amphiprora gigantea</i> Grunow		+					+				+			
<i>Amphiprora</i> sp.			+	+										
<i>Amphora ovalis</i> (Kützing) Kützing														+
<i>Asterionellopsis glacialis</i> (Castracane) Round		+	+	+			+	+			+			

Table 3. (Contd.).

Species	Gökalp (1972)	Polat <i>et al.</i> , (2000)	Polat & Işık (2002)	Polat & Piner (2002)	Polat (2004)	Polat <i>et al.</i> , (2005)	Aka & Polat (2006)	Polat <i>et al.</i> , (2006)	Polat (2007a)	Polat (2007b)	Polat (2007c)	Polat & Koray (2007)	Polat (2008)	This research
<i>Asterolampra grevillei</i> (Wallich) Greville	+	+									+			
<i>Asterolampra marylandica</i> Ehrenberg		+	+				+				+			+
<i>Asteromphalus flabellatus</i> (Brébisson) Greville		+	+								+			
<i>Asteromphalus heptactis</i> (Brébisson) Ralf in Pritchard			+								+			
<i>Asteromphalus hookeri</i> Ehrenberg							+							
<i>Bacillaria paxillifera</i> (O.F.Müller) Hendey		+	+	+				+			+			+
<i>Bacteriastrum delicatulum</i> Cleve		+	+	+				+			+			+
<i>Bacteriastrum elegans</i> Pavillard											+			+
<i>Bacteriastrum hyalinum</i> Lauder		+	+				+				+			
<i>Biddulphia pulchella</i> Gray			+											
<i>Campylodiscus</i> sp.		+	+								+			
<i>Cerataulina pelagica</i> (Cleve) Hendey		+	+	+			+	+		+	+		+	+
<i>Chaetoceros affinis</i> Lauder	+	+	+	+			+				+		+	+
<i>Chaetoceros anastomosans</i> Grunow		+					+				+			
<i>Chaetoceros atlanticus</i> Cleve		+	+	+			+	+			+		+	
<i>Chaetoceros brevis</i> Schütt							+							
<i>Chaetoceros coarctatus</i> Lauder		+												
<i>Chaetoceros compressus</i> Lauder		+	+	+			+				+			
<i>Chaetoceros constrictus</i> Gran		+					+				+			
<i>Chaetoceros costatus</i> Pavillard		+	+				+				+			
<i>Chaetoceros curvisetus</i> Cleve		+	+	+							+			
<i>Chaetoceros dadayi</i> Pavillard			+	+			+				+			+
<i>Chaetoceros danicus</i> Cleve		+					+							+
<i>Chaetoceros debilis</i> Cleve		+												
<i>Chaetoceros decipiens</i> Cleve		+	+	+			+				+		+	+
<i>Chaetoceros didymus</i> Ehrenberg		+	+	+			+	+			+		+	+
<i>Chaetoceros diversus</i> Cleve		+	+				+				+		+	+
<i>Chaetoceros eibenii</i> Grunow in Van Heurck							+				+			
<i>Chaetoceros holsaticus</i> Schütt							+							
<i>Chaetoceros laciniosus</i> Schütt		+	+	+			+				+			
<i>Chaetoceros lorenzianus</i> Grunow		+	+	+			+				+			+
<i>Chaetoceros messanensis</i> Castracane		+												
<i>Chaetoceros neogracilis</i> VanLand		+												
<i>Chaetoceros peruvianus</i> Brightwell		+	+	+			+				+			+
<i>Chaetoceros pseudocurvisetus</i> Mangin		+	+				+	+			+			
<i>Chaetoceros rostratus</i> Lauder		+												+
<i>Chaetoceros teres</i> Cleve											+			
<i>Chaetoceros tetrastichon</i> Cleve		+	+	+			+				+		+	
<i>Chaetoceros tortissimus</i> Gran		+	+				+				+			
<i>Chaetoceros wighamii</i> Brightwell							+							
<i>Climacosphaenia moniligera</i> Ehrenberg		+	+	+			+				+			
<i>Coscinodiscus centralis</i> Ehrenberg											+			
<i>Coscinodiscus perforatus</i> Ehrenberg		+	+				+							
<i>Coscinodiscus radiatus</i> Ehrenberg		+	+				+				+			+
<i>Cylindrotheca closterium</i> (Ehrenberg) Reimann & Lewin		+	+	+			+	+			+			+
<i>Cymbella</i> sp.							+							
<i>Dactyliosolen antarcticus</i> Castracane														+
<i>Dactyliosolen fragilissimus</i> (Bergon) Hasle		+		+			+							
<i>Diploneis bombus</i> (Ehrenberg) Ehrenberg ex Cleve														+
<i>Ditylum brightwellii</i> (T. West) Grunow in Van Heurck		+									+			
<i>Eucampia zoodiacus</i> Ehrenberg		+	+	+			+	+			+			+
<i>Fragilaria</i> sp.							+							
<i>Grammatophora marina</i> (Lyngbye) Kützing							+				+			
<i>Grammatophora</i> sp.		+												
<i>Guinardia delicatula</i> (Cleve) Hasle							+							
<i>Guinardia flaccida</i> (Castracane) H. Peragallo		+	+	+		+	+	+			+		+	
<i>Guinardia striata</i> (Stoltherfoth) Hasle		+	+	+		+	+				+		+	+

Table 3. (Contd.).

Species	Gökaltın (1972)	Polat <i>et al.</i> , (2000)	Polat & Işık (2002)	Polat & Piner (2002)	Polat (2004)	Polat <i>et al.</i> , (2005)	Aka & Polat (2006)	Polat <i>et al.</i> , (2006)	Polat (2007a)	Polat (2007b)	Polat (2007c)	Polat & Koray (2007)	Polat (2008)	This research
<i>Gyrosigma balticum</i> (Ehrenberg) Rabenhorst		+	+				+				+			
<i>Gyrosigma spencerii</i> (Quekett) Griffith & Henfrey		+												
<i>Gyrosigma tenuissimum</i> (W. Smith) Griffith & Henfrey		+					+							
<i>Helicotheca tamesis</i> (Shrubsole) Ricard		+									+			
<i>Hemiaulus hauckii</i> Grunow in Van Heurck		+	+	+		+	+				+		+	+
<i>Hemiaulus membranaceus</i> Cleve		+	+	+			+				+			
<i>Hemiaulus sinensis</i> Greville			+				+				+			
<i>Lauderia annulata</i> Gran		+					+							
<i>Leptocylindrus danicus</i> Cleve		+	+	+			+	+			+			
<i>Leptocylindrus mediterraneus</i> (H. Peragallo) Hasle		+	+								+			+
<i>Leptocylindrus minimus</i> Gran		+	+	+			+				+			+
<i>Licmophora abbreviata</i> Agardh		+	+	+							+			
<i>Lithodesmium undulatum</i> Ehrenberg		+	+				+				+			+
<i>Melosira moniliformis</i> (Müller) Agardh							+				+			
<i>Navicula transitans</i> Cleve														+
<i>Navicula</i> sp.		+	+	+			+				+			
<i>Nitzschia longissima</i> (Brébisson in Kützing) Ralfs in Pritchard	+	+	+	+			+				+			+
<i>Nitzschia lorenziana</i> Grunow in Cleve & Grunow														+
<i>Nitzschia sigma</i> (Kützing) W. Smith		+	+								+			+
<i>Nitzschia sigmaidea</i> (Nitzsch) W. Smith							+							
<i>Odontella aurita</i> (Lyngbye) Agardh							+	+			+			+
<i>Odontella mobiliensis</i> (J.W. Bailey) Grunow		+	+	+			+				+			+
<i>Paralia sulcata</i> (Ehrenberg) Cleve		+		+							+			
<i>Petrodictyon gemma</i> (Ehrenberg) D.G. Mann														+
<i>Pleurosigma elongatum</i> W. Smith		+	+	+			+				+			
<i>Pleurosigma normanii</i> Ralfs in Pritchard		+	+	+			+				+			+
<i>Podocystis perrinensis</i> Ricard				+							+			
<i>Podocystis</i> sp.		+												
<i>Proboscia alata</i> (Brightwell) Sundström		+	+	+		+	+	+		+	+		+	+
<i>Pseudo-nitzschia delicatissima</i> (Cleve) Heiden			+				+				+			
<i>Pseudo-nitzschia pseudodelicatissima</i> (Hasle) Hasle														+
<i>Pseudo-nitzschia pungens</i> (Grunow ex Cleve) Hasle		+	+	+			+	+			+			
<i>Pseudosolenia calcar-avis</i> (Schultze) Sundström	+	+	+	+			+				+		+	+
<i>Rhabdonema adriaticum</i> Kützing		+		+			+				+			+
<i>Rhizosolenia castracanei</i> H. Peragallo	+	+												
<i>Rhizosolenia habetata</i> (Bail) Gran	+							+			+			
<i>Rhizosolenia imbricata</i> Brightwell	+	+	+	+			+				+			+
<i>Rhizosolenia robusta</i> Norman in Pritchard		+	+	+			+				+			
<i>Rhizosolenia setigera</i> Brightwell		+	+				+	+			+			
<i>Rhizosolenia styliformis</i> Brightwell		+	+	+			+				+			+
<i>Skeletonema costatum</i> (Greville) Cleve		+									+			
<i>Stephanopyxis</i> sp.			+											
<i>Striatella unipunctata</i> (Lyngbye) Agardh		+	+				+				+			+
<i>Surirella fastuosa</i> (Ehrenberg) Kützing		+	+								+			
<i>Surirella</i> sp.				+										
<i>Synedra ulna</i> (Nitzsch) Ehrenberg		+	+	+			+				+			
<i>Thalassionema frauenfeldii</i> (Grunow) Hallegraeff	+	+	+	+		+	+			+	+		+	+
<i>Thalassionema nitzschioides</i> (Grunow) Van Heurck	+	+	+	+				+			+			+
<i>Thalassiophysa hyalina</i> (Greville) Paddock & Sims			+											
<i>Thalassiosira nordenskiöldii</i> Cleve								+						
<i>Thalassiothrix longissima</i> Cleve & Grunow														+
<i>Thalassiothrix mediterranea</i> Pavillard		+	+	+			+				+		+	
<i>Toxarium undulatum</i> Bailey			+	+							+			
<i>Triceratium</i> sp.			+											
Prasinophyceae														
<i>Halosphaera viridis</i> Schmitz		+												
Prymnesiophyceae														
<i>Anaplosolenia brasiliensis</i> (Lohmann) Deflandre				+							+			
<i>Scyphosphaera apsteinii</i> Lohmann				+							+			

Dinoflagellates (with 148 taxa) and diatoms (with 115 taxa) were dominant in terms of diversity. According to the records of adjacent regions, in northwestern Mediterranean, diatoms and dinoflagellate species represented, respectively, 51% and 36% of the total phytoplankton species (Velasquez & Cruzado, 1995). In the northeastern Mediterranean coast of Turkey, diatoms represented 57.4% and dinoflagellates 37.2% (Polat & Piner, 2002). In the Aegean Sea, diatoms and dinoflagellate species were represented by 45.8% and 41.2% (Koray, 1994). In this research, our data showed that diatoms and dinoflagellate species were represented by 43% and 55% respectively of the total species composition of microphytoplankton. In previous studies, similar results were reported such as 43% of diatoms and 52% of dinoflagellates in Villefranche Bay in northwestern Mediterranean (Gomez & Gorsky, 2003), 31% of diatoms and 48% of dinoflagellates in the Gulf of Genoa (Bernhard & Rampi, 1967), 40% of diatoms and 52% of dinoflagellates in the Sea of Marmara (Balkis, 2003) and 47% of diatoms and 50% of dinoflagellates in the coastal zone of Bozcaada Island (Balkis, 2009). The oligotrophic conditions result in a high diversity of dinoflagellates in the Mediterranean (Gomez, 2003). It was reported that water temperature in the Mediterranean has increased due to global warming and local influences (Turley, 1999) and that increase might have resulted in alterations in the marine biodiversity (Gomez & Claustre, 2003).

In terms of the qualitative distribution of phytoplankton, it can be said that most of the species listed in the present study consisted of phytoplanktonic algae reported in the different regions of the Mediterranean earlier. Most of the species in this Gulf were neritic, oceanic, temperate and subtropical species. In fact, marine species (*P. alata*, *T. nitzschioides* and *C. fusus*), brackish species (*C. closterium*) and some benthic species (*P. normanii* and *N. transitans*) which have adapted to plankton life were observed.

The most common genus from Dinophyceae was *Ceratium* with 40 species. Graham (1941) stated that the geographical distribution of *Ceratium* species were especially affected by temperature. During the sampling period of the present study, members of the *Ceratium* genus, *C. candelabrum* was observed at all stations (f=100) and *C. tripos* was observed at eight stations (f=89). According to Polat (1997), these species were not affected significantly by temperature changes during a year while Koray & Gökpinar (1983) categorized these species as species that live in habitats with a temperature between 12 and 26°C. In this study, it was determined that the *Ceratium* species lived in habitats with a temperature between 20.2 and 28.7°C. The temperature range in the present study is narrower than those of previous studies since the sampling was performed only in two seasons (November 2005, June 2006).

Pseudosolenia calcar-avis and *Hemiaulus hauckii* are distinctive species of oligotrophic waters that are poor of nutrients (Kimor, 1985). These species were encountered

during both sampling periods. According to the study conducted in the gulf, *Pseudosolenia calcar-avis* do not reproduce excessively during any of the sampling periods and *Hemiaulus hauckii* continue to maintain its existence in water environment also during the periods following the spring diatom increases (Polat, 1997).

It was reported that the phytoplankton species recorded in world seas, nearly 300 of them caused coloration on the surface of the seas by reproducing excessively and nearly 60 to 80 species negatively affected human health by producing toxins (Hallegraeff, 1993; Van Dolah, 2000). Since these species damage the coastal ecosystems of many countries and are called "Harmful Algal Bloom Species (HABs)" in the international literature. The frequency of harmful algal blooms, which are defined as the local, sudden and excessive growth of one or a small number of phytoplankton species in seas, has increased in recent years. This is generally caused by such factors as the increase of nutrient input into seas due to domestic, agricultural and industrial activities and seasonal changes. As a result of this event, which called "Harmful Algal Bloom", mass deaths are seen in the fauna, fishing and tourism are negatively affected, and the quality of water changes (Hallegraeff, 1993; Vila & Masó, 2005). Dinoflagellates and diatoms are the two most important groups that produce toxin. In the present study, of the 18 species that have excessive algal growth potential three produce toxin. Since the species were not examined quantitatively in the sampling, information about the abundance of these species is not available. In a study in the Gulf of İskenderun, Polat *et al.*, (2006) identified 39 phytoplankton species that caused excessive growth and reported that of these 4 species (*Dinophysis tripos*, *D. caudata*, *D. rotundata*, *Pseudo-nitzschia pungens*) were toxin-producing species.

Plant forms cause oxygen enrichment in surface waters as a result of photosynthesis. As seen in the present study, the oxygen values of surface waters which continuously interact with the atmosphere are high. In addition, it was found that the oxygen values in November when the temperature was lower compared to June were also higher than the oxygen values in June. In a study, Polat (1997) determined the highest dissolved oxygen value in April (8.6 mg l⁻¹) and the lowest in June (4.8 mg l⁻¹), and also noted that the oxygen content did not decrease to lower levels during the study. She explained that the reason for the higher value measured in April was the increase in the cell numbers of phytoplankton species and the reason for the lower value measured in summer months was the increasing temperature. The oxygen content in water changes depending on the water movements and the biological occurrences in the environment. Open sea waters rich in oxygen flow into the Gulf of İskenderun, circulates in the gulf and then flow out. Therefore, the Gulf of İskenderun has a quite dynamic structure (Polat, 1997).

To conclude, in the present study the phytoplankton species that are distributed in the Gulf of İskenderun and

the potentially harmful ones among these species were identified and a new species record was added to the phytoplankton species diversity in Turkish seas. The Mediterranean Sea is exposed to the migration of Red Sea species through the Suez Canal and to the passage of species from the Atlantic Ocean through the Strait of Gibraltar (Polat & Koray 2007). There is need for further research on this topic in order to identify the species that enter Turkish waters.

References

- Aka (Akiz), A. and S. Polat. 2006. Karataş kıyısı suları (Kuzey Doğu Akdeniz) planktonik diatomları, *Ege Üniversitesi Su Ürünleri Dergisi*, 23(1/1): 1-7.
- Aktan, Y. 2011. Large-scale patterns in summer surface water phytoplankton (except picophytoplankton) in the Eastern Mediterranean. *Estuarine, Coastal and Shelf Science*, 91: 551-558.
- Azov, Y. 1986. Seasonal patterns of phytoplankton productivity and abundance in nearshore oligotrophic waters of the Levant Basin (Mediterranean). *J. Plankton Res.*, 8(1): 41-53.
- Balkis, N. 2003. Seasonal variations in the phytoplankton and nutrient dynamics in the neritic water of Büyükçekmece Bay, Sea of Marmara. *J. Plankton Res.*, 25(7): 703-717.
- Balkis, N. 2005. Contributions to the knowledge of marine phytoplankton of Turkey. *Pak. J. Bot.*, 37(4): 807-814.
- Balkis, N. 2009. Seasonal variations of microphytoplankton assemblages and environmental variables in the coastal zone of Bozcaada Island in the Aegean Sea (NE Mediterranean Sea). *Aquat. Ecol.*, 43: 249-270.
- Bernhard, M. and L. Rampi. 1967. The annual cycle of the Utermöhl-phytoplankton in the Ligurian Sea in 1959 and 1962. *Pubbl. Staz. Zool. Napoli.*, 35: 137-169.
- Drira, Z., A. Hamza, M. Bel Hassen, H. Ayadi, A. Bouain and L. Aleya. 2010. Coupling of phytoplankton community structure to nutrients, ciliates and copepods in the Gulf of Gabès (South Ionian Sea, Tunisia). *J. Mar. Biol. Ass. U.K.*, 90: 1203-1215.
- Dugdale, R.C. and F.P. Wilkerson. 1988. Nutrient sources and primary production in the eastern Mediterranean, *Oceanol. Acta*, 9: 179-184.
- Faust, M.A. and R.A. Gulledge. 2002. Identifying harmful marine dinoflagellates. Contributions from the United States National Herbarium, 42: 1-144.
- Gazeau, F., S.V. Smith, B. Gentili, M. Frankignoulle and J.P. Gattuso. 2004. The European coastal zone: characterization and first assessment of ecosystem metabolism. *Estuarine Coastal and Shelf Science*, 60: 673-694.
- Gomez, F. 2003. Checklist of Mediterranean free-living dinoflagellates. *Bot. Mar.*, 46: 215-242.
- Gomez, F. 2005. A list of free-living dinoflagellate species in the world's oceans. *Acta Bot. Croat.*, 64(1): 129-212.
- Gomez, F. and H. Claustre. 2003. The genus *Asterodinium* (Dinophyceae) as a possible biological indicator of warming in the western Mediterranean. *J. Mar. Biol. Ass. U.K.*, 83: 173-174.
- Gomez, F. and G. Gorsky. 2003. Annual microplankton cycles in Villefranche Bay, Ligurian Sea, NW Mediterranean. *J. Plankton Res.*, 25(4): 323-339.
- Gökalp, N. 1972. A study of plankton conditions of Edremit, Bodrum and İskenderun Gulf. Publications of the Hydrobiological Research Institute, Faculty of Science, University of Istanbul, 3: 1-71.
- Graham, H.W. 1941. An oceanographic consideration of the dinoflagellate genus *Ceratium*. *Ecol. Monogr.*, 5(2): 99-116.
- Hallegraeff, G.M. 1993. A review of harmful algal blooms and their apparent global increase. *Phycologia*, 32(2): 79-99.
- Hasle, G.R. and E.E. Syvertsen. 1997. Marine diatoms. In: *Identifying marine phytoplankton*. (Ed.): C.R. Tomas. Academic Press a division of Harcourt Brace and Company, San Diego, USA, chapter 2, pp. 5-385.
- Ignatiades, L., S. Psarra, V. Zervakis, K. Pagou, E. Souvermezoglou, G. Assimakopoulou and O. Gotsis-Skretas. 2002. Phytoplankton size-based dynamics in the Aegean Sea (Eastern Mediterranean). *J. Mar. Syst.*, 36: 11-28.
- Ivanoff, A. 1972. Introduction al'océanographie. Tome I. Librairie Vuibert, Paris.
- Kimor, B. 1985. Round table on indicator species in marine phytoplankton, II. Background Presentation, *Rapp. Comm. Int. Mer Médit.*, 29(9): 65-79.
- Koray, T. 1994. Phytoplankton species succession, diversity and nutrients in neritic waters of the Aegean Sea (Bay of Izmir). *Tr. J. of Botany*, 19: 531-544.
- Koray, T. and Ş. Gökımar. 1983. *Ceratium* Schrank genusü türlerinin İzmir Körfezi'ndeki kalitatif ve kantitatif özellikleri, I. Ulusal Deniz ve Tatlısu Araştırmaları Kongresi tebliğleri, *Ege Üniversitesi Fen Fakültesi Dergisi*, I: 178-200.
- Krom, M.D., N. Kress, S. Brenner and L.I. Gordon. 1991. Phosphorus limitation of primary production in the eastern Mediterranean Sea. *Limnol. Oceanogr.*, 36: 424-432.
- Lakkis, S. and V. Lakkis. 1980. Composition, annual cycle and species diversity of phytoplankton in Lebanese coastal water. *J. Plankton Res.*, 3(1): 123-135.
- Lu, S. and I.J. Hodgkis. 2004. Harmful algal bloom causative collected from Hong Kong waters. *Hydrobiologia*, 512: 231-238.
- MarBEF Data System-European of Marine Species (ERMS) (<http://www.marbef.org/data/erms.php>) (2010).
- Polat, S. 1997. İskenderun Körfezi'nin Yumurtaalık Toros Gübre arasındaki kıyı bölgesinde fitoplankton dağılımı, yoğunluğu ve bunların zamana bağlı değişimleri, Ç. Ü. Fen Bilimleri Enstitüsü Su Ürünleri Anabilim Dalı, PhD. Thesis.
- Polat, S. 2002. Nutrients, chlorophyll *a* and phytoplankton in the İskenderun Bay (Northeastern Mediterranean). *P.S.Z.N.: Marine Ecology*, 23(2): 115-126.
- Polat, S. 2004. New record for a dinoflagellate species (*Citharistes regius* Stein) in the Northern Levantine Basin (Eastern Mediterranean). *Turk. J. Bot.*, 28: 507-509.
- Polat, S. 2007a. New record for a dinoflagellate species (*Gonyaulax pacifica* Kofoid) from Turkish coastal waters (Northeastern Mediterranean Sea). *Turk. J. Bot.*, 31: 67-70.
- Polat, S. 2007b. Effects of nutrient enrichment on coastal phytoplankton composition and abundance in the northeastern Mediterranean. *Pak. J. Bot.*, 39(6): 2087-2095.
- Polat, S. 2007c. Seasonal dynamics of phytoplankton in a coastal marine ecosystem: İskenderun Bay, Northeastern Mediterranean Sea. *Fresenius Environmental Bulletin*, 16(7): 756-763.
- Polat S. 2008. The distribution and seasonal dynamics of diatom and dinoflagellates in the İskenderun Bay (Northeastern Mediterranean). *Journal of Fisheries Sciences.com*, 2(2): 153-163.
- Polat, S. and O. Işık. 2002. Phytoplankton distribution, diversity and nutrients at the north-eastern Mediterranean coast of Turkey (Karataş-Adana), *Turk. J. Bot.*, 26: 77-86.

- Polat, S. and M.P. Piner. 2002. Nutrients and phytoplankton in the Babadillimani Bight, northeastern Mediterranean coast of Turkey. *Indian Journal of Marine Sciences*, 31(3): 188-194.
- Polat, S. and T. Koray. 2007. Planktonic dinoflagellates of the northern Levantine Basin, northeastern Mediterranean Sea. *European Journal of Protistology*, 43: 193-204.
- Polat, S., E. Sarihan and T. Koray. 2000. Seasonal changes in the phytoplankton of the northeastern Mediterranean (Bay of İskenderun). *Turk. J. Bot.*, 24: 1-12.
- Polat, S., A. Akiz and M.P. Olgunoğlu (Piner). 2005. Daily variations of coastal phytoplankton assemblages in summer conditions of the northeastern Mediterranean (Bay of İskenderun). *Pak. J. Bot.*, 37(3): 715-724.
- Polat, S., M.P. Olgunoğlu, A. Akiz Aka and T. Koray. 2006. Kuzeydoğu Akdeniz kıyısız sularında (İskenderun Körfezi) dağılım gösteren potansiyel zararlı fitoplankton türleri, *E. Ü. Su Ürünleri Dergisi*, 23(1-2): 169-172.
- Sorensen, T. 1948. A method of establishing groups of equal amplitude in plant sociology based on similar species content and its application to analyses of vegetation on Danish commons. *Biologiske Skrifter*, 5: 1-34.
- Turley, C.M. 1999. The changing Mediterranean Sea – a sensitive ecosystem? *Prog. Oceanogr.*, 44: 387-400.
- Van Dolah, F.M. 2000. Marine Algal Toxins: Origins, Health Effects and Their Increased Occurrence. *Environmental Health Perspectives Supplements*, 108(S1): 133-141.
- Velasquez, Z.R. and A. Cruzado. 1995. Inventory of the diatom flora of the NW Mediterranean Sea. *Vie Milieu*, 45: 249-263.
- Vila, M. and M. Masó. 2005. Phytoplankton functional groups and harmful algal species in anthropogenically impacted waters of the NW Mediterranean Sea. *Sci. Mar.*, 69(1): 31-45.
- Winkler, L.W. 1888. The determination of dissolved oxygen in water. *Berlin Deut. Chem. Ges.*, 21: 2843-2855.
- Yılmaz, A. and S. Tuğrul. 1998. The effect of cold and warm core eddies on the distribution and stoichiometry of dissolved nutrients in the northeastern Mediterranean, *J. Mar. Syst.*, 16: 253-268.
- Yılmaz, A., Ö. Baştürk, C. Saydam, D. Ediger, K. Yılmaz and E. Hatipoğlu. 1992. Eutrophication in İskenderun Bay, northeastern Mediterranean. *Sci. Total Environ.*, Supplement, 705-717.

(Received for publication 24 March 2011)