

**IN SILICO EXPLOITATION OF ANTIVIRAL PHYTOCHEMICALS AGAINST DENGUE****FAISAL NOUROZ<sup>1,2</sup>, MADIHA MEHBOOB<sup>1</sup>, TIBGHA MOBIN<sup>3</sup> AND SAJID KHAN<sup>1</sup>**<sup>1</sup>*Department of Bioinformatics, Hazara University Mansehra, Pakistan*<sup>2</sup>*Department of Botany, Hazara University Mansehra, Pakistan*<sup>3</sup>*Department of Biotechnology, COMSATS University Islamabad, Abbottabad Campus, Pakistan**\*Corresponding author's email: faisalnouroz@gmail.com***Abstract**

Phytochemicals are plants natural compounds, ethno-pharmacologically used as anti-hepatitis, anti-cancer, anti-diabetic, anti-fungal and anti-viral agents. Computer-aided drug designing (CADD) is flourishing technique nowadays and various natural plant products or phytochemicals are exploited to design potential drugs against various diseases. Dengue fever is caused by the dengue virus (DENV) and is a tropical mosquito-borne infectious disease. DENV infections are among the most important mosquito-infected diseases that are widespread in subtropical and tropical countries (>100). Currently, the dengue fever is becoming a serious health risk. There is need of treatment in contrary to the growing issues of dengue fever and the presence of resistant mutants of DENV in the world. In present research, potential and beneficial anti-dengue drug from plants natural compounds is proposed using CADD, which requires the utilization of information (Biochemical) of ligand-receptor interaction in turn to assume drug treatments. To find biochemical information, docking interactions of selected ligands with E-glycoprotein (4UT6) active site was done which showed effective results against the DENV. Ligand-receptor docking were interpreted by different interactions such as hydrogen bonding, ionic and hydrophobic interactions. Based on the interaction assessment and binding energies of the compounds, one was considered as a "lead compound". Six compounds were placed with in the active site of E-glycoprotein (PDB id: 4UT6). Ligands interaction with the active site of E-glycoprotein was assessed. On the bases of high binding interactions and activity, the assessed drug compounds will be proposed in the laboratory as a future plan for clinical testing and synthesis against Dengue virus.

**Key words:** *In-silico*, Drug, Dengue, E-glycoprotein, Ligand, Molecular docking**Introduction**

Phytochemicals are the natural products derived from plants and have been recognized as an important, effective and beneficial alternative to manage various infections or diseases such as dengue (Lee *et al.*, 2013). Medicinal plants as herbal or natural products have been used as a source of treatment for a variety of diseases due to traditional healing (Okonkwo, 2012), complex bioactive substances and rich drug sources (Agnaniet *et al.*, 2016). Several plants have been recently listed for their ethno-pharmacological uses especially treating the most important diseases such as hepatitis, cancer, diabetes and dengue fever. Numerous compounds from medicinal plants such as *Kalanchoe gracilis*, *Glycyrrhiza uralensis*, *Curcuma longa*, *Azadirachta indica* with anti-viral activities have been recently reported (Wang *et al.*, 2016). Natural products are less toxic and cost effective in comparison to synthetic drugs (Babar *et al.*, 2013). Current advances in computational biology methods have expanded the possibility of working in the field of drug development. It is now believed that molecular docking is the main method for predicting the predominant binding mode of a ligand with a defined 3D protein structure (Kiat *et al.*, 2006) in drug design and screening of antiviral compounds discovered against scary diseases.

Dengue virus (DENV) is a single stranded RNA virus of family Flaviviridae, genus Flavivirus and is the main causative agent of Dengue (feverish disease). Flaviviruses are transmitted during a blood meal by mosquitoes bite (*Aedes albopictus* or *Aedes aegypti*). The four serotypes of dengue are DEN-1, DEN-2, DEN-3 and DEN-4, which cause dengue hemorrhagic fever (DHF) or dengue shock syndrome (DSS). The dengue fever is prevalent mainly in

the tropical and subtropical regions and more than 100 countries are affected with an estimation of 390 million infections worldwide per annum, of which 96 million were clinically diagnosed (Bhatt *et al.*, 2013). The developing of serious dengue infection is mostly observed among individuals, who are infected by a different type of DENV from first time infection. This is called second or heterotypic infection. For this reason, an acute disease first arises amongst the persons in zones wherever more than one serotype travels simultaneously. Infection with Dengue virus provides long-life immunity against the disease resulting from that particular type (reinfection), which promotes the feasibility of developing a vaccine, and shows effective results against Dengue. However, the infection only provides short-term immunity to the other three types of Dengue virus (Mizumoto *et al.*, 2014).

According to World Health Organization (WHO), DENV is single stranded RNA virus causing Dengue, a mosquito-borne disease that affects both children and adults. Symptoms may appear three to fourteen days after infective mosquito bite. It is not directly transmitted to humans and requires a vector (mosquito) for its transmission. The symptoms are mild to high fever, acute headache, pain in the back of eyes, severe joint pain, muscle pain and heavy rashes. There is currently no WHO recommendation for the use of any specific drug for dengue fever or any effective treatment. Persons with dengue are advised to get plenty of rest, consume plenty of fluids, and relieve fever using paracetamol under medical advice (Anon., 2016). It is often needed to combat dengue infection in safe, low cost and effective drugs with inhibitory activity against DENV, particularly in the most affected countries with limited resources (Rothan *et al.*, 2014).

*In-silico* drug designing is a complete and useful technique to design new drugs using information about biochemical activities of biological processes, chemical compounds. In the current era, the bioinformatics tools and softwares are greatly facilitating the discovery of *in-silico* drugs. In the current study, anti-viral plants were selected for screening potential anti-dengue compounds using *in-silico* approaches. The envelop protein (E-glycoprotein) of DENV2 was chosen as the drug target against dengue. The objective of the study was the *in-silico* investigation of phytochemicals exhibiting potential anti-viral activities against DENV infection.

## Materials and Methods

**Template selection and optimization:** The experimentally designed 3D (crystal) structure of DANV-2 E-glycoprotein (PDB id: 4UT6) was recovered from Protein Data Bank (PDB) for current study. The structural E-glycoprotein was selected as target due to the presence of immune-dominance of the fusion loop epitope (FLE), a segment of the E-glycoprotein buried at the interface of the E dimers coating mature viral particles making it more virulent. After the target is defined; the candidate drug targets were chosen by the analysis of formerly acknowledged drugs with powerful anti-dengue effects or by designing new inhibitory compounds. The overall dengue drug designing pathway is represented in (Fig. 1). Initially, more than a hundred plants having anti-viral compounds were chosen. Subsequently, 40 potential plants with anti-viral phytochemicals were short-listed. Than all those phytochemical ligands that violate the Lipinski's rule of five were rejected and best phytochemicals were selected for further analysis.

**Screening of Anti-dengue inhibitors:** A number of E-glycoprotein inhibitors were identified and selected for *in-silico* drug designing and molecular docking studies. Six compounds were selected for docking analysis with the drug like properties against DANV. These six compounds were selected by assessing their efficacy and efficiency against viruses based on recent investigations on DANV. Selected drug inhibitors were retrieved by applying a series of filters such as FDA approved drug, toxicity class, structural organization, ADMET properties, control and drug screening. Drug scanning to control the satisfactory conditions of the compound as a drug candidate was performed by using ADMETSAR (Cheng *et al.*, 2012) to ensure that the compounds retained the appropriate molecular properties, being a candidate drug (Table 1).

**Molecular docking:** The selected six anti-dengue agents were taken from PubChem (<http://www.ncbi.nlm.nih.gov/pccompound>) and ZINC databases (Irwin *et al.*, 2012). Ligands docking such as inhibitors and the recommended drugs with the E-glycoprotein receptor (4UT6) structure were performed by utilizing AutoDock (4.2) (Morries *et al.*, 2009). AutoDock gives the ligand structure and the binding interactions of the six chosen inhibitors along with the target active site of proteins. Addition of atoms of hydrogen, removal of water molecules were done before the molecular docking. For ligand preparation, after entering the ligand, the Gasteiger charges were combined with non-polar hydrogen ligands. Root detection was selected in the majority of atoms of the ligand and creating rotatable or non-rotatable bonds. Using the autogrid and 0.547 Å spacing, the 90x90x90 grid box was utilized to cover the whole molecule (receptor).

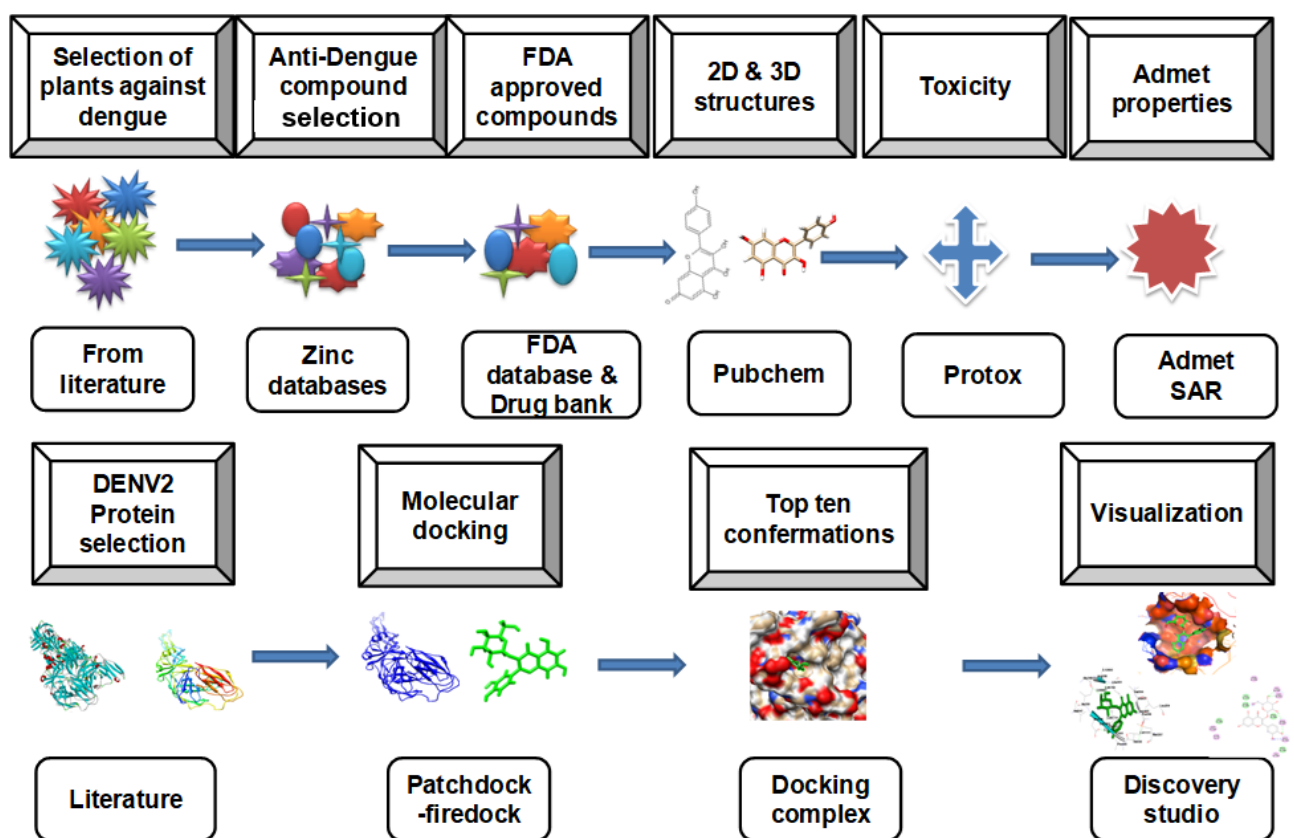


Fig. 1. Schematic representation of methodology used in current study for drug designing against DENV.

Table 1. Admet properties of the anti-dengue compounds.

| Properties        | ADMET properties              | Hyperoside | Cissampeloflavone | Ethylene glycol | Eugenol | Phenol  | Apigenin |
|-------------------|-------------------------------|------------|-------------------|-----------------|---------|---------|----------|
| <b>Absorption</b> | Blood-brain barrier           | BBB+       | BBB+              | BBB+            | BBB+    | BBB+    | BBB+     |
|                   | Human intestinal absorption   | HIA+       | HIA+              | HIA+            | HIA+    | HIA+    | HIA+     |
|                   | Aqueous solubility (LogS)     | -3.2198    | -3.5343           | 1.2264          | -3.7543 | -0.1541 | -2.7765  |
|                   | Caco-2 permeability (LogPapp) | Caco-2+    | Caco-2+           | Caco-2+         | Caco-2+ | Caco-2+ | Caco-2+  |
| <b>Metabolism</b> | P-glycoprotein inhibitor      | 0.9552     | 0.7135            | 0.9503          | 0.7379  | 0.9790  | 0.9543   |
|                   |                               | 0.9352     | 0.7968            | 0.9538          | 0.5145  | 0.9917  | 0.7525   |
|                   | CYP450 1A2 inhibitor          | 0.8896     | 0.9501            | 0.8432          | 0.9644  | 0.6114  | 0.9222   |
|                   | CYP450 2C9 inhibitor          | 0.8024     | 0.8448            | 0.9267          | 0.5201  | 0.9654  | 0.7746   |
|                   | CYP450 2D6 inhibitor          | 0.5844     | 0.9302            | 0.9729          | 0.9372  | 0.9746  | 0.9231   |
|                   | CYP450 2C19 inhibitor         | 0.6248     | 0.9606            | 0.9355          | 0.8993  | 0.8981  | 0.7043   |
|                   | CYP450 3A4 inhibitor          | 0.5153     | 0.8210            | 0.9657          | 0.6321  | 0.9523  | 0.9580   |
|                   | AMES toxicity                 | 0.6384     | 0.6571            | 0.9133          | 0.7310  | 0.9454  | 0.8906   |
| <b>Toxicity</b>   | Carcinogens                   | 0.6006     | 0.5690            | 0.9634          | 0.5396  | 0.8695  | 0.7316   |
|                   | Biodegradation                | BD+        | BD+               | BD+             | BD+     | BD+     | BD+      |
|                   | Acute oral toxicity           | NT         | NT                | NT              | NT      | NT      | NT       |
|                   | Carcinogenicity               | NC         | NC                | NC              | NC      | NC      | NC       |

Non-toxic; NC – Non-carcinogen

The docking parameter file for rigid docking is therefore made by identifying the molecule that is rigid after this specification of the ligand was done. Ligand-receptor docking was constructed using Patch Dock (Schneidman-Duhovny *et al.*, 2005). This method of docking was performed for six ligands having a receptor that has ten different conformations. These 10 different conformations were entered into Fire Dock for result refinement. Following, the molecular docking; protein-ligand complex was assessed to attain all the conformations. Out of these conformations, those with less binding energy value were chosen as the protein-ligand complex, which were recorded in PDB format. Once the molecular docking was completed, the protein-ligand complexes were closely analyzed using Discovery Studio (DS) software (Qamar *et al.*, 2014) and then ligand-receptor interactions were investigated. Ligand-receptor interactions containing hydrogen bonds, ionic bonds, and covalent bonds within a pocket were examined utilizing the Discovery Studio. This scheme is specified just for one ligand in molecular docking. For this reason, the docking of all the compounds to the protein molecule was done to capture each compound's complex and then examined in DS. Geometry optimization was done in Chimera 1.6.1 (<https://www.cgl.ucsf.edu/chimera/olddownload.html>). The whole methodology is summarized in Fig. 1.

## Results

**Molecular docking:** The docking process is initiated usually by binding of smaller molecule to a larger molecule like protein or enzymes. In the current study, initially above hundred plants with anti-viral compounds were chosen. Subsequently, 40 plants were short-listed with 200 anti-microbial phytochemicals, of which 6 most common anti-dengue compounds were selected. All those phytochemical ligands that violate Lipinski's rule of five were rejected. The six common anti-dengue phytochemicals were hyperoside, cissampeloflavone, ethylene glycol, eugenol, phenol and apigenin. Based on the binding of drugs (anti-dengue) to the E-glycoprotein (4UT6) (receptor protein), active sites were sought. Following this experiment, based on the x-score and lowermost binding energy, the best ligand-protein conformation was generated and selected to examine the

binding orientation and conformations within E-glycoprotein binding pocket. The amino acids contained by 5Å (i.e. active site) were acknowledged. According to this study the major factors of binding pockets were the residues GLY349, PHE337, GLU338 and LEU351. The active site and hydrophobic pocket of E-glycoprotein along with selected inhibitor molecules for all the six compounds is displayed in (Fig. 2). Protein-ligand interactions (docked complexes) showed that all bioactive compounds were fully buried within the binding pocket of E-glycoprotein (Fig. 3). The dockings were then performed utilizing the E-glycoprotein target protein in the test set compounds. After performing the docking of all the compounds with E-glycoprotein receptor protein, the Discovery Studio was used to visualize these docked complexes. The best docking complex was observed with hyperoside, while other compounds (cissampeloflavone, ethylene glycol, eugenol, phenol and apigenin) also showed efficient docking results. Then QSAR descriptors were used for the analysis of interaction of complexes such as hydrophobic interactions, ionic interaction and hydrogen bonds. The 2D structural analysis showed hydrophobic and ionic interactions of hyperoside with E-glycoprotein receptor protein.

**Lead identification:** Based on the binding interactions of six compounds, a "lead compound" was detected. The compounds hyperoside, cissampeloflavone, ethylene glycol, eugenol, phenol and apigenin displayed considerable interactions within the active site (5Å) of E-glycoprotein amino acids. The binding interaction of these showed that hyperoside was the lead compound forming five hydrogen bonds with the amino acids GLY349, PHE337, PHE367, GLU338 and LEU351 (Fig. 4) within the active site of E-glycoprotein with the distance of 2.96Å, 2.08Å, 2.36Å, 3.03Å, 3.06Å. Carbon bonds are formed by hyperoside with the E-glycoprotein receptor protein. The 3D crystal structure of E-glycoprotein showed residues close to the ligand molecule (Fig. 5). The medicinal activities of selected plants was identified, where majority of plants have shown the inhibitory activities against the DENV indicating potential control against dengue. Maximum activities were shown against Dengue, while anti-viral, anti-inflammatory, anti-cancer and antibacterial activities were also detected in these plants (Fig. 6).



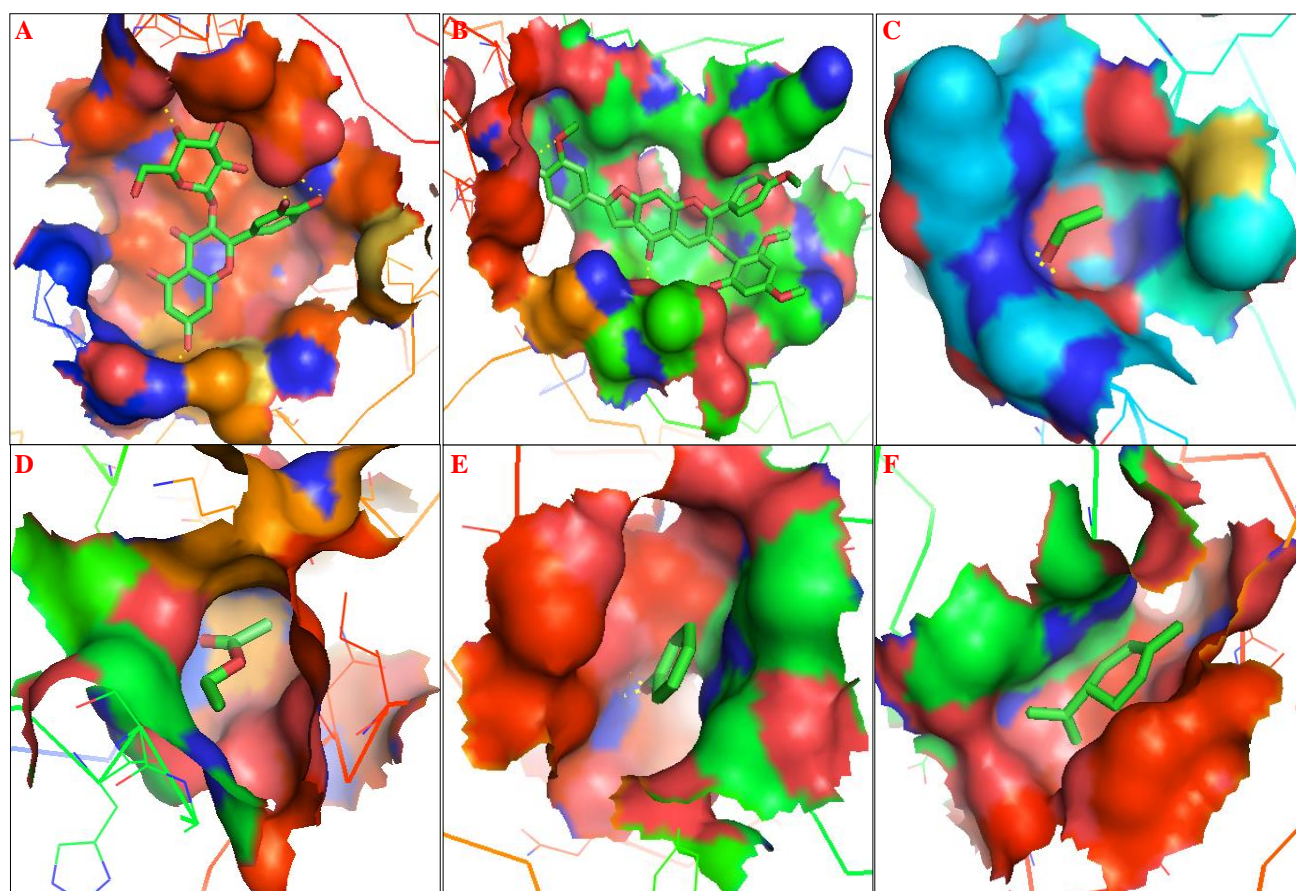


Fig. 2. Hydrophobic pocket of E-glycoprotein along with selected inhibitor molecules. Green structures represent small potential inhibitor molecules. A) hyperoside B) cissampeloflavone C) ethylene glycol D) eugenol E) phenol F) apigenin. The images of Figs. 2-3 were generated in Discovery Studio.

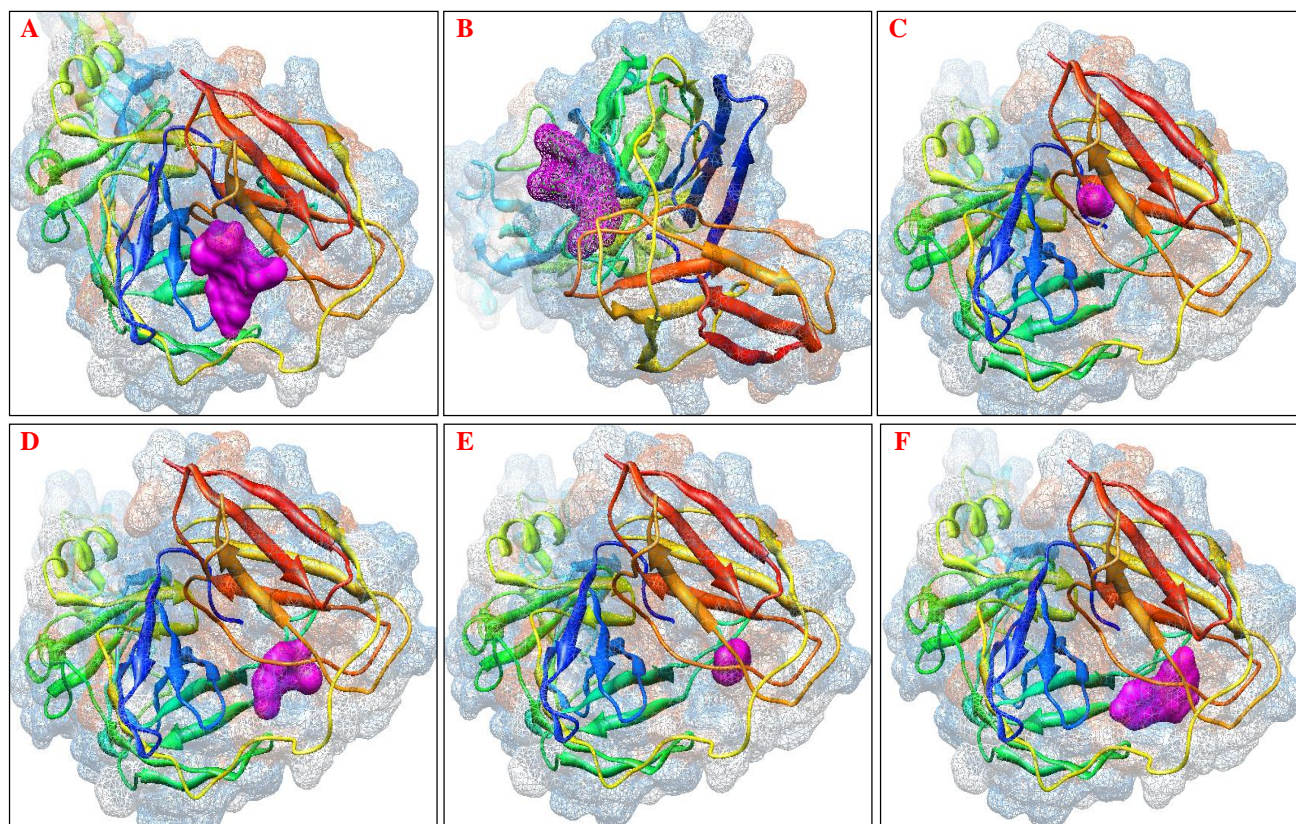


Fig. 3. Docked complexes of bioactive compounds with the binding pocket of E-glycoprotein; A) hyperoside B) cissampeloflavone C) ethylene glycol D) eugenol E) phenol F) apigenin.



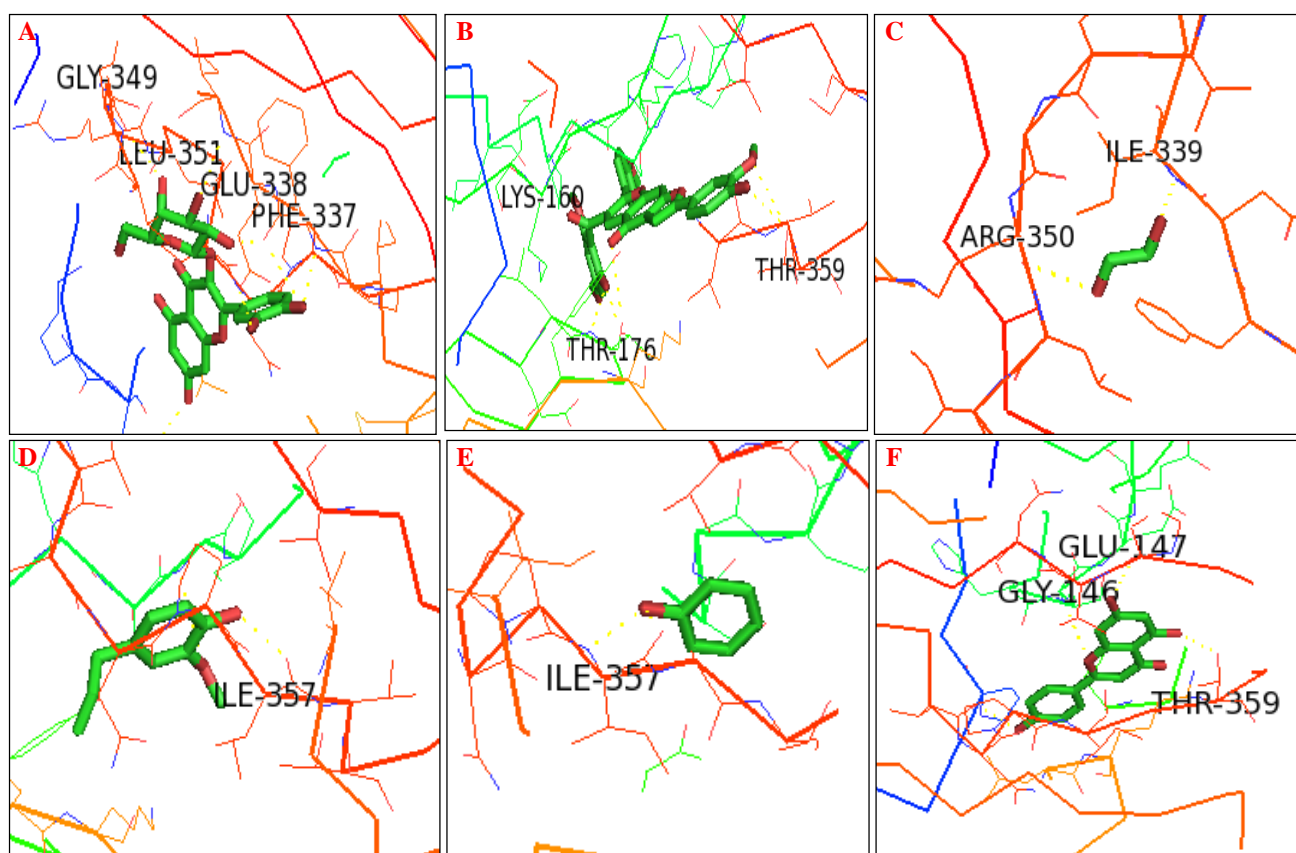


Fig. 4. Selected inhibitor molecules show hydrogen bonding with E-glycoprotein. A) hyperoside B) cissampeloflavone C) ethylene glycol D) eugenol E) phenol F) apigenin. The images of Figs. 4-5 were generated in Chimera 1.6.1 software.

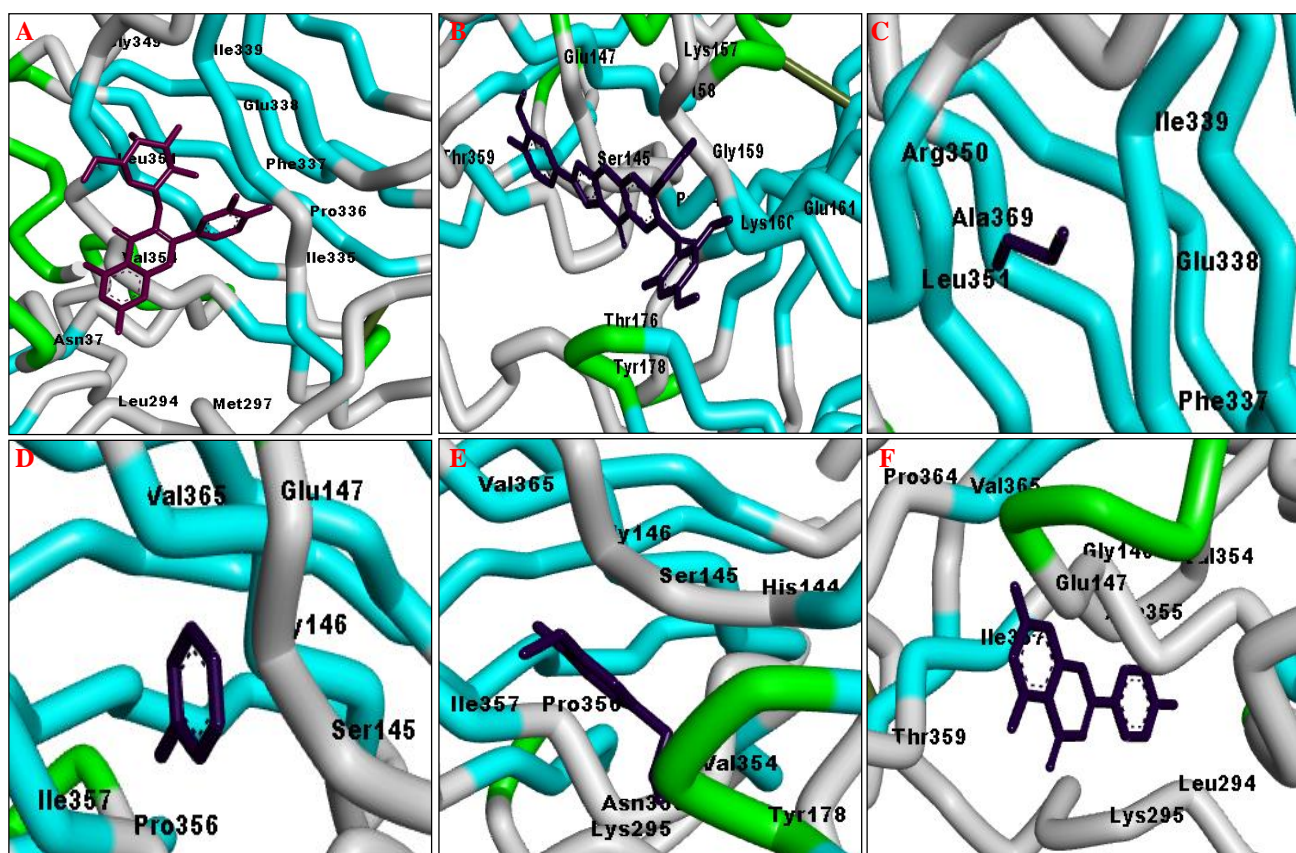


Fig. 5. The 3D crystal structure of E-glycoprotein shows residues close to the ligand molecule A) hyperoside B) cissampeloflavone C) ethylene glycol D) eugenol E) phenol F) apigenin.

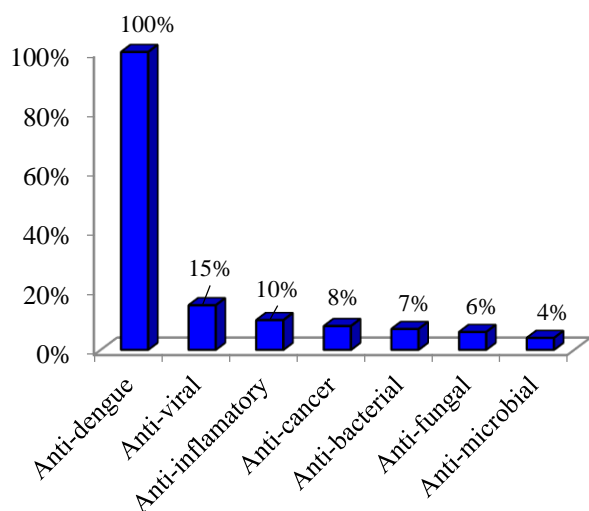


Fig. 6. Percentages of medicinal activities shown by selected plants.

From the six selected compounds, hyperoside is considered as a potential anti-dengue drug compound. The same method discussed earlier was utilized for docking of all the compounds with the E-glycoprotein active site. The selection of best conformation was based on the lowest binding energy of compounds in ten assessed conformations. Table 2 shows the QSAR descriptors of natural anti-dengue compounds and their energies. The conformation of each selected compound was analyzed after saving. For obtaining the interactions of the best conformation of the compound, PDB file of the protein was opened in the Discovery Studio. Hyperoside showed 5 hydrogen interactions and also have shown hydrophobic and ionic interactions respectively. However, cissampeloflavone showed 3 hydrogen interactions, whereas the ionic and hydrophobic interactions were also discovered. The ethylene glycol showed two hydrogen interactions as well as ionic and hydrophobic interactions. Single hydrogen interaction along with ionic and hydrophobic interactions were also shown by Eugenol. One hydrogen interaction was shown by phenol, where ionic and hydrophobic interactions were also displayed. It was found that 3 hydrogen interactions as well as ionic and hydrophobic interactions were investigated in Apigenin. Based on interactions with E-glycoprotein (target protein), maximum interactions with the target protein were shown by hyperoside. Although good interactions were shown by all the inhibitors but hyperoside showed the highest binding affinity. Five hydrogen interactions along with hydrophobic and ionic interactions were shown by hyperoside (lead compound) with the lowest binding pocket interactions' distance (3.5 Å). Hence, good interactions were shown by the hyperoside with our targeted protein presenting it a potential drug candidate against Dengue (Figs. 3-5).

**Establishing quantitative structure activity relationship (QSAR) and drug scan:** The data for six selected compounds was analyzed by QSAR to check the compounds having an exceptional symbol of attainment. The current QSAR study was conducted to identify and check the suitability of specified compound in the binding pattern of the selected target. Binding energy (kcal/mol), LogP and

molar refractivity (MR), toxicity class, RMSD values of the currently selected compounds were calculated using hydration energy and analyzed with the ArgusLab4.0.1 program on Chem Draw. The server Binding Affinity Prediction of Protein Ligand (BAPPL) aids in the estimation of binding energy of protein ligand complexes presented in Table 2. In order to confirm the requirement of the compound as a better drug candidate, an online cheminformatics tool (admetSAR) was used to perform the drug scan. In the end, in order to assess the likeliness of the drug compound molecular profile, Lipinski's rule was utilized (Lipinski, 2004), which states that the molecular weight of phytochemical drugs must be <500, LogP (Compound's Lipophilicity) <5, hydrogen donor numbers <5, and hydrogen acceptor numbers <10. Molecular properties were shown by this rule which are important for the understanding the drug's pharmacokinetics in the body of a human, along with their metabolism, distribution, absorption and excretion. Investigation showed that all selected six phytochemical drug compounds passed the essential filters. Table 2 illustrates the compounds inquiries for drug suitability of six compounds such as molecular formula, exact mass, polar surface area, hydrogen acceptor/donor, toxicity class, RMSD, boiling/melting points, Gibbs energy, LogP value and binding energies. After detailed analyses, a list of plants were compiled with the important anti-dengue compounds and their efficiency against dengue was checked using various bioinformatics approaches (Table 3). The list represents medicinal plants used in current study with their common names, family, medicinal uses, part used and phytochemical compounds with anti-dengue activities. The detailed investigations confirmed that most of the plants from various families have shown the availability of anti-dengue compounds like hyperoside, cissampeloflavone, ethylene glycol, eugenol, phenol and apigenin. The important plants with anti-dengue properties were *Andrographis paniculata*, *Azidarachta indica*, *Boerhaavia diffusa*, *Boesenbergia rotunda*, *Carica papaya*, *Chondrus crispus*, *Curcuma longa*, *Mimosa scabrella*, *Momordica charantia*, *Syzygium aromaticum* and *Quercus lusitanica*. Different plants parts are used for these herbal compounds such as leaves, stem, roots, fruits, seeds, bark or whole plant. The ethno-pharmacological uses of the listed plants have shown efficient activities such as antioxidant, anti-inflammatory, anticancer, anti-hepatitis, antibacterial and antiviral (anti-dengue) properties (Table 3; Fig. 6).

## Discussion

Dengue infection recurs as a serious threat to life, with an annual increase in infection rates. As a result, there is a need to create effective compounds (anti-dengue) to combat this widespread illnesses (Nair *et al.*, 2010). A novel perspective model molecule emerges as a call for the pharmaceutical industry because drug candidates cannot be produced as a consequence of the lack of a natural model. For this reason, it is vital to obtain an understanding of plants, since innate sources are required for model molecule isolation (Drwal *et al.*, 2014). Previous investigations have suggested quercetin as a major inhibitor since they exhibit better ligand enzyme stability and interactions, and DENV-2 exhibits significant anti-viral activity in vero cells inversely (Senthilvel *et al.*, 2013).



Table 3. Medicinal plants and their phytochemical compounds with anti-dengue activities.

| No. | Botanical name                     | Common names                     | Family names   | Ethno-pharmacological uses  | Parts used              | Anti-Dengue compounds |
|-----|------------------------------------|----------------------------------|----------------|---|-------------------------|-----------------------|
| 1.  | <i>Andrographis paniculata</i>     | Creat, Kariyat, Indian Echinacea | Acanthaceae    | It has a strong inhibitory effect against Dengue virus and increases the platelet level   | Whole plant, leaves     | Hyperoside            |
| 2.  | <i>Alternanthera philoxeroides</i> | Alligatorweed                    | Amaranthaceae  | It has shown a larvicidal effect against <i>Aedes aegypti</i> mosquitoes, also used in tuberculosis, measles and encephalitis B treatment   | Leaves                  | Cissampeloflavone     |
| 3.  | <i>Boesenbergia rotunda</i>        | Ginger, Finger root              | Zingiberaceae  | It has antioxidant, anti-inflammatory, antibacterial, antiviral, anticancer properties and shown inhibitory activity on the replication of Dengue virus   | Leaves                  | Ethylene glycol       |
| 4.  | <i>Castanospermum australe</i>     | Black Bean                       | Fabaceae       | This plant has antiviral activity and used in the treatment of dengue fever, HI and AIDS  | Seeds                   | Eugenol               |
| 5.  | <i>Chondrus crispus</i>            | Irish moss, Red seaweeds         | Gigartiniaceae | The components found in this plant were more effective inhibitors of Dengue virus-2   | Leaves                  | Phenol                |
| 6.  | <i>Cissampelos pareira</i>         | Velvetleaf                       | Menispermaceae | Methanolic extract of this plant have shown antiviral activities against dengue   | Leaves                  | Apigenin              |
| 7.  | <i>Gastrodia elata</i>             | Gastrodia                        | Orchidaceae    | The extract of this plant showed inhibitory activity on Dengue virus  | Roots                   | Hyperoside            |
| 8.  | <i>Euphorbia hirta</i>             | Asthma weed                      | Euphorbiaceae  | It has anti-bacterial, anti-viral and anti-fungal activities, treat bronchial asthma, laryngeal spasm, syphilis, and inhibits dengue  | Leaves                  | Cissampeloflavone     |
| 9.  | <i>Uncaria tomentosa</i>           | Catt's claw                      | Rubiaceae      | The hydro-alcoholic extract has anti-viral, anti-inflammatory, anti-microbial and anti-bacterial activities, used in the treatment of DANV-2, also used to treat wounds, treat stomach problems and cancer                                  | Vine bark, root         | Hyperoside            |
| 10. | <i>Kaempferia parviflora</i>       | Black ginger                     | Zingiberaceae  | It has anti-viral, anti-inflammatory and antioxidant activities   | Leaves and stem         | Hyperoside            |
| 11. | <i>Rhizophora apiculata</i>        | Mangrove                         | Rhizophoraceae | Petroleum ether extract has larvicidal activity against <i>A. aegypti</i> mosquito, shown inhibitory activities against flatulence; epilepsy, small pox, asthma, diabetes, rheumatism, stomach aches, fever, malaria, cholera and hepatitis | Leaves and stem         | Cissampeloflavone     |
| 12. | <i>Mimosa scabrella</i>            | Mimosa, Bracatinga               | Fabaceae       | The Galactomianians extracted from <i>Mimosa scabrella</i> seeds showed activities against Dengue virus-1 <i>In vitro</i> and <i>In vivo</i>  | Seeds                   | Cissampeloflavone     |
| 13. | <i>Leucaena leucocephala</i>       | White lead tree                  | Fabaceae       | This plant has antiviral activity against yellow fever and dengue   | Seeds                   | Eugenol               |
| 14. | <i>Phyllanthus urinaria</i>        | Chamber bitter, Gripe weed       | Phyllanthaceae | It has shown anti-cancer activities by inducing apoptosis. Its methanolic extract show anti-dengue activities   | Leaves                  | Ethylene glycol       |
| 15. | <i>Phyllanthus watsonii</i>        | -                                | Phyllanthaceae | The methanolic extract of this plant has anti-dengue activity   | Leaves                  | Ethylene glycol       |
| 16. | <i>Cladosiphon okamuranus</i>      | Brown seaweed                    | Chordariaceae  | Fucoidan extract showed strong anti-dengue, antitumor activities and is a potential therapeutic agent for patients with Adult T-cell leukemia (ATL)   | Leaves                  | Cissampeloflavone     |
| 17. | <i>Meristella gelidium</i>         | Tengusa, Makusa                  | Solieriaceae   | The extract and the fraction were more effective inhibitors of Dengue Virus   | Leaves                  | Apigenin              |
| 18. | <i>Oldenlandia affinis</i>         | Small-egg plant                  | Rubiaceae      | Showed inhibitory activities against dengue viral NS2B and NS3 protease   | Leaves, stems and roots | Phenol                |
| 19. | <i>Quercus infectori</i>           | Magic Nut, Masikai               | Fagaceae       | Methanol extracts of <i>Quercus infectori</i> have strongest inhibitory activity against Dengue Virus 2   | Leaves                  | Apigenin              |
| 20. | <i>Piper longum</i>                | Papal                            | Piperaceae     | The plant has shown control activities against <i>A. aegypti</i>  | Fruits                  | Hyperoside            |



Table 3. (Cont'd.).

| No. | Botanical name                 | Common names                         | Family names   | Ethno-pharmacological uses  | Parts used                     | Anti-Dengue compounds |
|-----|--------------------------------|--------------------------------------|----------------|---|--------------------------------|-----------------------|
| 21. | <i>Pimpinella anisum</i>       | Anisian, Aniseed                     | Apiaceae       | Showed insecticidal action against <i>A. aegypti</i> larvae   | Whole plant                    | Phenol                |
| 22. | <i>Curcuma longa</i>           | Turmeric                             | Zingiberaceae  | Ethyl acetate extract of <i>Curcuma longa</i> plant possess activity in inhibiting dengue fever   | Stem                           | Eugenol               |
| 23. | <i>Murraya koenigii</i>        | Curry leaf                           | Rutaceae       | Curry leaves possess anti-diabetic, antioxidant, hepatoprotective properties, strengthen the bones, improve digestion, and strengthen hair roots            | Leaves                         | Hyperoside            |
| 24. | <i>Caesalpinia pulcherrima</i> | Peacock flower, Red bird of paradise | Fabaceae       | Leaves are used as repellent for <i>A. aegypti</i>  | Leaves                         | Eugenol               |
| 25. | <i>Momordica charantia</i>     | Bitter melon                         | Cucurbitaceae  | Methanolic extract of <i>M. charantia</i> plant have inhibitory effect on Dengue Virus-1  | Leaves, stem, fruit, and resin | Hyperoside            |
| 26. | <i>Murraya koenigii</i>        | Kari patah or Kariapat               | Rutaceae       | The acetone and petroleum ether extracts of leaves has larvicidal effect against <i>A. aegypti</i>  | Leaves                         | Cissampeloflavone     |
| 27. | <i>Piper sarmentosum</i>       | Papal or Pippli                      | Piperaceae     | The ethanolic extract of plant have shown larvicidal activities against <i>A. aegypti</i> mosquitoes  | Fruit, root and stem           | Hyperoside            |
| 28. | <i>Houttuynia cordata</i>      | Fish mint Lizard tail                | Saururaceae    | Ethanol extracts from <i>Houttuynia cordata</i> plant shows an anti-dengue effect with 35.99% inhibition against Dengue Virus-1                             | Leaves                         | Hyperoside            |
| 29. | <i>Caricacarpaya</i>           | Papita                               | Caricaceae     | Aqueous extract of leaves of <i>Carica papaya</i> plant exhibited potential activity against Dengue fever   | Leaves                         | Hyperoside            |
| 30. | <i>Hippophae rhamnoides</i>    | Sea-buckthorn                        | Elaeagnaceae   | The extracts of leaves showed anti-dengue activity against DENV-2   | Leaves                         | Cissampeloflavone     |
| 31. | <i>Piper ribesoides</i>        | Papal or Pippli                      | Piperaceae     | The ethanolic extract of <i>Piper ribesoides</i> plant showed larvicidal activity against <i>A. aegypti</i> mosquitoes                                      | Fruit, root and stem           | Ethylene glycol       |
| 32. | <i>Pemphis acidula</i>         | Mentigi                              | Lythraceae     | Crude leaf extracts has larvicidal, ovicidal and repellent properties against <i>A. aegypti</i>   | Leaves                         | Eugenol               |
| 33. | <i>Azadirachta indica</i>      | Neem                                 | Meliaceae      | The aqueous extract of neem leaves has inhibitory effect on the replication of Dengue virus-2   | Leaves                         | Phenol                |
| 34. | <i>Boerhaavia diffusa</i>      | Spreading Hogweed,                   | Nyctaginaceae  | The plant showed antiviral efficacy against phytopathogenic viruses   | Roots, leaves and seeds        | Apigenin              |
| 35. | <i>Tinospora cordifolia</i>    | Amrita, Gilu, Giluncha               | Menispermaceae | The plant extract is used for diabetes, allergic rhinitis, high cholesterol, cancer, rheumatoid arthritis, hepatitis, dengue fever, syphilis and gonorrhoea | Stem and leaves                | Ethylene glycol       |
| 36. | <i>Andropogon citratus</i>     | Citronella grass                     | Poaceae        | Showed effectiveness in repelling <i>A. aegypti</i>   | Seeds                          | Eugenol               |
| 37. | <i>Psidium guajava</i>         | Guava leaves                         | Myrtaceae      | The leaf extracts reduce the growth of DENV   | Leaves                         | Phenol                |
| 38. | <i>Ervatamia coronaria</i>     | Pinwheel Jasmine                     | Apocynaceae    | <i>Ervatamia coronaria</i> plant is utilized as a herbal remedy to protect the mosquito bite  | Leaves                         | Apigenin              |
| 39. | <i>Syzygium aromaticum</i>     | Laung, clove                         | Myrtaceae      | <i>Syzygium aromaticum</i> oil is used as insect repellents including <i>A. aegypti</i>   | Seeds                          | apigenin              |
| 40. | <i>Quercus lusitanica</i>      | MazuPhal                             | Fagaceae       | The methanolic extract of <i>Quercus lusitanica</i> have shown inhibitory effect on the replication of DANV2  | Leaves                         | Eugenol               |

DENV-2 has a large binding cavity, so there are a number of active sites available for docking. Six phytochemical ligands from our docking examination indicated magnificent docking interactions with energies to DENV-2 (Table 2). These phytochemical agents were used to target the active site of the DENV-2 E protein. Phytochemicals small molecules can inhibit viral proteins by binding to glycoprotein active site of the protein. The results of the molecular docking provided a 2-D bioactive complex structure that specifically interacted with conserved target site residues. Interestingly, it was observed that Catechin, Laurifolin, Cianidanol had the highest binding energy to NS4B in DENV-1,2,4; which is due to the formation of more hydrogen bonds with amino acid residues at the binding site of the receptor. On the bases of results it was observed that the compound, particularly Catechin had anti-dengue activity (Paul *et al.*, 2016). The non-competitive inhibitor doxycycline showed a lower binding energy -5.15 kcal/mol than the predicted competitive inhibitors meclufenamic acid and comparable inhibitory inhibitors -3.64 and -3.21 kcal/mol respectively, against the NS2B-NS3 protease from DENV-2. Structural analyses showed compound doxycycline interacting with Lys74; an important amino acid residue linked to a specific allosteric region and linked to one of the Asp75 catalytic triads. Compounds meclufenamic acid and rolitetracycline showed direct binding with two of the His51 and Ser135 catalytic triads predicted as competitive inhibitors (Othman *et al.*, 2017).

It has been determined that the two ligands are the most suitable lead compounds, 1) 7-methyl-guanosine-5'-triphosphate-5'-(2'-O-methyl)-Guanosine and 2) Boctylglucucide had binding energy of (-123.2 and -84.61) respectively, for NS5 protein and envelope protein (Raikar *et al.*, 2017). ADMET properties and molecular docking results revealed 9 phytochemicals with potential inhibitory properties against Dengue virus namely, Isosilybin, Silydianin, Derrisin, Silymarin, Mundulinol, Narlumicine, Flavobion, Oxysanguinarine and Isopomorphine and to assess their *In vitro* and *In vivo* work can be considered as additional. These phytochemicals exhibited binding affinity  $\geq -8$  kcal/mol against Dengue virus 4-NS4B. The highest binding energy was shown for Palmatine -103.076 kcal/mol, delphinidin chloride -109.187 kcal/mol, squalene -109.975 kcal/mol and marmin -91.84 kcal/mol, 98.74 kcal/mol, while d-limonene and allicin exhibit a minimum binding energy to the binding sites of Dengue protein (PDB id. 3L6P) (Badoni *et al.*, 2015).

The present research was conducted to retrieve new drug molecules that can work as anti-dengue inhibitors by utilizing structure-based drug design methodology. E-glycoprotein was selected as a target in order to find new drugs. Hydrophobic cavity of DENV-2 E protein (Figs. 3-5A-F) showed a potential target for small-molecule inhibitors (Powers & Setzer, 2016). Six phytochemical drug compounds were chosen as a ligand, which were found effective against dengue virus. Every ligand was analyzed along with the target molecule complexes that were formed. Their energy values and binding affinities to the target were also assessed. On the bases of outcomes of current study, a lead compound was recognized. Six selected compounds were docked to find the best inhibitor

against Dengue. All inhibitors interacted well with the target E-glycoprotein, but phytochemical compound hyperoside showed the best docking complex with the target receptor. For this reason, the best of all other compounds is hyperoside and could be recommended as a drug for investigations in future. Anti-dengue inhibitors versus QSAR descriptors have been established to find addictive tendencies. Based on various descriptors, dengue virus showed dependence on drug activity. Diverse electronic and steric descriptors have been utilized to assess the association activity of the drug to the structure. The current study involved identification of six phytochemicals compounds with excellent docking properties with E-glycoprotein active site of DENV envelope protein. The results of this study reflect prior docking studies that showed phytochemicals to be astounding docking ligands to dengue protein targets (Powers *et al.*, 2016). This type of method aids to design the new drug with higher efficiency and inhibitors to Dengue virus.

## Conclusion

The current study achieved the desired goal by using a structure-based drug designing that provides the best inhibitor to serve as a powerful agent against DANV. The prominence of this study is reflected in the best definition of the six compounds as balanced DANV inhibitors. The lead compound hyperoside has been described in order to improve the effectiveness of antidepressant drugs due to the virus resistance. Proposed study regarding the potential drug designing against DENV might give important and effective data for the continuous exploration, innovation and design of new potential anti-viral drugs against DANV. The current study proposed the further formulations of these compounds and clinical trials to check their efficacy *In vivo*.

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