

ARABIDOPSIS TRICHOME MODEL ELUCIDATES THE MECHANISM IN COTTON FIBER INITIATION

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Abstract

Cotton is valued for its fiber and requires yield improvement to compete with synthetic textiles and for sustainable cotton production. Cotton fiber production from seed coat epidermal cells can be categorized into two stages: initiation and development. While a great deal of research has been conducted on the cotton fiber elongation and secondary cell wall biosynthesis, fiber initiation understanding is still in its infant stage due to the difficulty in studying the mechanism in cotton, namely lengthy transformation, lack of characterized mutants, and large and complex genomes. The Arabidopsis trichome differentiates from the leaf epidermal cells and presents an excellent model system to elucidate the cotton fiber initiation mechanism. Knowledge gained from the initiation mechanism of *Arabidopsis* trichomes will facilitate, as a comparative model system, in understanding of the cotton fiber initiation mechanisms.

Key words: Trichome, Fiber initiation, Trimeric complex, Diploid, Tetraploid.

Introduction

Cotton (*Gossypium*) fiber is the primary material for a textile industry, and currently, there is an immense interest in understanding the process of fiber initiation and development. With the recently published reference genome sequences of cotton species, the cotton fiber initiation and development has become an essential field of study (Chen *et al.*, 2020). Cotton fibers are unicellular trichomes originating from seed coat epidermal cells. Of these cells, approximately 30% are differentiated into fiber cells resulting in the production of roughly 20,000 fibers/ovule (Berlin, 1986). Increasing the number of fiber initials will result in additional fiber yield, will be benefitted ultimately the cotton producers and allied industries (Patel *et al.*, 2020). A mere 10% increase in initials results in about a 30% increase in the final fiber yield.

Cotton fibers, which are highly elongated and thickened cells, are one of the few cells in the plant kingdom that can significantly expand in size (up to 6.0cm) or composition during growth and development. These fibers, also known as seed trichomes, will quasi-synchronously undergo four distinct yet overlapping stages of development (Guan & Chen, 2013). Fiber initiation stage begins at approximately -3 days post-anthesis (DPA) to 5 DPA, in which ovular epidermal cells emerge and differentiate into fiber initials. Subsequently, from 3 to 21 DPA, morphologically-distinct fiber cells continue to expand up to 6 cm in length without further cell division (Wilkins & Arpat, 2005). From 14 to 40 DPA, a massive amount of cellulose is deposited which is known as the secondary cell wall biosynthesis stage. Finally, fiber cells mature at 50 to 60 DPA, and at this stage, the cotton fibers and seeds are ready for harvesting and industrial applications (Basra & Malik, 1984). Elucidating the molecular mechanism of fiber initiation will provide specific information on the genes involved in epidermal cell differentiation and will facilitate the design of novel genetic and molecular strategies to improve the number of initials, thereby improving cotton fiber yield.

Economic, environmental and scientific importance of cotton: Cotton is an essential raw material used to produce numerous commodities, including textile fabrics, medical

applications, fine paper, computer screens and automobile brakes; it is also used for cooking oil, cattle feed, and biodiesel fuel. Although additional commercial value can be captured from cottonseed and its associated products, the fundamental economic value originates from cotton fiber (Campbell & Hinze, 2010). Of over fifty documented species in the *Gossypium* genus (Wendel, 1989), four species (*G. hirsutum*, *G. barbadense*, *G. arboreum*, *G. herbaceum*) are widely cultivated around the world. They have had a significant impact on global trade and economy (Zhang & Feng, 2000). China, the United States (US), and India produce most of the world's cotton comprising more than 15.9 million metric tons of cotton lint and 30.4 million metric tons of cottonseed, which was approximately equivalent to 22.8 billion and 6 billion dollars, respectively (Bowman *et al.*, 2013). In 2019, these three countries contributed about 13.5 metric tons of cotton (<https://www.statista.com/>), corresponding to 40 to 100 million US dollars annually (Fig. 1). Globally, the economic impact of the cotton industry is estimated to be \$500 billion (US) per year, with more than 100 million families from approximately 150 countries directly or indirectly dependent on cotton crop (Bowman *et al.*, 2013).

Currently, the cotton fiber industry is facing fierce competition from companies producing synthetic textile fibers such as polyester, nylon, and polypropylene. Compared to cotton, synthetic fibers are not environmentally friendly as they are made from fossil fuel sources, are non-biodegradable, hydrophobic, burn and melt quickly posing health risks (<http://www.barnhardtcotton.net/blog/know-fibers-cotton-vs-synthetic-fibers/>). Besides, a recent study showed the presence of synthetic textile fibers in sea fish sold for human consumption (Rochman *et al.*, 2015), thus raising concerns about direct effect on human health. Cotton fiber is not only environmentally friendly, but it also helps to clean environmental pollutants such as oil spills and leaks; for example, 1 gram of raw cotton can absorb 30.5gram of crude oil (Singh *et al.*, 2013). Cotton fiber is one of the most critical cell types on earth, which has scientific, economic, and environmental significance; hence, there is a need to make cotton cultivation more profitable for sustainability and meet the demand of the growing world population.

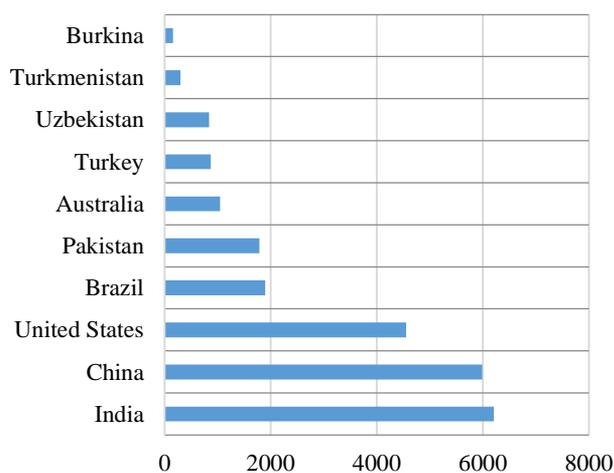


Fig. 1. Leading cotton-producing countries worldwide in 2018/2019 (in 1,000 metric tons). Source: US Department of Agriculture. (www.statista.com)

Phylogeny, genetics, and genomics of cotton:

Approximately 1.5 million years ago, the spontaneous interspecific hybridization and genome duplication event of two formerly independent diploid genomes, ($2n=2x=26$): extant D- genome species closely related to *G. raimondii* (D5), and A- genome species related to *G. arboreum* (A2)/*G. herbaceum* (A1), resulted in allotetraploid species ($2n=4x=52$) (Wendel & Cronn, 2003). The ancestral A-species produce spinnable fibers while the D- species do not. The polyploidization and subsequent evolution resulted in the emergence of six tetraploid species (Figs. 2 and 3). *G. hirsutum* (AD1), *G. barbadense* (AD2), *G. tomentosum* (AD3), *G. mustelinum* (AD4), *G. darwinii* (AD5) and recently described *G. ekmanianum* (AD6) (Grover *et al.*, 2015). Among the six allotetraploids, Upland or American cotton, *G. hirsutum*, represents more than 95% of annual world cotton production, while the remaining 5% is primarily produced from Pima cotton, *G. barbadense*, known for its finer and longer fiber. Diploid cotton is cultivated in South Asia; however, it contributes as little as 2% to total global cotton production (Wendel & Cronn, 2003).

Progress in understanding the role of subgenomes in fiber initiation and development using tetraploid cotton:

Though the ancestral D- diploid genome progenitors do not produce spinnable fibers, the QTL (Quantitative Trait Loci) mapping studies have showed that most of the QTLs influencing fiber quality and yield are located on D^T subgenome of the tetraploid species. In contrast, differential expression of RNA transcripts during early stages of fiber development in tetraploid species show selective enrichment of A^T subgenome specific genes, which is consistent with production of spinnable fibers in ancestral A- diploid species (Samuel *et al.*, 2006). Following this result, systematic mapping of fiber developmental genes in tetraploid species have demonstrated that more genes associated with fiber development are located on A^T subgenome, while the D^T subgenome provides more transcription factors which regulate the expression of the fiber genes in the A^T subgenome (Xu *et al.*, 2010).

In contrast, another study has showed that the ancestral D- genome provides many fiber genes after its hybridization with ancestral A- diploid species (Xu *et al.*, 2015). Additionally, studies using mapped fiber gene-

specific Simple Sequence Repeats (SSRs) indicate that both A^T and D^T subgenomes equally contribute to fiber traits (Han *et al.*, 2006). Overall, the reviews are inconclusive on the subgenome contribution towards fiber development. Moreover, all these studies show expression bias or association of genes with fiber development but do not demonstrate the functional role of these genes. Hence, identification and characterization of individual genes involved in fiber development is essential to understand the specific contribution of different genes.

Since fiber initiation is a result of an interaction of different proteins to form active complex as well as activation of several downstream genes (Guan *et al.*, 2007), a thorough understanding of this mechanism warrants a systematic and dedicated study. Current knowledge of the molecular mechanisms of cotton fiber initiation is in its infancy due to a complex and large genome, polyploidy, gene duplications, recalcitrance to genetic transformation (~1year), long growth cycles and lack of available (genetically characterized) mutants for functional studies (Pang *et al.*, 2013). Of the total seed coat epidermal cells, approximately 30% are differentiated into fiber cells, which further complicate the isolation of pure fiber initial cells for the molecular analysis. To circumvent these complications, the *Arabidopsis* trichome has been successfully employed as a model system for functional characterization of cotton fiber initiation genes.

Arabidopsis trichome initiation is regulated by counteracting positive and negative regulators:

The formation of an active trimeric complex is the prerequisite of trichome initiation process which is the composed of an R2R3-MYB, a basic helix-loop-helix (bHLH), and a WD40 protein abbreviated as MBW complex. Because of its simplicity, flexibility and plasticity, the MBW regulatory complex has been utilized extensively by plants (Ramsay & Glover, 2005). The MBW complex plays diverse roles in *Arabidopsis* such as anthocyanin production, stomatal-cell identity and root-hair formation (Walker *et al.*, 1999). Emerging evidence suggests that the same mechanism as in *Arabidopsis* may control trichome formation in other plant species. For example, MYB-like genes from *Mimulus guttatus* and peach mediate trichome formation (Scoville *et al.*, 2011; Vendramin *et al.*, 2014); ectopic expression of a R3 MYB gene from *Solanum lycopersicum* in *Arabidopsis* results in glabrous phenotypes (Tominaga-Wada *et al.*, 2013).

In *Arabidopsis*, the trichome initiation is positively mediated by a trimeric complex composed of GLABRA1 (GL1) (Oppenheimer *et al.*, 1991), GLABRA3 (GL3), which acts redundantly with its close homolog ENHANCER OF GLABRA3 (EGL3) (Payne *et al.*, 2000), and TRANSPARENT TESTA GLABRA1 (TTG1) (Serna & Martin, 2006). This trimeric activator complex up-regulates the expression of GLABROUS2 (GL2) (Hülkamp, 2004) and a small family of single-repeat MYB proteins lacking the typical transcriptional activation domains, including TRIPTYCHON (TRY) (Schellmann *et al.*, 2002), CAPRICE (CPC), ENHANCER OF TRY & CPC1 (ETC1, 2 and 3) (Tominaga *et al.*, 2008) and TRICHOME-LESS (TCL) (Wang *et al.*, 2007). The GL2 initiates the trichome patterning and differentiation while TRY, CPC, ETC1, 2, 3, TCL are six small, R3-single repeat MYB transcriptional regulators that repress trichome initiation from adjacent cells in a redundant manner (Tominaga *et al.*, 2008; Wester *et al.*, 2009).

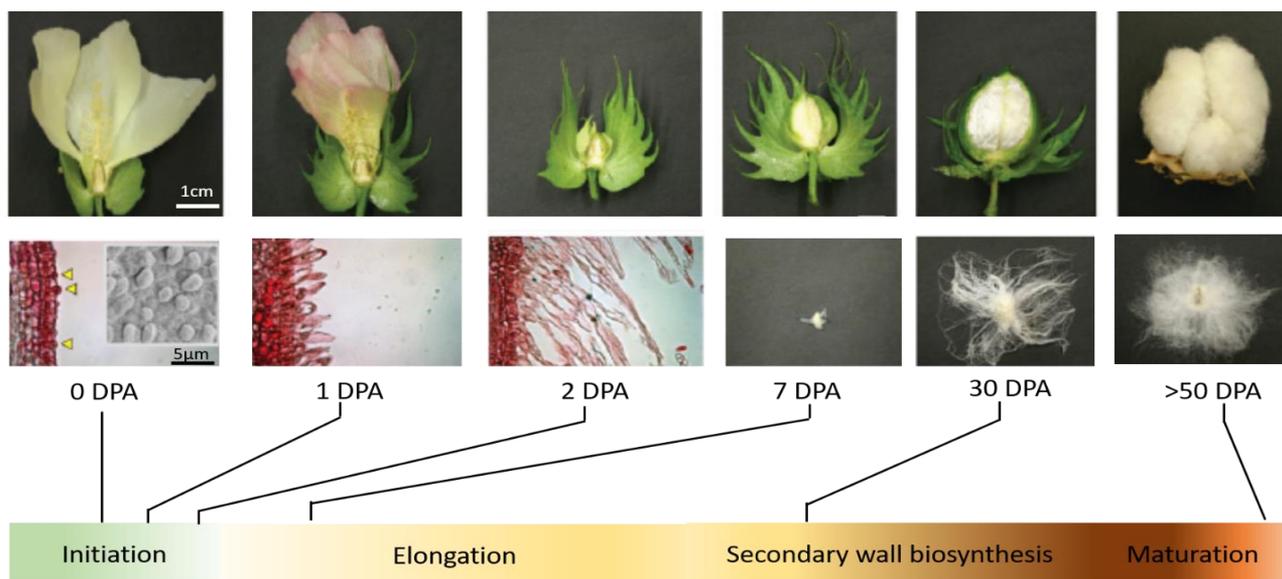


Fig. 2. Cotton (*Gossypium*) fiber initiation and elongation stages (Lee *et al.*, 2007).

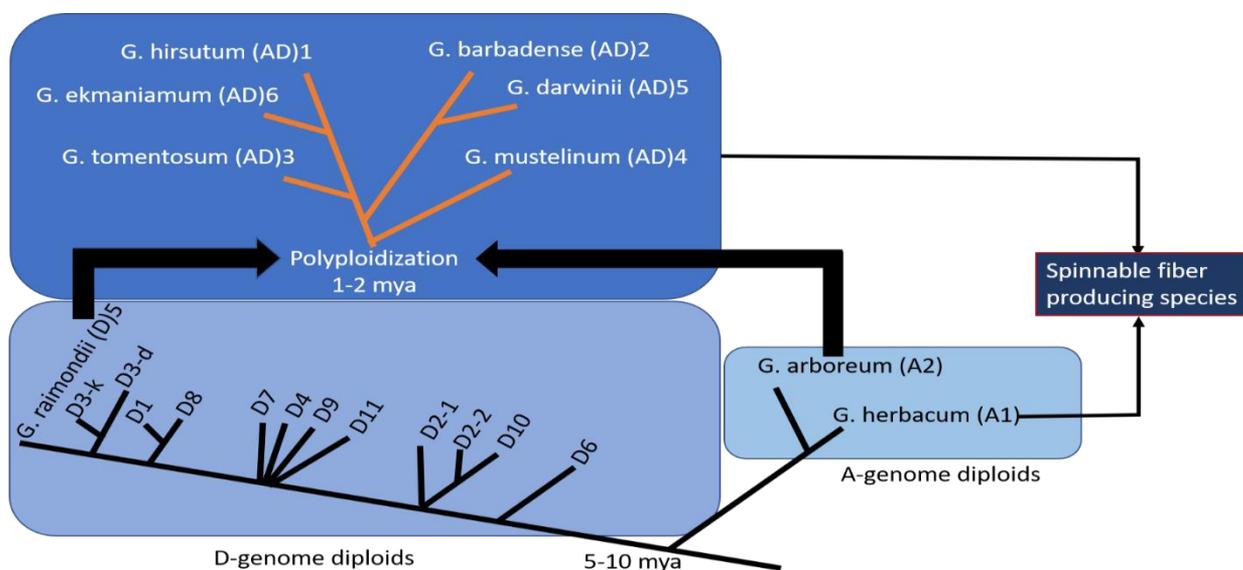


Fig. 3. Phylogenetic framework of cotton (*Gossypium*) diploid and tetraploid species.

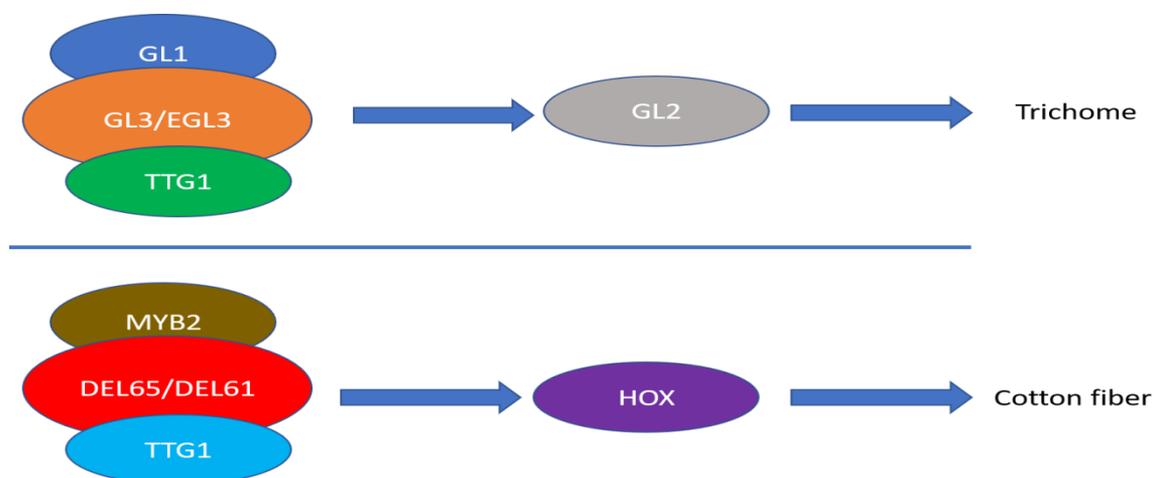


Fig. 4. Two MBW complexes is proposed to initiate Arabidopsis trichome and cotton fiber (by authors) A. Trimeric complex involved in the Arabidopsis trichome initiation. B. Proposed model of trimeric complex involved in cotton fiber initiation.

The regulation of trichome initiation is proposed to be mediated in a spatial and dose-dependent manner (Wester *et al.*, 2009). Once a certain threshold level of activator MBW complex has been reached, expression of downstream regulators will be triggered, including positive regulators of trichome cell fate GL2 and inhibitory R3 MYB proteins (Zhao *et al.*, 2008). The activation of GL2 mediates the trichome formation while the activated inhibitor proteins diffuse to adjacent cells and prevent them from becoming trichomes (lateral inhibition mechanism). The inhibitors, due to their smaller sizes and mobility, can laterally spread to neighboring cells to obstruct assembly of trimeric MBW activator complexes, thus preventing the adjacent cells from forming a trichome cell. The inhibitor proteins (TRY or CPC) prevent the formation of active MBW complex by competing with GL1, thus avoiding the trichome formation (Wester *et al.*, 2009). Overall, Arabidopsis trichome is the most studied cell type with a wealth of information and resources that serves as a useful model system to study the cotton fiber initiation mechanism.

Composition of cotton fiber initiation protein complex:

The current method for rapid characterization of cotton fiber initiation genes is by complementation of cotton homologs in corresponding Arabidopsis mutants and by examining the trichome recovery phenotype (Guan *et al.*, 2008; Guan *et al.*, 2014; Li *et al.*, 2017; Wang *et al.*, 2013) (Fig. 4). Ectopic expression of MYB2 from *G. arboreum*, which is homologous to AtGL1, rescues trichomeless phenotype of the Arabidopsis *gll* mutant, confirming MYB2-A is a functional homolog of AtGL1 (Guan *et al.*, 2014). Additionally, homologs of Arabidopsis GL3, TTG1, CPC, TRY, and GL2 have been isolated from *G. arboreum* (DEL65, TTG1, CPC, TRY, and HOX1, respectively) and functionally characterized using the Arabidopsis trichome model system (Guan *et al.*, 2008; Wang *et al.*, 2013). Formation of active trimeric complex is a prerequisite for the leaf epidermal cell differentiation into trichome in Arabidopsis (Li *et al.*, 2017). Functional characterization and protein-protein interaction of the genes involved in cotton fiber initiation indicate a trimeric protein complex, similar to the Arabidopsis trimeric complex, is involved in cotton fiber initiation (Fig. 2) (Guan *et al.*, 2008; Guan *et al.*, 2014; Wang *et al.*, 2013).

Despite the characterization of individual genes, there is currently no comprehensive understanding of the nature of the cotton trimeric complex and its function. The trichomes initiation follows defined pattern on Arabidopsis leaves while the cotton fibers appear randomly, with no design on the seed coat. The fundamental difference in the patterning mechanism remains unanswered due to lack of suitable tools.

Future research on fiber patterning on cotton fiber initiation: To address this fundamental question, creating a trimeric cotton complex in Arabidopsis is proposed. Currently, Arabidopsis mutant defective for one or two gene(s) is replaced with a cotton homolog for functional characterization of cotton fiber initiation genes. It essentially replaces only one component of Arabidopsis trimeric complex with a cotton homolog while retaining the other parts of the Arabidopsis complex. This approach proved to be highly useful to study individual genes. However, it still reflects the Arabidopsis trimeric complex in its interactions, or complex formation, or activation of downstream genes, which is evident from the patterned trichomes in the

Arabidopsis lines complemented with cotton genes. As a result, creating cotton fiber initiation complex in Arabidopsis without trichome initiation complex will be a novel tool to comprehensively understand the molecular basis for lack of fiber patterning on cotton seed. Comparative studies will be performed with leaf trichome and fuzz fiber systems to understand the intrinsic differences in these systems leading to differential pattern formation within cotton.

Conclusion Remarks

Cotton fiber, also known as seed trichome, is differentiated from the seed coat epidermal cells similar to Arabidopsis leaf trichome, which is differentiated from the leaf epidermal cells. Knowledge gained from the initiation mechanism of Arabidopsis trichomes will facilitate, as a comparative model system in understanding of the cotton fiber initiation mechanisms. Despite functional characterization of individual cotton fiber initiation genes, currently, there is not a comprehensive understanding of the mechanism behind cotton fiber initiation. Though there is a great deal of resemblance in initiation mechanism, there is a fundamental difference in the pattern formation of Arabidopsis trichomes and cotton fibers. The trichomes are well patterned on Arabidopsis leaves due to the lateral inhibition mechanism (Schellmann *et al.*, 2002), while there is no apparent pattern in fiber formation on cotton seed. We aim to address the fundamental differences in the pattern formation by developing a novel tool, cotton trimeric complex in Arabidopsis. The mechanistic studies will have broader implications in fiber production as they will have tremendous applications in improving the fiber yield.

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